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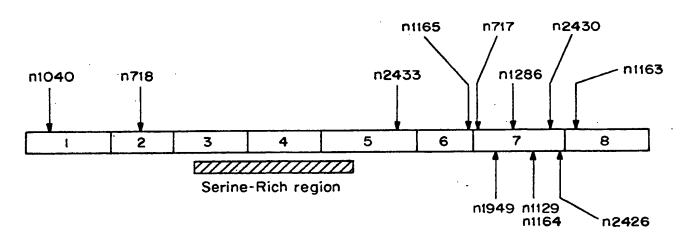
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(57) Abstract

Described herein is the discovery that human interleukin-1 β convertase (ICE) is structurally similar to the protein encoded by the *C. elegans* cell death gene, *ced-3*. Comparative and mutational analyses of the two proteins, together with previous observations, suggest that the Ced-3 protein may be a cysteine protease like ICE and that ICE may be a human equivalent of the nematode cell death gene. Another mammalian protein, the murine NEDD-2 protein, was also found to be similar to Ced-3. The NEDD-2 gene is implicated in the development of the murine central nervous system. On the basis of these findings, novel drugs for enhancing or inhibiting the activity of ICE, *ced-3*, or related genes are provided. Such drugs may be useful for treating inflammatory diseases and/or diseases characterized by cell deaths, as well as cancers, autoimmune disorders, infections, and hair growth and hair loss. Furthermore, such drugs may be useful for controlling pests, parasites and genetically engineered organisms. Furthermore, novel inhibitors of the activity of *ced-3*, ICE and related genes are described which comprise portions of the genes or their encoded products.

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INHIBITORS OF CED-3 AND RELATED PROTEINS

Background

Cell death is a fundamental aspect of animal development. Many cells die during the normal development of both vertebrates (Glucksmann, Biol. Rev. Cambridge Philos. Soc. 26:59-86 (1951)) and invertebrates (Truman, Ann. Rev. Neurosci. 7:171-188 (1984)). These deaths appear to function in morphogenesis, metamorphosis and tissue homeostasis, as well as in the generation of neuronal specificity and sexual dimorphism (reviewed by Ellis et al., Ann. Rev. Cell Biol. 7:663-698 (1991)). An understanding of the mechanisms that cause cells to die and that specify which cells are to live and which cells are to die is essential for an understanding of animal development.

The nematode Caenorhabditis elegans is an appropriate organism for analyzing naturally-occurring or programmed cell death (Horvitz et al., Neurosci. Comment. 1:56-65 (1982)). The generation of the 959 somatic cells of the adult C. elegans hermaphrodite is accompanied by the generation and subsequent deaths of an additional 131 cells (Sulston and Horvitz, Dev. Biol. 82:110-156 (1977); Sulston et al., Dev. Biol. 100:64-119 (1982)). The morphology of cells undergoing programmed cell death in C. elegans has been described at both the light and electron microscopic levels (Sulston and Horvitz, Dev. Biol. 82:100-156 (1977); Robertson and Thomson, J. Embryol. Exp. Morph. 67:89-100 (1982)).

Many genes that affect C. elegans programmed cell
death have been identified (reviewed by Ellis et al.,
Ann. Rev. Cell Biol. 7:663-698 (1991)). The activities
of two of these genes, ced-3 and ced-4, are required for
the onset of almost all C. elegans programmed cell
deaths (Ellis and Horvitz, Cell 44:817-829 (1986)).

when the activity of either ced-3 or ced-4 is eliminated, cells that would normally die instead survive and can differentiate into recognizable cell types and even function (Ellis and Horvitz, Cell 44:817-829 (1986); Avery and Horvitz, Cell 51:1071-1078 (1987); White et al., Phil. Trans. R. Soc. Lond. B. 331:263-271 (1991)). Genetic mosaic analyses have indicated that the ced-3 and ced-4 genes most likely act in a cell autonomous manner within dying cells, suggesting that the products of these genes are expressed within dying cells and either are cytotoxic molecules or control the activities of cytotoxic molecules (Yuan and Horvitz, Dev. Biol. 138:33-41 (1990)).

Summary of the Invention

This invention is based mainly on two experimental 15 findings and their implications: 1) that human interleukin-1 β convertase (ICE), which converts prointerleukin-1 β to the active cytokine and is involved in the inflammatory response in humans, has considerable 20 similarity to the protein encoded by the C. elegans cell death gene, ced-3; and 2) that fusion constructs containing amino-terminal portions of the ced-3 gene can prevent cell death in C. elegans. As discovered by Applicant, the human ICE and nematode Ced-3 proteins 25 have an overall amino acid identity of 28%. A higher degree of similarity was found in the carboxyl-terminal region, a region shown to be critical for the activities of both proteins. Furthermore, three sequences important for ICE activity, the region surrounding the 30 active cysteine and two autocleavage sites, have been shown to be conserved in the ced-3 gene product.

Thus, significant structural similarity has been shown between two proteins which previously were thought to be unrelated (to have dissimilar physiological

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roles). This finding leads to several implications, some of which are:

5

- 1) that the human ICE gene has an activity similar to that of ced-3 in causing cell death;
- 2) that the Ced-3 protein is also a cysteine protease with a substrate specificity similar to that of ICE;
- 3) that mutations in the ICE gene corresponding to mutations in the ced-3 gene will produce similar 10 effects, such as inactivation and constitutive activation:
- 4) that the ced-3 and ICE genes are members of a family of structurally related genes, referred to herein as the ced-3/ICE family, some of which are likely to be 15 cell death genes and some of which may encode substratespecific proteases;
- 5) that inhibitors of ICE, such as peptide aldehydes which contain the ICE recognition site or a substituted recognition site and the cowpox virus CrmA 20 protein, may also be useful for inhibiting cell deaths; and
 - 6) that inhibitors of ced-3, such as inhibitory portions of the gene or encoded product, may also be useful for inhibiting inflammation.
- This hitherto unknown connection between a cell death protein and a protease involved in the inflammatory response provides a basis for developing novel drugs and methods for the treatment of acute and chronic inflammatory disease, of leukemias in which IL-1β is implicated, and of diseases and conditions characterized by cell deaths (such as myocardial infarction, stroke, traumatic brain injury, viral and other types of pathogenic infection, neural and muscular degenerative diseases, aging, hair loss). In addition, drugs which increase cell deaths and which are useful

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for reducing the size or proliferative capacity of cell populations, such as cancerous cells, infected cells, cells which produce autoreactive antibodies, and hair follicle cells, as well as drugs which incapacitate or kill organisms, such as pests, parasites and recombinant organisms, can be developed using the ced-3, ICE, and other ced-3/ICE genes and their gene products.

This work also provides probes and methods for identifying additional members of the ced-3/ICE gene

10 family. Genes related to ced-3 and ICE are expected to exist in various organisms. Some of these may be cell death genes and/or proteases. The sequences of these related genes and their encoded products can be compared, for instance, using computer-based analysis,

15 to determine their similarities. Structural comparisons, for example, would indicate those regions or features of the genes or encoded products which are likely to be functionally similar and important. Such information can be used to design drugs which mimic or

20 alter the activity of the ced-3, ICE, or other ced-3/ICE genes, and which may, thus, be useful in the various medical and agricultural applications mentioned above.

In addition, another mammalian protein, the murine NEDD-2 protein, was also found to be similar to Ced-3.

25 Interestingly, NEDD-2 is not significantly similar to ICE. Thus, another potential mammalian cell death gene was identified.

Also described herein is the discovery that fusion constructs which encode an amino-terminal portion of the 30 Ced-3 protein fused to β-galactosidase act as inhibitors of cell death in C. elegans. Due to its structural similarity to Ced-3, constructs encoding corresponding portions of the human ICE protein are also expected to inhibit the enzymatic activity of ICE in cleaving 35 interleukin-1β. Thus, inhibitors comprising an amino-

terminal portion of the Ced-3 protein, ICE protein or another member of the Ced-3/ICE family and RNAs and DNA constructs which express these protein portions are potentially useful for decreasing cell deaths and/or inflammation involved in various pathologies. Methods for identifying other inhibitory portions of the ced-3 and ICE genes are also described.

Furthermore, deletion of the inhibitory aminoterminal portions of the ced-3 and ICE genes may result in constitutive activation of the genes. Constitutively activated carboxyl-terminal portions of the genes, or their encoded products, may thus be useful in applications where increased cell deaths or an increased inflammatory response are desired.

15 Brief Description of the Drawings

Figure 1 shows the physical and genetic maps of the ced-3 region on chromosome IV.

Figure 2 summarizes the experiments to localize ced-3 within C48D1. Restriction sites of plasmid C48D1

20 and subclone plasmids are shown. Ced-3 activity was scored as the number of cell corpses in the head of L1 young animals. ++, the number of cell corpses above 10.

+, the number of cell corpses below 10 but above 2. -, the number of cell corpses below 2.

Figures 3A-H show the nucleotide sequence (SEQ ID NO:1) of ced-3 and deduced amino acid sequence (SEQ ID NO:2). The genomic sequence of the ced-3 region was obtained from plasmid pJ107. The introns and the positions of 12 ced-3 mutations are indicated. The likely translation initiation site is indicated by a solid arrowhead. The SL1 splice acceptor of the RNA is boxed. Repetitive elements are indicated as arrows above the relevant sequences. Numbers on the sides

indicate nucleotide positions. Numbers under the amino acid sequence indicate codon positions.

Figure 4A shows the genomic structure of the ced-3 gene and the location of the mutations. The sizes of 5 the introns and exons are given in bp. The downward arrows indicate the positions of 12 EMS-induced mutations of ced-3. The arrow pointing right indicates the direction of transcription. The solid arrowhead indicates the translation initiation site. The open arrowhead indicates the termination codon.

Figure 4B shows the locations of the mutations relative to the exons (numbered 1-7) and the encoded serine-rich region in ced-3.

Figure 5 is a Kyte-Doolittle hydrophobicity plot of the Ced-3 protein.

Figures 6A-B show the alignment of the amino acid sequences of Ced-3 (SEQ ID NO:2) and human interleukin-1β convertase (ICE; SEQ ID NO:4). Vertical bars indicate identical amino acids and single and double dots indicate similar amino acids, where double dots signifies closer similarity than a single dot. The serine-rich region and inactivating mutations of Ced-3 are indicated. The active site and autocleavage sites of ICE are indicated. The portions of the Ced-3 protein encoded by the BGAFQ and PBA constructs are also shown.

Figure 7 shows the alignment of the amino acid sequences of Ced-3 (SEQ ID NO:2) and murine NEDD-2 (SEQ ID NO:13). Vertical bars and single and double dots signify degrees of similarity as in Figures 6A-B.

30 Inactivating mutations of Ced-3 are shown.

Figure 8A shows the alignment of the amino-terminal regions of the Ced-3 proteins of three nematode species (C. briggsae, C. elegans, and C. vulgaris) and mouse (SEQ ID NO:14) and human ICEs. A consensus sequence is also shown. Amino acid positions with the same residue

in more than half of the sequences are shaded. Completely conserved amino acids are also boxed.

Figure 8B shows the alignment of carboxyl-terminal regions of the three nematode Ced-3 proteins, human and 5 mouse ICEs, and the mouse NEDD-2 protein. Except for NEDD-2, these sequences are contiguous with the corresponding sequences shown in Figure 8A. A consensus sequence and amino acid conservation are also shown.

Figure 9 shows a comparison of the Ced-3 proteins

10 of C. elegans (line 1; SEQ ID NO:2) and two related

nematode species, C. briggsae (line 2; SEQ ID NO:5) and

C. vulgaris (line 3; SEQ ID NO:6). The conserved amino

acids are indicated by ".". Gaps inserted in the

sequence for the purpose of alignment are indicated by

15 "".

Figure 10 is the interleukin-1 β convertase cDNA sequence (SEQ ID NO:3).

Figure 11A is a schematic representation of two fusion constructs that can prevent programmed cell 20 death.

Figure 11B is a schematic representation of the lacz-containing portion of the fusion constructs.

Detailed Description of the Invention

This invention is based on the discovery that the

25 human enzyme interleukin-1\$\beta\$ convertase (ICE) has

significant structural similarity to the protein product

of the \$C\$. elegans cell death gene, \$ced-3\$. The

activities of \$ced-3\$ and another cell death gene, \$ced-4\$,

have been shown to be required for almost all the cell

30 deaths which occur during the development of the

nematode. ICE is a cysteine protease whose

physiological significance has been thought to be

related to its role in the maturation of one form of

interleukin-1 (IL-1), a major mediator of the immune and

inflammatory response (Fuhlbrigge et al., in: The Year in Immunology, Cruse and Lewis (eds.), Karger, Basel, 1989, pp. 21-37). There are two distantly related forms of IL-1, α and β , of which the β form is the predominant 5 species. ICE selectively converts pro-interleukin-1 β to the active cytokine, IL-1 β . The production of active IL-1 β has been implicated in acute and chronic inflammatory diseases, septic shock, and other physiological processes, including wound healing and 10 resistance to viral infection (Ray et al., Cell 69:597-604 (1992)). As a result of this discovery, an enzyme which has been known to be involved in the inflammatory response and inflammatory diseases is implicated as having a role in cell death processes. This discovery 15 is consistent with the notion that cell death genes equivalent to the nematode ced-3 gene function in a variety of organisms. The structural similarity between their gene products suggests that the ICE gene is a human equivalent of the ced-3 cell death gene. As 20 further described below, the conservation of certain features of ICE in the Ced-3 protein further suggests that Ced-3 is a protease with a substrate-specificity similar to that of ICE.

Furthermore, the identification of ced-3 and ICE as structurally related genes (i.e., genes whose encoded products, or which themselves, are structurally similar) presents the possibility that a family of structurally related genes exists and provides probes to identify additional members of this ced-3/ICE gene family.

30 Comparison of the genes within this family could indicate functionally important features of the genes or their gene products, and thus, provide information for designing drugs which are useful for treating conditions characterized by cell deaths and/or inflammatory

35 disease.

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This discovery provides novel drugs based on the ced-3, ICE and other ced-3/ICE genes and encoded products that inhibit the production of IL-1\beta and are useful for treatment (preventive and therapeutic) of 5 acute and chronic inflammatory disease, as well as drugs which reduce cell deaths and are useful for treatment of diseases and conditions involving cell deaths (such as myocardial infarction, stroke, traumatic brain injury, viral and other types of pathogenic infection,
10 degenerative diseases, aging, and hair loss). These drugs may also be useful for treating leukemias in which IL-1\beta has been implicated.

Drugs or agents which increase cell deaths can also be developed based on the ced-3, ICE, and related genes and gene products; such drugs or agents may be useful for killing or incapacitating undesired cell populations (such as cancerous cells, infected cells, cells which produce autoreactive antibodies and hair follicle cells) or undesired organisms (such as pests, parasites, and genetically engineered organisms). Drugs are also provided which increase IL-1\$\beta\$ production and, therefore, the inflammatory and immune response. These drugs may be helpful for providing increased resistance to viral and other types of infection.

Also described herein is the discovery that fusion constructs containing amino-terminal portions of the ced-3 gene can inhibit the activity of the intact gene when expressed in otherwise wild-type worms. Due to the similarity between ICE and Ced-3, it is likely that the corresponding amino-terminal portions of the ICE gene will also inhibit the enzymatic activity of ICE in cleaving interleukin-1β. Thus, novel inhibitors of the ced-3 and ICE genes are provided which may be useful for decreasing cell deaths and/or inflammation involved in various pathologies.

This work has also shown that Ced-3 and the murine NEDD-2 protein are structurally similar. Thus, drugs for increasing or decreasing cell deaths can be developed based on the NEDD-2 gene and its encoded 5 products.

The above-described discoveries, and their implications, and novel drugs and treatments for diseases related to cell death and/or inflammation arising therefrom are described in further detail below.

10 As used herein, the activity of a gene is intended to include the activity of the gene itself and of the encoded products of the gene. Thus, drugs and mutations which affect the activity of a gene include those which affect the expression as well as the function of the encoded RNA and protein. The drugs may interact with the gene or with the RNA or protein encoded by the gene, or may exert their effect more indirectly.

The ced-3 Gene

The C. elegans ced-3 gene was cloned by mapping DNA restriction fragment length polymorphisms (RFLPs) and chromosome walking (Example 1; Figure 1). The gene was localized to a 7.5 kb fragment of cloned genomic DNA by complementation of the ced-3 mutant phenotype (Figure 2). A 2.8 kb transcript was further identified. The ced-3 transcript was found to be most abundant in embryos, but was also detected in larvae and young adults, suggesting that ced-3 is expressed not only in cells undergoing programmed cell death.

A 2.5 kb cDNA corresponding to the ced-3 mRNA was sequenced. The genomic sequence cloned in the plasmid pJ107 was also determined (Figure 3; SEQ ID NO:1). A comparison with the cDNA sequence revealed that the ced-3 gene has 7 introns which range in size from 54 to 1195 bp (Figure 4A). The four largest introns, as well as

sequences 5' of the start codon, contain repetitive elements (Figure 3), some of which have been previously characterized in non-coding regions of other C. elegans genes such as fem-1 (Spence et al., Cell 60:981-990 (1990)), lin-12 (J. Yochem, personal communication), and myoD (Krause et al., Cell 63:907-919 (1990)). The transcriptional start site was also mapped (Figure 3), and a ced-3 transcript was found to be trans-spliced to a C. elegans splice leader, SL1.

10 Twelve EMS-induced ced-3 alleles were also sequenced. Eight of the mutations are missense mutations, three are nonsense mutations, and one is a putative splicing mutation (Table 1). This identification of ced-3 null alleles, together with 15 results of genetic analysis of nematodes homozygous for these null mutations in ced-3, indicate that, like ced-4, ced-3 function is not essential to viability. In addition, 10 out of the 12 mutations are clustered in the carboxyl-terminal region of the gene (exons 6-8, Figure 4B), suggesting that this portion of the encoded protein may be important for activity.

The ced-3 gene encodes a putative protein of 503 amino acids (Figure 3; SEQ ID NO:2). The protein is very hydrophilic and no significantly hydrophobic region can be found that might be a transmembrane domain (Figure 5). One region of the Ced-3 protein is very rich in serine (Figures 6A-B). Comparison of the C. elegans protein with the Ced-3 proteins of two related nematodes species, C. briggsae and C. vulgaris, shows conservation of the serine-rich feature without conservation of the amino acid sequence in this region (Figure 9; SEQ ID NO:5-6). This suggests that the exact sequence of this serine-rich region may not be important but that the serine-rich feature is. This hypothesis is supported by analysis of ced-3 mutations: none of 12

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EMS-induced ced-3 mutations is in the serine-rich region (Figure 4B). It is possible that the serine-rich region in Ced-3 is another example of semi-specific protein-protein interaction, similar to acid blobs in transcription factors and basic residues in nuclear localization signals. In all these cases, the exact primary sequence is not important.

The serine-rich region may function as a site for post-translational regulation of Ced-3 activity through 10 protein phosphorylation of the serine residues by a Ser/Thr kinase. McConkey et al. (J. Immunol, 145:1227-1230 (1990)) have shown that phorbol esters, which stimulate protein kinase C, can block the death of cultured thymocytes induced by exposure to Ca++ 15 ionophores or glucocorticoids (Wyllie, Nature 284:555-556 (1980); Wyllie et al., J. Path. 142:67-77 (1984)). It is possible that protein kinase C may inactivate certain cell death proteins by phosphorylation and, thus, inhibit cell death and promote cell proliferation. 20 Several agents that can elevate cytosolic cAMP levels have been shown to induce thymocyte death, suggesting that protein kinase A may also play a role in mediating thymocyte death. Further evidence suggests that abnormal phosphorylation may play a role in the 25 pathogenesis of certain cell-degenerative diseases. For example, abnormal phosphorylation of the microtubuleassociated protein Tau is found in the brains of Alzheimer's disease and Down's syndrome patients (Grundke-Igbal et al., Proc. Natl. Acad. Sci. USA 30 83:4913-4917 (1986); Flament et al., Brain Res. 516:15-19 (1990)). Thus, it is possible that phosphorylation may have a role in regulating programmed cell death in C. elegans. This is consistent with the fairly high levels of ced-3 and ced-4 transcripts which suggest that transcriptional regulation alone may be insufficient to regulate programmed cell death.

Structural Relatedness of the ced-3 and Human Interleukin-1\beta Convertase Genes and Functional

5 Implications

A search of GenBank, PIR and SWISS-PROT databases using the Blast program (National Center for Biotechnology Information) revealed that human interleukin-1β convertase (ICE) has a 28% amino acid identity with the Ced-3 protein (Figures 6A-B). A comparable level of overall similarity was found between ICE and the Ced-3 proteins from two other nematode species, C. briggsae and C. vulgaris.

The carboxyl-terminal regions of Ced-3 and ICE 15 (amino acids 250-503 and amino acids 166-404, respectively) were found to be more conserved (33% identity) than the amino-terminal portions of the two proteins (22% identity). A comparison of human and murine ICEs also indicated a high degree of similarity 20 (80% identity) in the carboxyl-terminal region compared with an overall identity of 62% (Cerretti et al., Science 256:97-100 (1992)). Furthermore, deletion analysis of the ICE cDNA sequence has shown that the amino-terminal 119 amino acids of ICE are not required 25 for enzymatic activity, but that deletions of the carboxyl-terminal region eliminate the enzyme's ability to process pro-IL-1 β (Cerretti et al., 1992 supra). observation that most of the inactivating mutations of ced-3 cluster in the carboxyl-terminal region (Figure 30 4B) suggests that the activity of Ced-3 also resides (at least partially) in this region. Thus, the identification of the carboxyl-terminal regions of the two proteins as functional domains and the marked similarity of these regions suggest that the Ced-3 and

ICE proteins have similar activities, i.e., that ICE has cell death activity similar to Ced-3 and Ced-3 has protease activity similar to ICE.

The possiblity that Ced-3 has protease activity is 5 further supported by the observation that the region surrounding the active cysteine and two autocleavage sites of ICE appear to be conserved in the Ced-3 protein. As shown in Figures 6A-B, the five amino acids (QACRG, amino acids 283 to 287) surrounding the active 10 cysteine of ICE (Thornberry et al., Nature 356:768-774 (1992)) are conserved in amino acids 356 to 360 of Ced-3; this pentapeptide is the longest conserved sequence between ICE and Ced-3. This peptide is also conserved in the Ced-3 proteins of C. briggsae and C. vulgaris 15 (Figure 9). One inactivating mutation of ced-3, n2433, introduces a glycine to serine change near the putative active cysteine (Figures 6A-B). The human ICE gene encodes a precursor enzyme which is autoproteolytically cleaved at two major sites (amino acids 103 and 297) by 20 the active form of the enzyme (Thornberry et al., 1992 supra). The Asp-Ser dipeptides of both autocleavage sites are conserved in Ced-3 (at amino acids 131 and 371) (Figures 6A-B). The conservation of these functionally important sequences strongly suggests that, 25 like ICE, Ced-3 is a cysteine protease with a similar substrate-specificity. Ced-3 would, therefore, be expected to cleave the IL-1 β precursor, as well as other substrates of ICE.

The possibility that ICE is a cell death gene is consistent with evidence which suggests that the production of active IL-1β is involved with cell death processes. Firstly, a variety of studies has suggested that IL-1β can prevent cell death (McConkey et al., J. Biol. Chem. 265:3009-3011 (1990); Mangan et al., J.

35 Immun. 146:1541-1546 (1991); Sakai et al., J. Exp. Med.

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166:1597-1602 (1987); Cozzolino et al., Proc. Natl.
Acad. Sci. USA 86:2369-2373 (1989)). Secondly, active,
mature IL-1β appears to be released from cells
undergoing cell death. Studies on murine macrophages
5 suggest that release of the active form seems not to be
merely due to the lysis of the cells or leaking of cell
contents. When murine peritoneal macrophages were
stimulated with lipopolysaccharide (LPS) and induced to
undergo cell death by exposure to extracellular ATP,
10 mature active IL-1β was released into the culture
supernatant. In contrast, when the cells were injured
by scraping, IL-1β was released exclusively as the
inactive proform (Hogquist et al., Proc. Natl. Acad.
Sci. USA 88:8485-8489 (1991)).

The similarity between ICE and Ced-3 strongly supports the hypothesis that ICE is involved in cell death. Since Ced-3 is necessary for cell death, one suggestion is that ICE is also necessary for cell death. It is possible that IL-1 β can cause cell death.

20 Alternatively, ICE could produce products besides IL-1 β , one or more of which can cause cell death. The observation that the ICE transcript is detected in cells that lack IL-1 β expression (Cerretti et al., 1992 supra) supports this idea.

The finding of a human gene related to the nematode ced-3 gene is consistent with the idea that cell death genes which are structurally related and/or functionally similar to the nematode ced-3 gene exist in a variety of organisms. This idea is supported by evidence that cell deaths occurring in a variety of organisms, including vertebrates and invertebrates, and possibly microbes and plants, as well as cell deaths observed in various developmental and pathologic situations share a common genetic mechanism. Evidence for this hypothesis is discussed in Example 2. The structural relatedness of

ICE suggests that it is a mammalian equivalent of the nematode cell death gene, ced-3. The cDNA sequence of ICE is shown in Figure 10 (SEQ ID NO:3).

The ced-3/ICE Gene Family and Uses Thereof

The ICE and ced-3 genes can be used to isolate additional structurally related genes, including genes from other organisms. Such genes may be identified using probes derived from both the ced-3 and ICE gene sequences and known techniques, including nucleic acid 10 hybridization, polymerase chain reaction amplification of DNA, screening of cDNA or genomic libraries, and antibody screening of expression libraries. The probes can be all or portions of the genes which are specific to the genes, RNA encoded by the genes, degenerate 15 oligonucleotides derived from the sequences of the encoded proteins, and antibodies directed against the encoded proteins. The sequences of the genes and their protein products can also be used to screen DNA and protein databases for structurally similar genes or 20 proteins.

One strategy for detecting structurally related genes in a number of organisms is to initially probe animals which are taxonomically closely related to the source of the probes, for example, probing other worms

25 with a ced-3-derived probe, or probing other mammals with an ICE-derived probe. Closely related species are more likely to possess related genes or gene products which are detected with the probe than more distantly related organisms. Sequences conserved between ced-3 or ICE and these new genes can then be used to identify similar genes from less closely related species. Furthermore, these new genes provide additional sequences with which to probe the molecules of other animals, some of which may share conserved regions with

the new genes or gene products but not with the original probe. This strategy of using structurally related genes in taxonomically closer organisms as stepping stones to genes in more distantly related organisms can be referred to as walking along the taxonomic tree.

Together, ced-3, ICE, and related genes obtained as described above would comprise a family of structurally related genes, referred to herein as the ced-3/ICE gene family. It is highly likely that at least some of these 10 additional family members would exhibit cell death and/or protease activity. The new genes can be tested for protease activity using known assay methods. For example, the sequence of the protein encoded by a new gene may indicate an active site and substrate-15 specificity similar to that of ICE, such as observed in Ced-3. This activity can then be verified using the transient expression assays and purified enzyme assays previously described (Cerretti et al., Science 256:97-100 (1992); Thornberry et al., Nature 356:768-774 20 (1992)). Cell death activity can be tested in bioassays using transgenic nematodes. A candidate cell death gene, such as the ICE gene, can be injected into Ced-3deficient mutant animals to determine whether the gene

The ced-3, ICE and other related genes which have cell death activity can be used to develop and identify drugs which reduce or increase cell deaths. Drugs which reduce cell deaths are potentially useful for treating diseases and conditions characterized by cell deaths, such as myocardial infarction, stroke, viral and other pathogenic infections (e.g., human immunodeficiency virus), traumatic brain injury, neural and muscular degenerative diseases, and aging. Drugs which cause cell deaths can be used to control or reduce undesired

complements the *ced-3* mutation. Expression libraries 25 can also be screened for cell death genes by this assay.

cell populations, such as neoplastic growths and other cancerous cells, infected cells, and cells which produce autoreactive antibodies. Undesired organisms, such as pests, parasites, and recombinant organisms, may also be incapacitated or killed by such drugs.

ICE has been implicated in the growth of certain leukemias (Sakai et al., J. Exp. Med. 166:1597 (1987); Cozzolino et al., Proc. Natl. Acad. Sci. U.S.A. 86:2369 (1989); Estrov et al., Blood 78:1476 (1991); Bradbury et al., Leukemia 4:44 (1990); Delwel et al., Blood 74:586 (1989); Rambaldi et al., Blood 78:3248 (1991)). The observation that the human ICE gene maps to chromosome location 11q23, a site frequently involved in DNA rearrangements seen in human cancers (C. Cerretti et al., Science 256: 97-100 (1992)), further suggests that ICE is involved in cancer. The finding that ICE probably functions in cell death implies that ICE and other related genes, like ced-3, may be used to develop drugs to control cancerous growth.

In addition, since cell death plays an important role in mammalian hair growth, it seems likely that by controlling cell death, one could cause or prevent hair loss. It has been found that bcl-2, a human gene which is structurally related to the gene which prevents cell deaths in nematode development (ced-9), is expressed in the hair follicle in a cell-cycle dependent manner. ced-9 has been shown to act by antagonizing the activities of the cell death genes, ced-3 and ced-4. Together, these findings suggest that genes equivalent to the ced-3, ced-4, and ced-9 genes are involved in the physiology of mammalian hair growth and loss.

Drugs which increase cell deaths may comprise ced-3, ICE, and other ced-3/ICE family members, their RNA and protein products, constitutively activated mutants of the genes and encoded products, and peptide

and non-peptide mimetics of the proteins. Drugs which decrease cell deaths may comprise antisense RNA complementary to the mRNA of a cell death gene, or mutant cell death genes or encoded products, that no 5 longer cause cell death and interfere with the function of wild-type genes. Furthermore, drugs comprising agonists and antagonists of the cell death genes can be designed or identified using the genes or their gene products as targets in bioassays. The bioassays can be 10 conducted in wild-type, mutant, or transgenic nematodes, in which an alteration in programmed cell deaths is an indicator of an effective agonist or antagonist. Bioassays can also be performed in cultured cells transfected with the target cell death gene, into which 15 the substance being tested is introduced. In bioassays for antagonists of cell death, the cultured cells should be put under conditions which induce the activity of the target cell death gene.

Uses of bioassays utilizing *C. elegans* are 20 exemplified by the following:

- 1) use of normal, wild-type nematodes to screen for drugs or genes that inactivate ced-3 and hence, prevent programmed cell deaths;
- 2) use of normal, wild-type nematodes to screen 25 for drugs or genes that activate ced-3 and hence, cause excess cell deaths;
- 3) use of mutant nematodes which overexpress ced-3 or which express a constitutively activated ced-3 gene to identify drugs or genes that prevent excess cell 30 deaths caused by the ced-3 mutation;
 - 4) use of mutant nematodes which underexpress ced-3 or which express an inactivated ced-3 gene to identify drugs or genes that mimic or complement the ced-3 mutation;

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- 5) use of transgenic nematodes (with an inactivated endogenous ced-3 gene) in which either a wild-type or mutant form of ICE or other ced-3/ICE family member causes excess cell deaths to identify drugs or genes which antagonize the activity of the transgene; and
- 6) use of transgenic nematodes which carry a transgene that inhibits cell death (e.g., a transgene that expresses an inhibitory fragment of ced-3, ICE or 10 related gene, as described below) to identify drugs that overcome this inhibition and cause cell death.

Drugs can be introduced into nematodes by diffusion, ingestion, microinjection, shooting with a particle gun or other methods. They can be obtained 15 from traditional sources such as extracts (e.g., bacterial, fungal or plant) and compound libraries, or can be provided by newer methods of rationale drug design. Information on functionally important regions of the genes or gene products, gained by sequence 20 comparisons and/or mutational analysis may provide a basis for drug design. Genes can be microinjected into nematodes to produce transgenic nematodes. Individual genes or cDNA and genomic DNA libraries can be screened in this manner.

Agonists and antagonists may also be derived from genes which are not cell death genes, but which interact with, regulate or bypass cell death genes. Such interacting genes may be tested by the bioassays mentioned above, as well as by in vivo genetics in nematodes. In this latter method, interacting genes are identified as secondary mutations which suppress or enhance the ced-3 mutation. The sequences of these interacting genes can then be used to identify structurally related interacting genes in other organisms.

Similarly, anti-inflammatory drugs may be developed or identified using ced-3, ICE and other family members and their encoded products. Drugs which enhance ICE activity may also be useful for boosting the inflammatory response to viral and other infections.

In addition, the availability of a number of structurally related genes makes it possible to carry out structural comparisons. Conserved regions or features of the genes or their encoded products are likely to be functionally significant for cell death and/or protease activity. This information could be helpful in designing or selecting drugs which would mimic or affect the activity of the genes.

Moreover, conservation of functional domains among

ced-3/ICE family members or their encoded products
suggests not only that these genes have similar
activities, but that they and their encoded products
function via similar mechanisms. This suggests that
mutations in conserved regions, mimetics based on

conserved regions, and agonists and antagonists which
affect the function of conserved regions of one ced3/ICE gene or encoded protein will similarly affect
other genes or encoded proteins in the family. This is
the rationale behind the use of Ced-3 inhibitors to

inhibit ICE and inflammation, and the use of antiinflammatory drugs which act by inhibiting ICE to
inhibit the ced-3 gene and reduce cell deaths (described
further below).

Furthermore, drugs which affect the cell death and/or inflammatory activities of the ced-3 and ICE genes may also affect other as yet undiscovered activities of these genes. The biology of IL-1 β and ICE is only incompletely understood at the present time, and it is very likely that other functions of both IL-1 β and ICE may be discovered. These may include new activities

or new physiological processes or diseases in which the respective cytokinetic and proteolytic activities of these molecules are involved. In either case, drugs (such as inhibitory protein portions) which affect ICE activity are likely to affect the new activities and processes, as well.

In addition, mutations and drugs which alter or mimic the activity of one member of the ced-3/ICE family can be engineered based on what is known about mutations 10 and drugs affecting another family member with which it shares a conserved region. Mutations in conserved regions which correspond to those found in another family member could be used to produce similar effects. For example, five out of nine inactivating point 15 mutations analyzed in ced-3 were found to result in alterations of amino acids which are conserved between ICE and Ced-3 (Figures 6A-B). Amino acid substitutions in ICE corresponding to those in Ced-3 are also expected to result in inactivation. The inhibitory amino-20 terminal gene portions and constitutively activated carboxyl-terminal gene portions described below are further examples of corresponding mutations which can be made in genes of the ced-3/ICE family.

Comparison of Ced-3, ICE, and related proteins may
25 also provide insights into the substrate-specificity of
ICE and related enzymes. Previous studies on ICE have
not identified a consistent consensus cleavage site. A
comparison of the Ced-3 and ICE autocleavage sites,
together with the cleavage site of pro-IL-1\beta, reveals
30 that cleavage always occurs after an Asp residue. For
this reason, it is likely that Ced-3, ICE, and related
proteins are proteases which cleave after some aspartate
residues or, perhaps at lower efficiencies, all
aspartate residues.

A further use of ced-3/ICE family members is to provide diagnostic probes (DNA, RNA, oligonucleotides and antibodies) for diseases involving cell deaths and inflammation in humans and other organisms. It is 5 likely that such diseases are associated with abnormalities in ced-3/ICE genes and their gene products. The probes can be used to detect abnormalities in the sequence, level and/or activities of the genes and encoded RNA and protein products. 10 diseases may be genetic, in which case, the probes may be used in patient and pre-natal testing, or nongenetic, in which case, RNAs and proteins may be examined. In particular, the finding that ICE is a putative cell death gene makes this gene and its 15 derivative molecules potentially useful as diagnostic probes for diseases characterized by cell deaths. Similarly, ced-3 and its derivative molecules are potentially useful for detecting abnormalities in pathologies in which inflammation is evident. The 20 usefulness of these probes may be multiplied as more genes with known physiological functions are found to be structurally related to ced-3 and ICE.

Structural Relatedness of ced-3 and the Murine NEDD-2 Gene

Database searches also revealed that another mammalian protein is similar to the Ced-3 protein (Figure 7). The murine NEDD-2 protein has 27% amino acid identity and 55% similarity to a carboxyl-terminal portion of Ced-3. The NEDD-2 protein is expressed in the brain of mouse embryos and much less in the murine adult brain; the protein is thought to be involved in the development of the murine central nervous system (Kumar et al., Biochem. Biophys. Res. Comm. 185(3):1155-1161 (1992)). The structural similarity between the

NEDD-2 and ced-3 gene products suggests that the NEDD-2 gene is also involved in cell death processes which occur during development, and further supports the hypothesis that genes which are structurally and functionally related to the nematode ced-3 gene function in a variety of organisms. Interestingly, the NEDD-2 amino acid sequence is not significantly similar to that of human ICE.

The similarity of the amino acid sequences of Ced-3 and NEDD-2 further suggests that mutations of the NEDD-2 gene which produce alterations in the protein corresponding to alterations in Ced-3 resulting from the mutations, n1129, n1164, n2426 and n1163 (see Figure 7), will inactivate the NEDD-2 gene.

This invention includes all and portions of the NEDD-2 gene, mutated NEDD-2 genes corresponding to known ced-3 mutations, RNAs and proteins encoded by the wild-type and mutated genes, and mimetics and other drugs derived from these genes and gene products, which are useful for controlling cell death.

Figures 8A and 8B show alignments of the aminoterminal and carboxyl-terminal regions, respectively, of the Ced-3 proteins of the three nematode species (C. briggsae, C. elegans, and C. vulgaris), the human and murine ICEs and the murine NEDD-2 protein (in 8B only). As shown in these figures (boxed portions), a number of amino acids are completely conserved among these structurally related proteins, and thus, are likely to be important functionally. Mutations of these sites would be expected to alter the activity of the genes.

Inhibitory Portions of the ced-3 Gene

Fusion constructs containing portions of the ced-3 gene were found to prevent programmed cell death when expressed in wild-type C. elegans. These constructs are

represented schematically in Figure 11A. The BGAFQ construct contains a portion of the ced-3 gene fused 5' of the E. coli lacZ gene and another ced-3 portion fused 3' of lacZ. The 5' ced-3 portion is the genomic sequence from a BamHI site located about 300 base pairs upstream of nucleotide 1 of the sequence shown in Figure

sequence from a BamHI site located about 300 base pairs upstream of nucleotide 1 of the sequence shown in Figure 3 to a SalI site at nucleotide 5850. This portion spans sequences 5' of the SL1 acceptor site (nucleotide 2161) to include the 372 codons of the amino-terminal region.

10 The 3' ced-3 portion of BGAFQ is the genomic sequence from a NotI site at nucleotide 5927 in the ced-3 gene to an ApaI site located about 1.5 kb downstream of nucleotide 7653 of the sequence in Figure 3. This portion contains the carboxyl-terminal codons from 398 to the end (codon 503) and 3' untranslated sequences.

The PBA construct has a smaller portion of the ced3 gene which is the genomic sequence from the same BamHI
site as in BGAFQ to a BglII site at nucleotide 3020
(Figure 11A) fused 5' of the lacZ gene. This ced-3
20 portion spans sequences 5' of the SL1 acceptor site to
include the first 149 codons of the amino-terminal
region.

Both constructs were made using the pBluescript vector (Stratagene) and fragments containing the lacz construct from the pPD vectors of Fire (EMBO J. 5:2673-2690 (1986)). The lacz-containing portion has the entire lacz coding sequence except for the first 11 codons. In addition, there is a synthetic intron and a nuclear localization signal upstream of the lacz gene and a fragment of the 3' end of the unc-54 gene downstream of the lacz gene (Figure 11B). Construct PBA was made by inserting a BamHI-ApaI fragment containing the lacz construct shown in Figure 11B from Andy Fire's vector, pPD22.04, into the BglII-ApaI fragment of the ced-3-containing plasmid, pJ40. Construct BGAFQ was

made by inserting a SalI-EagI fragment containing the same lacZ construct from pPD22.04 into the SalI-NotI fragment of pJ40A, which is pJ40 without the NotI site in the vector.

nematodes with the two constructs. These results indicate that the BGAFQ and PBA fusion constructs prevent cell deaths which normally occur in the development of the nematodes. These fusion constructs were further observed to prevent cell deaths and the apparently associated inviability caused by a loss-of-function mutation in ced-9, a gene which functions to keep certain cells from dying during nematode development, and which has been shown to act by antagonizing ced-3 and second cell death gene, ced-4.

Both constructs express β-galactosidase activity in wild-type nematodes. Since the pBluescript vector does not contain eukaryotic transcriptional or translational start sites, these signals are probably supplied by the 20 ced-3 gene portions fused 5' of lacz. Furthermore, since the PBA construct works to prevent cell death, it seems that the ced-3 portion in BGAFQ needed for inhibition is the portion fused upstream of lacz (as opposed to the portion located downstream of lacz).

25 Presumably, only the region from the BamHI site to nucleotide 3020 is needed in BFAGQ, since this is all

A construct that contains the PBA ced-3 portion but not any of the lacz portion did not prevent cell death, 30 suggesting that fusion to portions of lacz is needed for expression or action of the inhibitory gene portion.

that is contained in PBA.

These observations indicate that the amino-terminal portion of the Ced-3 protein, possibly in conjunction with a portion of $E.\ coli\ \beta$ -galactosidase, can act to prevent programmed cell deaths in $C.\ elegans$. One

plausible mechanism is that this portion of the Ced-3 protein acts in a dominant negative or antimorphic fashion, to prevent the activity of the normal Ced-3 protein. (It is known that inactivation of the Ced-3 5 protein results in an absence of programmed cell deaths.) Such dominant negative activity could be a result of the partial Ced-3 protein binding to and, thereby, inactivating the normal Ced-3 protein; consistent with this model is the finding that the active form of the structurally similar ICE protein is dimeric. Alternatively, the partial Ced-3 protein may bind to a molecule with which the normal Ced-3 protein must interact to function and by preventing this interaction, inhibits Ced-3 activity.

Due to the structural similarity of ICE to the Ced3 protein, fusion constructs encoding amino-terminal
portions of ICE would also be expected to inhibit the
activity of the ced-3 gene. In particular, those
portions of the ICE gene corresponding to the ced-3 gene
portions in BGAFQ and PBA, i.e., ICE codons 1 to 298 and
codons 1 to 111, or active subportions of these, are
expected to inhibit ced-3. A further extension of this
reasoning suggests that corresponding gene portions of
any structurally related ced-3/ICE family member would
also have an inhibitory effect on ced-3 activity.

Furthermore, the structural relatedness of the ced3 and ICE genes implies that the ICE enzyme could also
be inhibited by fusion constructs containing aminoterminal portions of the ICE gene, as well as
30 corresponding portions of other structurally related
genes, such as ced-3.

Identification of portions of the ced-3, ICE, and related genes which inhibit the ced-3 gene can be carried out by testing expression constructs containing these gene portions or their encoded products in

bioassays for cell death activity. Identification of gene portions or encoded products which inhibit ICE can be carried out using previously described assays for ICE 1) wild-type worms can be activity. For example: 5 injected with portions of the ced-3 or other structurally related gene, such as ICE, to determine if they prevent programmed cell death; 2) portions of the ICE protein or other structurally similar protein, such as Ced-3, can be co-expressed with ICE and pro-IL-1 β in 10 nematodes or cultured mammalian cells to see if they inhibit ICE-catalyzed cleavage of the IL-1 β precursor; and 3) peptides or nucleic acids containing portions of the amino acid or coding sequence of ICE or similar protein, such as Ced-3, can be tested using purified ICE 15 and synthetic substrates.

Inhibitory portions of the ced-3 gene, ICE, and structurally related genes, their encoded RNAs and proteins, and peptide and non-peptide mimetics of the proteins may be used to reduce cell deaths and/or 20 inflammation, and are, thus, useful for the treatment of diseases involving these processes. The encoded proteins and peptide and non-peptide mimetics can be delivered by various known methods and routes of drug delivery. For example, they can be administered orally 25 or by another parenteral route or by a non-parenteral route (e.g., by injection intramuscularly, intraperitoneally or intravenously or by topical administration). Alternatively, expression constructs containing the gene portions can be made using 30 heterologous transcriptional and translational signals or signals native to the gene portions. The constructs can be delivered into cells by various methods of gene therapy, such as retroviral infection.

Interestingly, those ICE gene portions
35 corresponding to the ced-3 portions of BGAFQ and PBA

encode approximately the protein fragments which result from cleavage at each of the two autocleavage sites (amino acids 103 and 297). This observation suggests that autoproteolytic conversion of the proenzyme to 5 active ICE involves cleaving off the inhibitory aminoterminal portions of the protein. Active ICE is a heterodimer composed of subunits of about 20 and 10 kilodaltons (Thornberry et al., Nature 356:768-774 (1992)). These subunits have been shown to be derived 10 from the ICE proenzyme and correspond to amino acids 120 to 297 (p20) and 317 to 404 (p10). Kinetic studies suggest that association of the two subunits is required for activity of the enzyme. It is possible that the amino-terminal region of the protein interferes with 15 this association.

This implies that mutant proteins in which the inhibitory amino-terminal regions are deleted may be constitutively activated. Thus, carboxyl-terminal portions of the Ced-3, ICE, and related proteins, and 20 constructs and RNAs expressing these portions, are potentially useful for increasing cell deaths and/or IL- 1β production. Constructs which may be used include those which express the carboxyl region of ICE, which encodes the two subunits of the active enzyme, as well 25 as those which express each of these subunits separately. In addition, it is possible that the amino region of ICE, which is not needed for ICE enzymatic activity in vitro, is important for ICE activity or the regulation of ICE activity in vivo. Consistent with 30 this idea is the finding that two of the ced-3 mutations map in this region. For this reason, a construct which expresses the amino region of Ced-3, ICE or a Ced-3/ICE gene family member may also be used. Furthermore, the NEDD-2 protein, which is similar to a carboxyl-terminal 35 portion of the Ced-3 portion, may also exhibit

constitutive activity in causing cell deaths. Thus, all or active portions of NEDD-2, and DNA and RNA encoding NEDD-2 proteins, would be expected to produce cell death activity when expressed. Drugs comprising activated 5 molecules derived from the carboxyl-terminal regions of Ced-3, ICE and other proteins of the Ced-3/ICE family and from the NEDD-2 protein, DNAs and RNAs encoding these proteins and protein fragments, as well as peptide and non-peptide mimetics, are potentially useful for 10 controlling or reducing the size of undesirable cell populations, such as cancerous cells, infected cells, cells producing autoreactive antibodies and hair follicle cells. Such drugs may also be useful for incapacitating or killing undesired organisms, such as 15 parasites, pests, and genetically engineered organisms. For example, a number of nematodes are human, animal and plant parasites.

ICE Inhibitors As Inhibitors of Cell Death

The conservation of the active site of ICE (active cysteine and surrounding amino acids) in the Ced-3 protein implies that Ced-3 is a cysteine protease which interacts with its substrate by a similar mechanism. Hence, it is likely that inhibitors of ICE which interfere with this mechanism, or chemical analogs of these inhibitors, will also inhibit Ced-3 function.

Peptide aldehydes containing the ICE recognition site:

or a substituted site in which P2 is Ala, His, Gln, Lys, Phe, Cha, or Asp, have been shown to be effective, specific, and reversible inhibitors of the protease activity of ICE (Thornberry et al., Nature 356:768-774

(1992)). These molecules are thought to act as transition analogs, which compete for ICE binding to its substrate, pro-IL-1β. Three such inhibitors have been described: Inhibitor B (Ac-Tyr-Val-Ala-Asp-CHO);
5 Inhibitor C (Ac-Tyr-D-Ala-Ala-Asp-CHO); and Inhibitor D (Ac-Tyr-Val-Lys-Asp-CHO). Of these, Inhibitor B is the most potent, with a K_i=0.76 nM compared to K_i=3 nM for D and K_i=1.5 μM for C.

In addition, the crmA gene of cowpox virus has been found to encode a serpin which specifically inhibits ICE (Ray et al., Cell 69:597-604 (1992)). The serpin acts by preventing the proteolytic activation of ICE. This inhibitor of ICE is also expected to inhibit structurally similar proteins, such as Ced-3. The crmA gene and methods for obtaining purified CrmA protein have been described (Pickup et al., Proc. Natl. Acad. Sci. USA 83:7698-7702 (1986); Ray et al., 1992 supra). This invention includes the use of inhibitors of ICE, such as peptide aldehydes, and particularly inhibitor B, and the CrmA protein, as drugs for decreasing the activity of cell death genes and, thus, for treatment of diseases characterized by cell deaths.

Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, 25 many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the following claims. For example, functional equivalents of DNAs and RNAs may be nucleic acid sequences which, through the degeneracy of the genetic code, encode the same proteins as those specifically claimed. Functional equivalents of proteins may be substituted or modified amino acid sequences, wherein the substitution or modification does not change the activity or function of the protein. A

"silent" amino acid substitution, such that a chemically similar amino acid (e.g., an acidic amino acid with another acidic amino acid) is substituted, is an example of how a functional equivalent of a protein can be produced. Functional equivalents of nucleic acids or proteins may also be produced by deletion of nonessential sequences.

The following examples illustrate the invention and are not intended to be limiting in any way.

10

EXAMPLE 1

CLONING, SEQUENCING, AND CHARACTERIZATION OF THE CED-3 GENE

MATERIALS AND METHODS

General Methods and Strains

- The techniques used for the culturing of *C. elegans* were as described by Brenner (*Genetics* 77:71-94 (1974)).

 All strains were grown at 20°C. The wild- type parent strains were *C. elegans* variety Bristol strain N2,

 Bergerac strain EM1002 (Emmons et al., Cell 32:55-65
- 20 (1983)), C. briggsae and C. vulgaris (obtained from V. Ambros). The genetic markers used are described below. These markers have been described by Brenner (1974 supra), and Hodgkin et al. (In: The Nematode Caenorhabditis elegans, Wood and the Community of C.
- 25 elegans Researchers (eds.), Cold Spring Harbor
 Laboratory, 1988, pp 491-584). Genetic nomenclature
 follows the standard system (Horvitz et al., Mol. Gen.
 Genet. 175:129-133 (1979)):
 - LG I: ced-1(e1375); unc-54(r323)
- 30 LG VI: unc-31(e928), unc-30(e191), ced-3(n717, n718, n1040, n1129, n1163, n1164, n1165, n1286,

n1949, n2426, n2430, n2433), unc-26(e205), dpy-4(e1166)

LG V: eql-1(n986); unc-76(e911)

LG X: dpy-3 (e27)

5 Isolation of Additional Alleles of ced-3

A non-complementation screen was designed to isolate new alleles of ced-3. Because animals heterozygous for ced-3(n717) in trans to a deficiency are viable (Ellis and Horvitz, Cell 44:817-829 (1986)), 10 animals carrying a complete loss-of-function ced-3 allele generated by mutagenesis were expected to be viable in trans to ced-3(n717), even if the new allele was inviable in homozygotes. Fourteen EMS mutagenized eg1-1 males were mated with ced-3(n717) unc-26(e205); 15 eg1-1(n487); dpy-3(e27) hermaphrodites. eg1-1 was used as a marker in this screen. Dominant mutations in eg1-1 cause the two hermaphrodite specific neurons, the HSNs, to undergo programmed cell death (Trent et al., Genetics 104:619-647 (1983)). The HSNs are required for normal 20 egg-laying, and egl-1(n986) hermaphrodites, which lack HSNs, are egg-laying defective (Trent et al., 1983 supra). The mutant phenotype of egl-1 is suppressed in a ced-3; egl-1 strain because mutations in ced-3 block programmed cell deaths. eg1-1 males were mutagenized 25 with EMS and crossed with ced-3(n717), unc-26(e205); eg1-1(n487); dpy-3(e27). Most cross progeny were egglaying defective because they were heterozygous for ced-3 and homozygous for eg1-1. Rare egg-laying competent animals were picked as candidates for carrying new 30 alleles of ced-3. Four such animals were isolated from about 10,000 F1 cross progeny of EMS-mutagenized animals. These new mutations were made homozygous to confirm that they carried recessive mutations of ced-3.

Molecular Biology

Standard techniques of molecular biology were used (Maniatis et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, 1983).

Two cosmid libraries were used extensively in this work: a Sau3AI partial digest genomic library of 7000 clones in the vector pHC79 and a Sau3AI partial digest genomic library of 6000 clones in the vector pJB8 (Ish-Horowicz and Burke, Nucleic Acids Res. 9:2989 (1981)).

The "right" end of MMM-C1 was cloned by cutting it with HindIII and self-ligating. The "left" end of MMM-C1 was cloned by cutting it with BglII or SalI and self-ligating.

The "right" end of Jc8 was made by digesting Jc8

15 with EcoRI and self-ligating. The "left" end of Jc8 was

made by digesting Jc8 by SalI and self-ligating.

C. elegans RNA was extracted using guanidine isothiocyanate (Kim and Horvitz, Genes & Dev. 4:357-371 (1990)). Poly(A) + RNA was selected from total RNA by a poly(dT) column (Maniatis et al., 1983 supra). To prepare stage-synchronized animals, worms were synchronized at different developmental stages (Meyer and Casson, Genetics 106:29-44 (1986)).

For DNA sequencing, serial deletions were made
25 according to a procedure developed by Henikoff (Gene
28:351-359 (1984)). DNA sequences were determined using
Sequenase and protocols obtained from US Biochemicals
with minor modifications.

The Tc1 DNA probe for Southern blots was pCe2001,
30 which contains a Bergerac Tc1 element (Emmons et al.,
Cell 32:55-65 (1983)). Enzymes were purchased from New
England Biolabs, and radioactive nucleotides were from
Amersham.

Primer extension procedures followed the proby Robert E. Kingston (In: Current Protocols in Molecular Biology, Ausubel et al. (eds.), Greene Publishing Associates and Wiley-Interscience, Ne

Polymerase chain reaction (PCR) was carried or using standard protocols supplied by the GeneAmp Kit (Perkin Elmer). The primers used for primer extension and PCR are as follows:

10 Pex2: 5' TCATCGACTTTTAGATGACTAGAGAACATC 3' (SEQ ID NO:7);

5 p. 4.8.1) with minor modifications.

Pex1: 5' GTTGCACTGCTTTCACGATCTCCCGTCTCT 3' (SEQ ID NO:8);

SL1: 5' GTTTAATTACCCAAGTTTGAG 3' (SEQ ID NO:9);

15 SL2: 5' GGTTTTAACCAGTTACTCAAG 3' (SEQ ID NO:10);

Log5: 5' CCGGTGACATTGGACACTC 3' (SEQ ID NO:11); and

Oligo10: 5' ACTATTCAACACTTG 3' (SEQ ID NO:12).

Germline Transformation

The procedure for microinjection basically follows

that of A. Fire (EMBO J. 5:2673-2680 (1986)) with
modifications: Cosmid DNA was twice purified by CsC1gradient. Miniprep DNA was used when deleted cosmids
were injected. To prepare miniprep DNA, DNA from 1.5 ml
overnight bacterial culture in superbroth (12 g Bactotryptone, 24 g yeast extract, 8 ml 50% glycerol, 900 ml
H₂O, autoclaved; after autoclaving, 100 ml 0.17 M KH₂PO₄
and 0.72 M KH₂PO₄ were added) was extracted by alkaline
lysis method as described in Maniatis et al. (1983
supra). DNA was treated with RNase A (37°, 30 minutes)
and then with protease K (55°, 30 minutes), extracted
with phenol and then chloroform, precipitated twice
(first in 0.3 M sodium acetate and second in 0.1 M

potassium acetate, pH 7.2), and resuspended in 5 μ l injection buffer as described by A. Fire (1986 supra). The DNA concentration for injection is in the range of .00 ug to 1 mg per ml.

All transformation experiments used ced-1(e1735); unc-31(e928) ced-3(n717) strain. unc-31 was used as a marker for co-transformation (Kim and Horvitz, 1990 supra). ced-1 was present to facilitate scoring of the Ced-3 phenotype. The mutations in ced-1 block the 10 engulfment process of cell death, which makes the corpses of the dead cells persist much longer than in wild-type animals (Hedgecock et al., Science 220:1277-1280 (1983)). The Ced-3 phenotype was scored as the number of dead cells present in the head of young L1 15 animals. The cosmid C10D8 or the plasmid subclones of C10D8 were mixed with C14G10 (unc-31(+)-containing) at a ratio of 2:1 or 3:1 to increase the chances that a Unc-31(+) transformant would contain the cosmid or plasmid being tested as well. Usually, 20-30 animals were 20 injected in one experiment. Non-Unc F1 progeny of the injected animal were isolated three to four days later. About 1/2 to 1/3 of the non-Unc progeny transmitted the non-Unc phenotype to F2 progeny and established a transformant line. The young L1 progeny of such non-Unc 25 transformant were checked for the number of dead cells present in the head using Nomarski optics.

RESULTS

Isolation of Additional ced-3 Alleles

All of the ced-3 alleles that existed previously
30 were isolated in screens designed to detect viable
mutants displaying the Ced phenotype (Ellis and Horvitz,
Cell 44:817-829 (1986)). Such screens may have
systematically missed any class of ced-3 mutations that

is inviable as homozygotes. For this reason, a scheme was designed that could isolate recessive lethal alleles of ced-3. Four new alleles of ced-3 (n1163, n1164, n1165, n1286) were isolated in this way. Since new 5 alleles were isolated at a frequency of about 1 in 2500, close to the frequency expected for the generation of null mutations by EMS in an average C. elegans gene (Brenner, Genetics 77:71-94 (1974); Greenwald and Horvitz, Genetics 96:147-160 (1980)), and all four 10 alleles are homozygous viable, it was concluded that the null allele of ced-3 is viable.

Mapping RFLPs near ced-3

Tc1 is a C. elegans transposable element that is thought to be immobile in the common laboratory Bristol 15 strain and in the Bergerac strain (Emmons et al., Cell 32:55-65 (1983)). In the Bristol strain, there are 30 copies of Tc1, while in the Bergerac strain, there are more than 400 copies of Tc1 (Emmons et al., 1983 supra; Finney, Ph.D. thesis, Massachusetts Institute of 20 Technology, Cambridge, Massachusetts, 1987). Because the size of the C. elegans genome is small (haploid genome size 8 x 107 bp) (Sulston and Brenner, Genetics 77:95-104 (1976)), a polymorphism due to Tc1 between the Bristol and Bergerac strains would be expected to occur 25 about once every 200 kb. Restriction fragment length polymorphisms (RFLPs) can be used as genetic markers and mapped in a manner identical to conventional mutant phenotypes. A general scheme has been designed to map Tcl elements that are dimorphic between the Bristol and 30 Bergerac strains near any gene of interest (Ruvkun et al., Genetics 121:501-516 (1989)). Once tight linkage of a particular Tc1 to a gene of interest has been

established, that Tc1 can be cloned and used to initiate chromosome walking.

A 5.1 kb Bristol-specific Tc1 EcoRI fragment was tentatively identified as containing the Tc1 closest to 5 ced-3. This Tcl fragment was cloned using cosmids from a set of Tc1-containing C. elegans Bristol genomic DNA fragments. DNA was prepared from 46 such Tc1-containing cosmids and screened using Southern blots to identify the cosmids that contain a 5.1 kb EcoRI Tcl-containing 10 fragment. Two such cosmids were identified: MMM-C1 and MMM-C9. The 5.1 kb EcoRI fragment was subcloned from MMM-C1 into pUC13 (Promega). Since both ends of Tc1 contain an EcoRV site (Rosenzweig et al., Nucleic Acids Res. 11:4201-4209 (1983)), EcoRV was used to remove Tcl 15 from the 5.1 kb EcoRI fragment, generating a plasmid that contains only the unique flanking region of this Tc1-containing fragment. This plasmid was then used to map the specific Tc1 without the interference of other Tc1 elements.

unc-30(e191) ced-3(n717) dpy-4(e1166)/+++ males 20 were crossed with Bergerac (EM1002) hermaphrodites, and Unc non-Dpy or Dpy non-Unc recombinants were picked from among the F2 progeny. The recombinants were allowed to self-fertilize, and strains that were homozygous for 25 either unc-30(e191) dpy-4(Bergerac) or unc-30(Bergerac) dpy-4(e1166) were isolated. After identifying the ced genotypes of these recombinant strains, DNA was prepared from these strains. A Southern blot of DNA from these recombinants was probed with the flanking sequence of 30 the 5.1 kb EcoRI Tcl fragment. This probe detects a 5.1 kb fragment in Bristol N2 and a 3.4 kb fragment in Bergerac. Five out of five unc-30 ced-3 dpy(+Berg) recombinants, and one of one unc-30(+Berg) ced-3 dpy-4 recombinants showed the Bristol pattern. Nine of ten 35 unc-30(+Berg) dpy-4 recombinants showed the Bergerac

pattern. Only one recombinant of unc-30(+Berg) dpy-4 resulted from a cross-over between ced-3 and the 5.1 kb Tc1 element. The genetic distance between ced-3 and dpy-4 is 2 map units (mu). Thus, this Tc1 element is located 0.1 mu on the right side of ced-3.

Cosmids MMM-C1 and MMM-C9 were used to test whether any previously mapped genomic DNA cosmids overlapped with these two cosmids. A contig of overlapping cosmids was identified that extended the cloned region near ced
10 3 in one direction.

To orient MMM-C1 with respect to this contig, both ends of MMM-C1 were subcloned and these subclones were used to probe the nearest neighboring cosmid C48D1. The "right" end of MMM-C1 does not hybridize to C48D1, while 15 the "left" end does. Therefore, the "right" end of MMM-C1 extends further away from the contig. To extend this contig, the "right" end of MMM-C1 was used to probe the filters of two cosmid libraries (Coulson et al., Proc. Natl. Acad. Sci. USA 83:7821-7825 (1986)). One clone, 20 Jc8, was found to extend MMM-C1 in the opposite direction of the contig.

RFLPs between the Bergerac and Bristol strains were used to orient the contig with respect to the genetic map. Bristol (N2) and Bergerac (EM1002) DNA was digested with various restriction enzymes and probed with different cosmids to look for RFLPs. Once such an RFLP was found, DNA from recombinants of the Bristol and Bergerac strains between ced-3 and unc-26, and between unc-30 and ced-3 was used to determine the position of the RFLP with respect to ced-3.

The "right" end of Jc8, which represents one end of the contig, detects an RFLP (nP33) when N2 and EM1002 DNA was digested with HindIII. A Southern blot of DNA from recombinants between three ced-3(+Berg) unc-26 was probed with the "right" end of Jc8. Three of three

+Berg unc-26 recombinants showed the Bristol pattern, while two of two ced-3 unc-26(+Berg) recombinants showed the Bergerac pattern. Thus, nP33 mapped very close or to the right side of unc-26.

The "left" end of Jc8 also detects a HindIII RFLP (nP34). The same Southern blot was reprobed with the Jc8 "left" end. Two of the two ced-3 unc-26(+Berg) recombinants and two of the three ced-3(+Berg) unc-26 recombinants showed the Bergerac pattern. One of the three ced-3(+Berg) unc-26 recombinants showed the Bristol pattern. The genetic distance between ced-3 and unc-26 is 0.2 mu. Thus, nP34 was mapped between ced-3 and unc-26, about 0.1 mu on the right side of ced-3.

The flanking sequence of the 5.1 kb EcoRI Tc1

15 fragment (named nP35) was used to probe the same set of recombinants. Two of three ced-3(+Berg) unc-26 recombinants and two of two ced-3 unc-26(+Berg) recombinants showed the Bristol pattern. Thus, nP35 was also found to be located between ced-3 and unc-26, about 0.1 mu on the right side of ced-3.

A similar analysis using cosmid T10H5 which contains the *HindIII* RFLP (nP36), and cosmid B0564, which contains a *HindIII* RFLP (nP37), showed that nP36 and nP37 mapped very close or to the right of unc-30.

These experiments localized the ced-3 gene to an interval of three cosmids. The positions of the RFLPs, and of ced-3, unc-30 and unc-26 on chromosome IV, and their relationships to the cosmids are shown in Figure 1. It has been demonstrated by microinjection that cosmids C37G8 and C33F2 carry the unc-30 gene (John Sulston, personal communication). Thus, the region containing the ced-3 gene was limited to an interval of two cosmids. These results are summarized in Figure 1.

Complementation of ced-3 by Germline Transformation

Cosmids that were candidates for containing the ced-3 gene were microinjected into a ced-3 mutant to see if they rescue the mutant phenotype. The procedure for 5 microinjection was that of A. Fire (EMBO J. 5:2673-2680 (1986)) with modifications. unc-31, a mutant defective in locomotion, was used as a marker for cotransformation (Kim and Horvitz, Genes & Dev. 4:357-371 (1990)), because the phenotype of ced-3 can be examined only by 10 using Nomarski optics. Cosmid C14G10 (containing unc-31(+)) and a candidate cosmid were coinjected into ced-1(e1375); unc-31(e928) ced-3(n717) hermaphrodites, and F1 non-Unc transformants were isolated to see if the non-Unc phenotype could be transmitted and established 15 as a line of transformants. Young L1 progeny of such transformants were examined for the presence of cell deaths using Nomarski optics to see whether the Ced-3 phenotype was suppressed. Cosmid C14G10 containing unc-31 alone does not rescue ced-3 activity when injected 20 into a ced-3 mutant. Table 4 summarizes the results of these transformation experiments.

As shown in Table 3, of the three cosmids injected (C43C9, W07H6 and C48D1), only C48D1 rescued the Ced-3 phenotype (2/2 non-Unc transformants rescued the Ced-3 phenotype). One of the transformants, nEX2, appears to be rescued by an extra-chromosomal array of injected cosmids (Way and Chalfie, Cell 54:5-16 (1988)), which is maintained as an unstable duplication, since only 50% of the progeny of a non-Unc Ced(+) animal are non-Unc Ced(+). Since the non-Unc Ced(+) phenotype of the other transformant (nIS1) is transmitted to all of its progeny, it is presumably an integrated transformant. L1 ced-1 animals contain an average of 23 cell corpses in the head. L1 ced-1; ced-3 animals contain an average of 0.3 cell corpses in the head. ced-1; unc-31 ced-3; nIS1 and ced-1; unc-31 ced-3; nEX2 animals contain an

average of 16.4 and 14.5 cell corpses in the head, respectively. From these results, it was concluded that C48D1 contains the ced-3 gene.

In order to locate ced-3 more precisely within the cosmid C48D1, this cosmid was subcloned and the subclones were tested for the ability to rescue ced-3 mutants. C48D1 DNA was digested with restriction enzymes that cut rarely within the cosmid and the remaining cosmid was self-ligated to generate a subclone. Such subclones were then injected into a ced-3 mutant to look for completion. When C48D1 was digested with BamHI and self-ligated, the remaining 14 kb subclone (named C48D1-28) was found to rescue the Ced-3 phenotype when injected into a ced-3 mutant (Figure 2 and Table 4). C48D1-28 was then partially digested with BglII and self-ligated. Clones of various lengths were isolated and tested for their ability to rescue ced-3.

One clone, C48D1-43, which did not contain a 1.7 kb 20 BqlII fragment of C48D1-28, was able to rescue ced-3 (Figure 2 and Table 4). C48D1-43 was further subcloned by digesting with BamHI and ApaI to isolate a 10 kb BamHI-ApaI fragment. This fragment was subcloned into pBSKII+ to generate pJ40. pJ40 can restore Ced-3+ 25 phenotype when microinjected into a ced-3 mutant. pJ40 was subcloned by deleting a 2 kb BglII-ApaI fragment to generate pJ107. pJ107 was also able to rescue the Ced-3 phenotype when microinjected into a ced-3 mutant. Deletion of 0.5 kb on the left side of pJ107 could be 30 made by ExoIII digestion (as in pJ107del28 and pJ107del34) without affecting Ced-3 activity; in fact, one transgenic line, nEX17, restores full Ced-3 activity. However, the ced-3 rescuing ability was significantly reduced when 1 kb was deleted on the left 35 side of pJ107 (as in pJ107del12 and pJ107del27), and the WO 93/25694 PCT/US93/05705

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ability was completely eliminated when a 1.8 kb Sall-BglII fragment was deleted on the right side of pJ107 (as in pJ55 and pJ56), suggesting that this SalI site is likely to be in the ced-3 coding region. From these experiments, ced-3 was localized to a DNA fragment of 7.5 kb. These results are summarized in Figure 2 and Table 4.

ced-3 Transcript

pJ107 was used to probe a Northern blot of N2 RNA

10 and detected a band of 2.8 kb. Although this transcript
is present in 12 ced-3 mutant animals, subsequent
analysis showed that all 12 ced-3 mutant alleles contain
mutations in the genomic DNA that codes for this mRNA
(see below), thus establishing this RNA as a ced-3

15 transcript.

The developmental expression pattern of ced-3 was determined by hybridizing a Northern blot of RNA from animals of different stages (eggs, L1 through L4 larvae and young adult) with the ced-3 cDNA subclone pJ118.

20 Such analysis revealed that the ced-3 transcript is most abundant during embryonic development, which is the period when most programmed cell deaths occur, but it was also detected during the L1 through L4 larval stages and is present in relatively high levels in young

25 adults. This result suggests that ced-3 is not only expressed in cells undergoing programmed cell death.

Since ced-3 and ced-4 are both required for programmed cell death in C. elegans, one of the genes might act as a regulator of transcription of the other 30 gene. To examine if ced-4 regulates the transcription of ced-3, RNA was prepared from eggs of ced-4 mutants (n1162, n1416, n1894, and n1920), and a Northern blot was probed with the ced-3 cDNA subclone pJ118. The presence of RNA in each lane was confirmed with an actin

I probe. Such an experiment showed that the level of ced-3 transcript is normal in ced-4 mutants. This indicates that ced-4 is unlikely to be a transcriptional regulator of ced-3.

5 Isolation of a ced-3 cDNA

To isolate cDNA of ced-3, pJ40 was used as a probe to screen a cDNA library of N2 (Kim and Horvitz, Genes & Dev. 4:357-371 (1990)). Seven cDNA clones were isolated. These cDNAs can be divided into two groups:

10 one is 3.5 kb and the other 2.5 kb. One cDNA from each group was subcloned and analyzed further. pJ85 contains the 3.5 kb cDNA. Experiments showed that pJ85 contains a ced-3 cDNA fused to an unrelated cDNA; on Northern blots of N2 RNA, the pJ85 insert hybridizes to two RNA transcripts, and on Southern blots of N2 DNA, pJ85 hybridizes to one more band than pJ40 (ced-3 genomic DNA) does. pJ87 contains the 2.5 kb cDNA. On Northern blots, pJ87 hybridizes to a 2.8 kb RNA and on Southern blots, it hybridizes only to bands to which pJ40 hybridizes. Thus, pJ87 contains only ced-3 cDNA.

To show that pJ87 does contain the ced-3 cDNA, a frameshift mutation was made in the SalI site of pJ40 corresponding to the SalI site in the pJ87 cDNA. Constructs containing the frameshift mutation failed to rescue the Ced-3 phenotype when microinjected into ced-3 mutant animals, suggesting that ced-3 activity has been eliminated.

ced-3 Sequence

The DNA sequence of pJ87 was determined (Figure 3).

30 pJ87 contains an insert of 2.5 kb which has an open reading frame of 503 amino acids (Figure 3; SEQ ID NO:2). The 5' end of the cDNA contains 25 bp of poly-A/T sequence, which is probably an artifact of cloning

and is not present in the genomic sequence. The cDNA ends with a poly-A sequence, suggesting that it contains the complete 3' end of the transcript. 1 kb of pJ87 insert is untranslated 3' region and not all of it is essential for ced-3 expression, since genomic constructs with deletions of 380 bp of the 3' end can still rescue ced-3 mutants (pJ107 and its derivatives, see Figure 2).

To confirm the DNA sequence obtained from the ced-3 cDNA and to study the structure of the ced-3 gene, the 10 genomic sequence of the ced-3 gene in the plasmid pJ107 was determined (Figure 3; SEQ ID NO:1). Comparison of the ced-3 genomic and cDNA sequences revealed that the ced-3 gene has seven introns that range in size from 54 bp to 1195 bp (Figure 4A). The four largest introns, as 15 well as sequences 5' of the start codon, were found to contain repetitive elements (Figure 3). Five types of repetitive elements were found, some of which have been previously characterized in non-coding regions of other C. elegans genes, such as fem-1 (Spence et al., Cell 20 60:981-990 (1990)), lin-12 (J. Yochem, personal communication), and myoD (Krause et al., Cell 63:907-919 (1990)). Of these, repeat 1 was also found in fem-1 and myoD, repeat 3 in lin-12 and fem-1, repeat 4 in lin-12, and repeats 2 and 5 were novel repetitive elements.

amplification was used to determine the location and nature of the 5' end of the ced-3 transcript. Two primers (Pex1 and Pex2) were used for the primer extension reaction. The Pex1 reaction yielded two major bands, whereas the Pex2 reaction gave one band. The Pex2 band corresponded in size to the smaller band from the Pex1 reaction, and agreed in length with a possible transcript that is trans-spliced to a C. elegans splice leader (Bektesh, Genes & Devel. 2:1277-1283 (1988)) at a consensus splice acceptor at position 2166 of the

genomic sequence (Figure 3). The nature of the larger Pex1 band is unclear.

To confirm the existence of this trans-spliced message in wild-type worms, total *C. elegans* RNA was PCR 5 amplified using the SL1-Log5 and SL2-Log5 primer pairs, followed by a reamplification using the SL1-Oligo10 and SL2-Oligo10 primer pairs. The SL1 reaction yielded a fragment of the predicted length. The identity of this fragment was confirmed by sequencing. Thus, at least some, if not most, of the *ced-3* transcript is transspliced to SL1. Based on this result, the start codon of the *ced-3* message was assigned to the methionine encoded at position 2232 of the genomic sequence (Figure 3).

The DNA sequences of 12 EMS-induced ced-3 alleles 15 were also determined (Figure 3 and Table 1). Nine of the 12 are missense mutations. Two of the 12 are nonsense mutations, which might prematurely terminate the translation of ced-3. These nonsense ced-3 mutants 20 confirmed that the ced-3 gene is not essential for viability. One of the 12 mutations is an alteration of a conserved splicing acceptor G, and another has a change of a 70% conserved C at the splice site, which could also generate a stop codon even if the splicing is 25 correct. Interestingly, these EMS-induced mutations are in either the N-terminal quarter or C-terminal half of In fact, 9 of the 12 mutations occur the protein. within the region of ced-3 that encodes the last 100 amino acids of the protein. Mutations are notably 30 absent from the middle part of the ced-3 gene (Figure 4A).

Ced-3 Protein Contains A Region Rich in Serines

The Ced-3 protein is very hydrophilic and no 35 significantly hydrophobic region can be found that might

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be a trans-membrane domain (Figure 5). The Ced-3 protein is rich in serine. From amino acid 78 to amino acid 205 of the Ced-3 protein, 34 out of 127 amino acids are serine. Serine is often the target of

5 serine/threonine protein kinases (Edelman, Ann. Rev. Biochem. 56:567-613 (1987)). For example, protein kinase C can phosphorylate serines when they are flanked on their amino and carboxyl sides by basic residues (Edelman, 1987 supra). Four of the serines in the Ced-3 protein are flanked by arginines (Figures 6A-B). The same serine residues might also be the target of related Ser/Thr kinases.

To identify the functionally important regions of the Ced-3 protein, genomic DNAs containing the ced-3

15 genes from two related nematode species, C. briggsae
(SEQ ID NO:5) and C. vulgaris (SEQ ID NO:6) were cloned and sequenced. Sequence comparison of the three ced-3
gene products showed that the non-serine-rich region of the proteins is highly conserved (Figure 9). In C.

20 briggsae and C. vulgaris, many amino acids in the serine-rich region are dissimilar compared to the C. elegans Ced-3 protein. It seems that what is important in the serine-rich region is the overall serine-rich feature rather than the exact amino acid sequence.

25 This hypothesis is also supported by analysis of

This hypothesis is also supported by analysis of ced-3 mutations in C. elegans: none of the 12 EMS-induced mutations is in the serine-rich region, suggesting that mutations in this region might not affect the function of the Ced-3 protein and thus, could not be isolated in the screen for ced-3 mutants.

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EXAMPLE 2

A COMMON MECHANISM OF CELL DEATH IN VERTEBRATES AND INVERTEBRATES

Results from previous studies reported in the 5 scientific literature suggest that cell deaths in a variety of organisms, including vertebrates as well as invertebrates, share a common mechanism which involves the activation of genes. These studies are consistent with the hypothesis that genes similar to the C. elegans 10 ced-3 and ced-4 genes may be involved in the cell deaths that occur in vertebrates, although certain observations have led some to distinguish vertebrate cell deaths from the programmed cell deaths observed in such invertebrates as nematodes and insects. Some vertebrate 15 cell deaths share certain characteristics with the programmed cell deaths in C. elegans that are controlled by ced-3 and ced-4. For example, up to 14% of the neurons in the chick dorsal root ganglia die immediately after their births, before any signs of differentiation 20 (Carr and Simpson, Dev. Brain Res. 2:57-162 (1982)). Genes like ced-3 and ced-4 could well function in this class of vertebrate cell death.

and ced-4 genes are expressed by cells that undergo
programmed cell death, so that these genes may not act
through cell-cell interactions (Yuan and Horvitz, Dev.
Biol. 138:33-41 (1990)). Many cell deaths in
vertebrates seem different in that they appear to be
controlled by interactions with target tissues. For
example, it is thought that a deprivation of targetderived growth factors is responsible for vertebrate
neuronal cell deaths (Hamburger and Oppenheim, Neurosci.
Comment. 1:39-55 (1982)); Thoenen et al., in: Selective
Neuronal Death, Wiley, New York, 1987, Vol. 126, pp. 82-

85). However, even this class of cell death could involve genes like ced-3 and ced-4, since pathways of cell death involving similar genes and mechanisms might be triggered in a variety of ways. Supporting this idea 5 are several in vitro and in vivo studies which show that the deaths of vertebrate as well as invertebrate cells can be prevented by inhibitors of RNA and protein synthesis, suggesting that activation of genes are required for these cell deaths (Martin et al., J. Cell 10 Biol. 106:829-844 (1988); Cohen and Duke, J. Immunol. 132:38-42 (1984); Oppenheim and Prevette, Neurosci. Abstr. 14:368 (1988); Stanisic et al., Invest. Urol. 16:19-22 (1978); Oppenheim et al., Dev. Biol. 138:104-113 (1990); Fahrbach and Truman, in: Selective Neuronal 15 Death, Ciba Foundation Symposium, 1987, No. 126, pp. 65-81). It is possible that the genes induced in these dying vertebrate and invertebrate cells are cell death genes which are structurally related to the C. elegans ced-3 or ced-4 genes.

Also supporting the hypothesis that cell death in C. elegans is mechanistically similar to cell death in vertebrates is the observation that the protein product of the C. elegans gene ced-9 is similar in sequence to the human protein Bcl-2. ced-9 has been shown to prevent cells from undergoing programmed cell death during nematode development by antagonizing the activities of ced-3 and ced-4 (Hengartner, et al., Nature 356:494-499 (1992)). The bcl-2 gene has also been implicated in protecting cells against cell death.

30 It seems likely that the genes and proteins with which ced-9 and bcl-2 interact are similar as well.

Sites of Mutations in the ced-3 Gene

Table 1

Consequence	L to F	G to R	G to S	Q to termination	Splice acceptor loss	Q to termination	W to termination	A to V	A to V	A to V	E to K	S to F
Codon	27	65	360	403	ı	412	428	449	449	466	483	486
Nucleotide	2310	2487	5757	5940	6297	6322	6342	6434	6434	6485	6535	7020
Mutation	C to T	G to A	G to A	C to T	G to A	c to T	G to A	c to T	C to T	C to T	G to A	C to T
Allele	n1040	n718	n2433	n1164	n717	n1949	n.1286	n1129	n1165	n2430	n2426	n1163

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Nucleotide and codon positions correspond to the numbering in Figure 3.

Table 2

<u>ced-3-lacZ</u> Fusions Which

<u>Prevent Programmed Cell Death</u>

		Average #	Number
Strain Name	Construct	Extra Cells	of Animals
N2 (wild-type)	-	0.1	40
nEx 121	PBA	2.0	23
nEx 70	PBA	2.4	31
nEx 67	BGAFQ	2.1	18
nEx 66	BGAFQ	2.1	25

Table 3
Summary of Transformation Experiments
Using Cosmids in the ced-3 Region

Cosmid <u>injected</u>	No. of non-Unc transformants	Ced-3 phenotype	Strain name
C43C9; C14G10	1	-	MT4302
W07H6; C14G10	3	-	MT4299
		-	MT4300
		-	MT4301
C48D1; C14G10	2	+	MT4298
		+	MT4303

Animals injected were of genotype: ced-1(e1735); unc-31(e929) ced-3(n717).

-53Table 4

The expression of ced-3(+) transformants

Genotype	DNA injected	Average No. cell deaths in L1 head	No. Animals scored
ced-1	-	23	20
ced-1; ced-3	-	0.3	10
ced-1; nIS1	C48D1;	16.4	20
unc-31 ced-3	C14G10	:	
ced-1; unc-31 ced-3; nIS1/+		14.5	20
ced-1; unc-31	C48D1;	13.2	10/14
ced-3; nEX2	C14G10	0	4/14
ced-1; unc-31 ced-3; nEX10	C48D1-28; C14G10	12	9/10
-	•	0	1 of 10
ced-1; unc-31 ced-3; nEX9	C48D1-28; C14G10	12	10
ced-1; unc-31 ced-3; nEX11	C48D1-43	16.7	10/13
		Abnormal cell deaths	3/13
ced-1; unc-31 ced-3; nEX13	pJ40; C14G10	13.75	4/4

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1	T	2	b	1	e	4	C	0	n	t	<u>i</u>	n	u	e	₫

ced-1; unc-31 ced-3; nEX17	pJ107de128, pJ107de134 C14G10	23	12/14
		0	2/14
ced-1; unc-31 ced-3; nEX18	pJ107de128, pJ107de1134 C14G10	12.8	9/10
		0	1/10
ced-1; unc-31 ced-3; nEX19	pJ107de128, pJ107de134 G14G10	10.6	5/6
		0	1/6
ced-1; unc-31 ced-3; nEX16	pJ107del12, pJ107del27 C14G10	7.8	12/12

Alleles of the genes used are ced-1(e1735), unc-31(e928), and ced-3(n717).

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT
 - (A) NAME: Massachusetts Institute of Technology
 - (B) STREET: 77 Massachusetts Avenue
 - (C) CITY: Cambridge
 - (D) STATE OR PROVINCE: Massachusetts

 - (E) COUNTRY: U.S.A. (F) POSTAL CODE: 02139
- (ii) TITLE OF INVENTION: Inhibitors of Ced-3 and Related Proteins
- (iii) NUMBER OF SEQUENCES: 14
- (iv) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: diskette

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 7653 base pairs (B) TYPE: nucleic acid

 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

AGATCTGAAA	TAAGGTGATA	AATTAATAAA	TTAAGTGTAT	TTCTGAGGAA	ATTTGACTGT	60
TTTAGCACAA	TTAATCTTGT	TTCAGAAAAA	AAGTCCAGTT	TTCTAGATTT	TTCCGTCTTA	120
TTGTCGAATT	AATATCCCTA	TTATCACTTT	TTCATGCTCA	TCCTCGAGCG	GCACGTCCTC	180
AAAGAATTGT	GAGAGCAAAC	GCGCTCCCAT	TGACCTCCAC	ACTCAGCCGC	CAAAACAAAC	240
GTTCGAACAT	TCGTGTGTTG	TGCTCCTTTT	CCGTTATCTT	GCAGTCATCT	TTTGTCGTTT	300
TTTTCTTTGT	TCTTTTTGTT	GAACGTGTTG	CTAAGCAATT	ATTACATCAA	TTGAAGAAAA	360
GGCTCGCCGA	TTTATTGTTG	CCAGAAAGAT	TCTGAGATTC	TCGAAGTCGA	TTTTATAATA	420
TTTAACCTTG	GTTTTTGCAT	TGTTTCGTTT	AAAAAAACCA	CTGTTTATGT	GAAAAACGAT	480
TAGTTTACTA	ATAAAACTAC	TTTTAAACCT	TTACCTTTAC	CTCACCGCTC	CGTGTTCATG	540
GCTCATAGAT	TTTCGATACT	CAAATCCAAA	AATAAATTTA	CGAGGGCAAT	TAATGTGAAA	600
CAAAAACAAT	CCTAAGATTT	CCACATGTTT	GACCTCTCCG	GCACCTTCTT	CCTTAGCCCC	660
ACCACTCCAT	CACCTCTTTG	GCGGTGTTCT	TCGAAACCCA	CTTAGGAAAG	CAGTGTGTAT	720
CTCATTTGGT	ATGCTCTTTT	CGATTTTATA	GCTCTTTGTC	GCAATTTCAA	TGCTTTAAAC	780

AATCCAAATC	GCATTATATT	TGTGCATGGA	GGCAAATGAC	GGGGTTGGAA	TCTTAGATGA	840
GATCAGGAGC	TTTCAGGGTA	AACGCCCGGT	TCATTTTGTA	CCACATTTCA	TCATTTTCCT	900
GTCGTCCTTG	GTATCCTCAA	CTTGTCCCGG	TTTTGTTTTC	GGTACACTCT	TCCGTGATGC	960
CACCTGTCTC	CGTCTCAATT	ATCGTTTAGA	AATGTGAACT	GTCCAGATGG	GTGACTCATA	1020
TTGCTGCTGC	TACAATCCAC	TTTCTTTTCT	CATCGGCAGT	CTTACGAGCC	CATCATAAAC	1080
TTTTTTTCC	GCGAAATTTG	CAATAAACCG	GCCAAAAACT	TTCTCCAAAT	TGTTACGCAA	1140
TATATACAAT	CCATAAGAAT	ATCTTCTCAA	TGTTTATGAT	TTCTTCGCAG	CACTTTCTCT	1200
TCGTGTGCTA	ACATCTTATT	TTTATAATAT	TTCCGCTAAA	ATTCCGATTT	TTGAGTATTA	1260
ATTTATCGTA	AAATTATCAT	AATAGCACCG	AAAACTACTA	AAAATGGTAA	AAGCTCCTTT	1320
TAAATCGGCT	CGACATTATC	GTATTAAGGA	ATCACAAAAT	TCTGAGAATG	CGTACTGCGC	1380
AACATATTTG	ACGGCAAAAT	ATCTCGTAGC	GAAAACTACA	GTAATTCTTT	AAATGACTAC	1440
TGTAGCGCTT	GTGTCGATTT	ACGGGCTCAA	TTTTTGAAAA	TAATTTTTTT	TTTCGAATTT	1500
TGATAACCCG	TAAATCGTCA	CAACGCTACA	GTAGTCATTT	AAAGGATTAC	TGTAGTTCTA	1560
GCTACGAGAT	ATTTTGCGCG	CCAAATATGA	CTGTAATACG	CATTCTCTGA	ATTTTGTGTT	1620
TCCGTAATAA	TTTCACAAGA	TTTTGGCATT	CCACTTTAAA	GGCGCACAGG	ATTTATTCCA	1680
ATGGGTCTCG	GCACGCAAAA	AGTTTGATAG	ACTTTTAAAT	TCTCCTTGCA	TTTTTAATTC	1740
AATTACTAAA	ATTTTCGTGA	ATTTTTCTGT	TAAAATTTTT	AAAATCAGTT	TTCTAATATT	1800
TTCCAGGCTG	ACAAACAGAA	ACAAAAACAC	AACAAACATT	TTAAAAATCA	GTTTTCAAAT	1860
TAAAAATAAC	GATTTCTCAT	TGAAAATTGT	GTTTTATGTT	TGCGAAAATA	AAAGAGAACT	1920
GATTCAAAAC	AATTTTAACA	AAAAAAAACC	CCAAAATTCG	CCAGAAATCA	AGATAAAAAA	1980
TTCAAGAGGG	TCAAAATTTT	CCGATTTTAC	TGACTTTCAC	CTTTTTTTC	GTAGTTCAGT	2040
GCAGTTGTTG	GAGTTTTTGA	CGAAAACTAG	GAAAAAAATC	GATAAAAATT	ACTCAAATCG	2100
AGCTGAATTT	TGAGGACAAT	GTTTAAAAAA	AAACACTATT	TTTCCAATAA	TTTCACTCAT	2160
TTTCAGACTA	AATCGAAAAT	CAAATCGTAC	TCTGACTACG	GGTCAGTAGA	GAGGTCAACC	2220
ATCAGCCGAA	GATGATGCGT	CAAGATAGAA	GGAGCTTGCT	AGAGAGGAAC	ATTATGATGT	2280
TCTCTAGTCA	TCTAAAAGTC	GATGAAATTC	TCGAAGTTCT	CATCGCAAAA	CAAGTGTTGA	2340
ATAGTGATAA	TGGAGATATG	ATTAATGTGA	GTTTTTAATC	GAATAATAAT	TTTAAAAAAA	2400
AATTGATAAT	ATAAAGAATA	TTTTTGCAGT	CATGTGGAAC	GGTTCGCGAG	AAGAGACGGG	2460
AGATCGTGAA	AGCAGTGCAA	CGACGGGGAG	ATGTGGCGTT	CGACGCGTTT	TATGATGCTC	2520

TTCGCTCTAC	GGGACACGAA	GGACTTGCTG	AAGTTCTTGA	ACCTCTCGCC	AGATCGTAGG	2580
TTTTTAAAGT	TCGGCGCAAA	AGCAAGGGTC	TCACGGAAAA	AAGAGGCGGA	TCGTAATTTT	2640
GCAACCCACC	GGCACGGTTT	TTTCCTCCGA	AAATCGGAAA	TTATGCACTT	TCCCAAATAT	2700
TTGAAGTGAA	ATATATTTA	TTTACTGAAA	GCTCGAGTGA	TTATTTATTT	TTTAACACTA	2760
ATTTTCGTGG	CGCAAAAGGC	CATTTTGTAG	ATTTGCCGAA	AATACTTGTC	ACACACACAC	2820
ACACACATCT	CCTTCAAATA	TCCCTTTTTC	CAGTGTTGAC	TCGAATGCTG	TCGAATTCGA	2880
GTGTCCAATG	TCACCGGCAA	GCCATCGTCG	GAGCCGCGCA	TTGAGCCCCG	CCGGCTACAC	2940
TTCACCGACC	CGAGTTCACC	GTGACAGCGT	CTCTTCAGTG	TCATCATTCA	CTTCTTATCA	3000
GGATATCTAC	TCAAGAGCAA	GATCTCGTTC	TCGATCGCGT	GCACTTCATT	CATCGGATCG	3060
ACACAATTAT	TCATCTCCTC	CAGTCAACGC	ATTTCCCAGC	CAACCTTGTA	TGTTGATGCG	3120
AACACTAAAT	TCTGAGAATG	CGCATTACTC	AACATATTTG	ACGCGCAAAT	ATCTCGTAGC	3180
GAAAAATACA	GTAACCCTTT	AAATGACTAT	TGTAGTGTCG	ATTTACGGC	TCGATTTTCG	3240
AAACGAATAT	ATGCTCGAAT	TGTGACAACG	AATTTTAATT	TGTCATTTTT	GTGTTTTCTT	3300
TTGATATTTT	TGATCAATTA	ATAAATTATT	TCCGTAAACA	GACACCAGCG	CTACAGTACT	3360
CTTTTAAAGA	GTTACAGTAG	TTTTCGCTTC	AAGATATTTT	GAAAAGAATT	TTAAACATTT	3420
TGAAAAAAA	TCATCTAACA	TGTGCCAAAA	CGCTTTTTTC	AAGTTTCGCA	GATTTTTTGA	3480
TTTTTTTCAT	TCAAGATATG	CTTATTAACA	CATATAATTA	TCATTAATGT	GAATTTCTTG	3540
TAGAAATTTT	GGGCTTTTCG	TTCTAGTATG	CTCTACTTTT	GAAATTGCTC	AACGAAAAA	3600
TCATGTGGTT	TGTTCATATG	AATGACGAAA	AATAGCAATT	TTTTATATAT	TTTCCCCTAT	3660
TCATGTTGTG	CAGAAAAATA	GTAAAAAAGC	GCATGCATTT	TTCGACATTT	TTTACATCGA	3720
ACGACAGCTC	ACTTCACATG	CTGAAGACGA	GAGACGCGGA	GAAATACCAC	ACATCTTTCT	3780
GCGTCTCTCG	TCTTCAGCAT	GTGAAATGGG	ATCTCGGTCG	ATGTAAAAAA	ATGTCGAATA	3840
ATGTAAAAA	TGCATGCGTT	TTTTTACACT	TTTCTGCACA	AATGAATAGG	GGGAAAATGT	3900
ATTAAAATAC	ATTTTTTGTA	TTTTTCAACA	TCACATGATT	AACCCCATTA	TTTTTTCGTT	3960
GAGCAACTTA	AAAAGTAGAG	AATATTAGAG	CGAAAACCAA	AATTTCTTCA	AGATATTACC	4020
TTTATTGATA	ATTATAGATG	TTAATAAGCA	TATCTTGAAT	GAAAGTCAGC	AAAAATATGT	4080
GCGAAACACC	TGAAAAAAAT	CAAAAATTCT	GCGAAAATTG	AAAAAATGCA	TTAAAATACA	4140
TTTTTGCATT	TTTCTACATC	ACATGAATGT	AGAAAATTAA	AAGGGAAATC	AAAATTTCTA	4200
GAGGATATAA	TTGAATGAAA	CATTGCGAAA	TTAAAATGTG	CGAAACGTCA	AAAAAGAGGA	4260

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AATTTGGGTA	TCAAAATCGA	TCCTAAAACC	AACACATTTC	AGCATCCGCC	AACTCTTCAT	4320
TCACCGGATG	CTCTTCTCTC	GGATACAGTT	CAAGTCGTAA	TCGCTCATTC	AGCAAAGCTT	4380
CTGGACCAAC	TCAATACATA	TTCCATGAAG	AGGATATGAA	CTTTGTCGAT	GCACCAACCA	4440
TAAGCCGTGT	TTTCGACGAG	AAAACCATGT	ACAGAAACTT	CTCGAGTCCT	CGTGGAATGT	4500
GCCTCATCAT	AAATAATGAA	CACTTTGAGC	AGATGCCAAC	ACGGAATGGT	ACCAAGGCCG	4560
ACAAGGACAA	TCTTACCAAT	TTGTTCAGAT	GCATGGGCTA	TACGGTTATT	TGCAAGGACA	4620
ATCTGACGGG	AAGGGTACGG	CGAAATTATA	TTACCCAAAC	GCGAAATTTG	CCATTTTGCG	4680
CCGAAAATGT	GGCGCCCGGT	CTCGACACGA	CAATTTGTGT	TAAATGCAAA	AATGTATAAT	4740
TTTGCAAAAA	ACAAAATTTT	GAACTTCCGC	GAAAATGATT	TACCTAGTTT	CGAAATTTTC	4800
GTTTTTTCCG	GCTACATTAT	GTGTTTTTC	TTAGTTTTTC	TATAATATTT	GATGTAAAAA	4860
ACCGTTTGTA	AATTTTCAGA	CAATTTTCCG	CATACAAAAC	TTGATAGCAC	GAAATCAATT	4920
TTCTGAATTT	TCAAAATTAT	CCAAAAATGC	ACAATTTAAA	ATTTGTGAAA	ATTGGCAAAC	4980
GGTGTTTCAA	TATGAAATGT	ATTTTTAAAA	ACTTTAAAAA	CCACTCCGGA	AAAGCAATAA	5040
AAATCAAAAC	AACGTCACAA	TTCAAATTCA	AAAGTTATTC	ATCCGATTTG	TTTATTTTTG	5100
CAAAATTTGA	AAAAATCATG	AAGGATTTAG	AAAAGTTTTA	TAACATTTT	TCTAGATTTT	5160
TCAAAATTTT	TTTTAACAAA	TCGAGAAAAA	GAGAATGAAA	AATCGATTTT	AAAAATATCC	5220
ACAGCTTCGA	GAGTTTGAAA	TTACAGTACT	CCTTAAAGGC	GCACACCCCA	TTTGCATTGG	5280
ACCAAAAATT	TGTCGTGTCG	AGACCAGGTA	CCGTAGTTTT	TGTCGCAAAA	ATTGCACCAT	5340
TGGACAATAA	ACCTTCCTAA	TCACCAAAAA	GTAAAATTGA	AATCTTCGAA	AAGCCAAAAA	5400
ATTCAAAAAA	AAAGTCGAAT	TTCGATTTT	TTTTTGGTTT	TTTGGTCCCA	AAAACCAAAA	5460
AAATCAATTT	TCTGCAAAAT	ACCAAAAAGA	AACCCGAAAA	AATTTCCCAG	CCTTGTTCCT	5520
AATGTAAACT	GATATTTAAT	TTCCAGGGAA	TGCTCCTGAC	AATTCGAGAC	TTTGCCAAAC	5580
ACGAATCACA	CGGAGATTCI	GCGATACTC	TGATTCTATC	: ACACGGAGAF	GAGAATGTGA	5640
TTATTGGAGT	TGATGATATA	CCGATTAGTA	CACACGAGAT	TATGATCT1	CTCAACGCGG	5700
CAAATGCTCC	CCGTCTGGCG	AATAAGCCG	AAATCGTTTI	TGTGCAGGC	TGTCGAGGCG	5760
GTTCGTTTTT	TATTTTAT	TTAATATAA!	A TATTTTAAA	RAATTCATT	TCAGAACGTC	5820
GTGACAATGG	ATTCCCAGTO	TTGGATTCT	TCGACGGAGT	TCCTGCATT	CTTCGTCGTG	5880
GATGGGACAF	1 TCGAGACGG	CCATTGTTC	A ATTTTCTTGG	ATGTGTGCG	CCGCAAGTTC	5940
AGGTTGCAAT	TTAATTTCT	GAATGAGAA	r attecttear	A AAAATCTAAJ	ATAGATTTTT	6000

ATTCCAGAAA	GTCCCGATCG	AAAAATTGCG	ATATAATTAC	GAAATTTGTG	ATAAAATGAC	6060
AAACCAATCA	GCATCGTCGA	TCTCCGCCCA	CTTCATCGGA	TTGGTTTGAA	AGTGGGCGGA	6120
GTGAATTGCT	GATTGGTCGC	AGTTTTCAGT	TTAGAGGGAA	TTTAAAAATC	GCCTTTTCGA	6180
AAATTAAAA	TTGATTTTTT	CAATTTTTC	GAAAAATATT	CCGATTATTT	TATATTCTTT	6240
GGAGCGAAAG	CCCCGTCCTG	TAAACATTTT	TAAATGATAA	TTAATAAATT	TTTGCAGCAA	6300
GTGTGGAGAA	AGAAGCCGAG	CCAAGCTGAC	ATTCTGATTC	GATACGCAAC	GACAGCTCAA	6360
TATGTTTCGT	GGAGAAACAG	TGCTCGTGGA	TCATGGTTCA	TTCAAGCCGT	CTGTGAAGTG	6420
TTCTCGACAC	ACGCAAAGGA	TATGGATGTT	GTTGAGCTGC	TGACTGAAGT	CAATAAGAAG	6480
GTCGCTTGTG	GATTTCAGAC	ATCACAGGGA	TCGAATATTT	TGAAACAGAT	GCCAGAGGTA	6540
CTTGAAACAA	ACAATGCATG	TCTAACTTTT	AAGGACACAG	AAAAATAGGC	AGAGGCTCCT	6600
TTTGCAAGCC	TGCCGCGCGT	CAACCTAGAA	TTTTAGTTTT	TAGCTAAAAT	GATTGATTTT	6660
GAATATTTTA	TGCTAATTTT	TTTGCGTTAA	ATTTTGAAAT	AGTCACTATT	TATCGGGTTT	6720
CCAGTAAAAA	ATGTTTATTA	GCCATTGGAT	TTTACTGAAA	ACGAAAATTT	GTAGTTTTTC	6780
AACGAAATTT	ATCGATTTT	AAATGTAAAA	AAAAATAGCG	AAAATTACAT	CAACCATCAA	6840
GCATTTAAGC	CAAAATTGTT	AACTCATTTA	AAAATTAATT	CAAAGTTGTC	CACGAGTATT	6900
ACACGGTTGG	CGCGCGGCAA	GTTTGCAAAA	CGACGCTCCG	CCTCTTTTTC	TGTGCGGCTT	6960
GAAAACAAGG	GATCGGTTTA	GATTTTTCCC	CAAAATTTAA	ATTAAATTTC	AGATGACATC	7020
CCGCCTGCTC	AAAAAGTTCT	ACTTTTGGCC	GGAAGCACGA	AACTCTGCCG	TCTAAAATTC	7080
ACTCGTGATT	CATTGCCCAA	TTGATAATTG	TCTGTATCTT	CTCCCCAGT	TCTCTTTCGC	7140
CCAATTAGTT	TAAAACCATG	TGTATATTGT	TATCCTATAC	TCATTTCACT	TTATCATTCT	7200
ATCATTTCTC	TTCCCATTTT	CACACATTTC	CATTTCTCTA	CGATAATCTA	AAATTATGAC	7260
GTTTGTGTCT	CGAACGCATA	ATAATTTTAA	TAACTCGTTT	TGAATTTGAT	TAGTTGTTGT	7320
GCCCAGTATA	TATGTATGTA	CTATGCTTCT	ATCAACAAAA	TAGTTTCATA	GATCATCACC	7380
CCAACCCCAC	CAACCTACCG	TACCATATTC	ATTTTTGCCG	GGAATCAATT	TCGATTAATT	7440
TTAACCTATT	TTTTCGCCAC	AAAAAATCTA	ATATTTGAAT	TAACGAATAG	CATTCCCATC	7500
TCTCCCGTGC	CGGAATGCCT	CCCGGCCTTT	TAAAGTTCGG	AACATTTGGC	AATTATGTAT	7560
AAATTTGTAG	GTCCCCCCA	TCATTTCCCG	CCCATCATCT	CAAATTGCAT	TCTTTTTTCG	7620
CCGTGATATC	CCGATTCTGG	TCAGCAAAGA	TCT			7653

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 503 amino acids
 - (B) TYPE: amino acid (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Met Arg Gln Asp Arg Arg Ser Leu Leu Glu Arg Asn Ile Met Met

Phe Ser Ser His Leu Lys Val Asp Glu Ile Leu Glu Val Leu Ile Ala

Lys Gln Val Leu Asn Ser Asp Asn Gly Asp Met Ile Asn Ser Cys Gly

Thr Val Arg Glu Lys Arg Arg Glu Ile Val Lys Ala Val Gln Arg Arg

Gly Asp Val Ala Phe Asp Ala Phe Tyr Asp Ala Leu Arg Ser Thr Gly

His Glu Gly Leu Ala Glu Val Leu Glu Pro Leu Ala Arg Ser Val Asp

Ser Asn Ala Val Glu Phe Glu Cys Pro Met Ser Pro Ala Ser His Arg

Arg Ser Arg Ala Leu Ser Pro Ala Gly Tyr Thr Ser Pro Thr Arg Val

His Arg Asp Ser Val Ser Ser Val Ser Ser Phe Thr Ser Tyr Gln Asp

Ile Tyr Ser Arg Ala Arg Ser Arg Ser Arg Ser Arg Ala Leu His Ser

Ser Asp Arg His Asn Tyr Ser Ser Pro Pro Val Asn Ala Phe Pro Ser

Gln Pro Ser Ser Ala Asn Ser Ser Phe Thr Gly Cys Ser Ser Leu Gly

Tyr Ser Ser Ser Arg Asn Arg Ser Phe Ser Lys Ala Ser Gly Pro Thr

Gln Tyr Ile Phe His Glu Glu Asp Met Asn Phe Val Asp Ala Pro Thr

Ile Ser Arg Val Phe Asp Glu Lys Thr Met Tyr Arg Asn Phe Ser Ser 225 230 235

Pro Arg Gly Met Cys Leu Ile Ile Asn Asn Glu His Phe Glu Gln Met - 250

Pro Thr Arg Asn Gly Thr Lys Ala Asp Lys Asp Asn Leu Thr Asn Leu Phe Arg Cys Met Gly Tyr Thr Val Ile Cys Lys Asp Asn Leu Thr Gly Arg Gly Met Leu Leu Thr Ile Arg Asp Phe Ala Lys His Glu Ser His 295 Gly Asp Ser Ala Ile Leu Val Ile Leu Ser His Gly Glu Glu Asn Val Ile Ile Gly Val Asp Asp Ile Pro Ile Ser Thr His Glu Ile Tyr Asp Leu Leu Asn Ala Ala Asn Ala Pro Arg Leu Ala Asn Lys Pro Lys Ile Val Phe Val Gln Ala Cys Arg Gly Glu Arg Arg Asp Asn Gly Phe Pro **Val Leu Asp Ser Val Asp Gly Val Pro Ala Phe Leu Arg Arg Gly Trp** Asp Asn Arg Asp Gly Pro Leu Phe Asn Phe Leu Gly Cys Val Arg Pro Gln Val Gln Gln Val Trp Arg Lys Lys Pro Ser Gln Ala Asp Ile Leu Ile Arg Tyr Ala Thr Thr Ala Gln Tyr Val Ser Trp Arg Asn Ser Ala Arg Gly Ser Trp Phe Ile Gln Ala Val Cys Glu Val Phe Ser Thr His Ala Lys Asp Met Asp Val Val Glu Leu Leu Thr Glu Val Asn Lys Lys Val Ala Cys Gly Phe Gln Thr Ser Gln Gly Ser Asn Ile Leu Lys Gln Met Pro Glu Met Thr Ser Arg Leu Leu Lys Lys Phe Tyr Phe Trp Pro 485 Glu Ala Arg Asn Ser Ala Val 500

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1373 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA

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(ix) FEATURE:

- (A) NAME/KEY: CDS
 (B) LOCATION: 18..1232
 (D) OTHER INFORMATION: /product= "human interleukin-1β convertase"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

AAAAGGAGAG AAAAGCC		AAG GTC CTG AAG GAG Lys Val Leu Lys Glo 5		50
CTG TTT ATC CGT TC Leu Phe Ile Arg Se 15				98
GAA TTA TTA CAG AC. Glu Leu Leu Gln Th . 30		Asn Lys Glu Glu M		146
AAA CGT GAA AAT GC Lys Arg Glu Asn Al 45	T ACA GTT ATG a Thr Val Met 50	GAT AAG ACC CGA G Asp Lys Thr Arg A 55	CT TTG ATT GAC la Leu Ile Asp	194
TCC GTT ATT CCG AA Ser Val Ile Pro Ly 60				242
ATT TGT GAA GAA GA Ile Cys Glu Glu As 8	p Ser Tyr Leu			290
GAT CAA ACA TCT GG Asp Gln Thr Ser Gl 95				338
CTT TCT TCC TTT CC Leu Ser Ser Phe Pr 110		Ala Val Gln Asp A		386
CCC ACA TCC TCA GG Pro Thr Ser Ser Gl 125	C TCA GAA GGG y Ser Glu Gly 130	AAT GTC AAG CTT T Asn Val Lys Leu C 135	GC TCC CTA GAA Cys Ser Leu Glu	434
GAA GCT CAA AGG AT Glu Ala Gln Arg Il 140				482
ATG GAC AAG TCA AG Met Asp Lys Ser Se 16	r Arg Thr Arg			530
GAA TTT GAC AGT AT Glu Phe Asp Ser Il 175				578

					CTA Leu											626
					TCG Ser											674
					AAG Lys 225											722
					GAA Glu											770
GTC Val	CCA Pro	GAT Asp	ATA Ile 255	CTA Leu	CAA Gln	CTC Leu	AAT Asn	GCA Ala 260	ATC Ile	TTT Phe	AAC Asn	ATG Met	TTG Leu 265	AAT Asn	ACC Thr	818
					TTG Leu											866
					AGC Ser											914
					CTA Leu 305											962
					GCC Ala											1010
					AAT Asn											1058
					CTC Leu											1106
		Val					Arg								CAG Gln	1154
CCA Pro 380	Asp	GGT Gly	AGA Arg	GCG Ala	CAG Gln 385	ATG Met	CCC Pro	ACC Thr	ACT Thr	GAA Glu 390	Arg	GTG Val	ACT Thr	TTG Leu	ACA Thr 395	1202
	GA TGT TTC TAC CTC TTC CCA GGA CAT TAAAATAAGG AAACTGTATG								1249							
Arg	Сув	Phe	Tyr	Leu 400		Pro	Gly	His	405	405						
AATGTCTGCG GGCAGGAAGT GAAGAGATCG TTCTGTAAAA GGTTTTTGGA ATTATGTCTG										G 1309						

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CTGAATAATA	AACTTTTTT	GAAATAATAA	ATCTGGTAGA	AAAATGAAAA	AAAAAAAA	1369
AAAA						1373

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 404 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4: Met Ala Asp Lys Val Leu Lys Glu Lys Arg Lys Leu Phe Ile Arg Ser Met Gly Glu Gly Thr Ile Asn Gly Leu Leu Asp Glu Leu Leu Gln Thr Arg Val Leu Asn Lys Glu Glu Met Glu Lys Val Lys Arg Glu Asn Ala Thr Val Met Asp Lys Thr Arg Ala Leu Ile Asp Ser Val Ile Pro Lys 50 60 Gly Ala Gln Ala Cys Gln Ile Cys Ile Thr Tyr Ile Cys Glu Glu Asp
65 70 75 80 Ser Tyr Leu Ala Gly Thr Leu Gly Leu Ser Ala Asp Gln Thr Ser Gly Asn Tyr Leu Asn Met Gln Asp Ser Gln Gly Val Leu Ser Ser Phe Pro 105 Ala Pro Gln Ala Val Gln Asp Asn Pro Ala Met Pro Thr Ser Ser Gly

Ser Glu Gly Asn Val Lys Leu Cys Ser Leu Glu Glu Ala Gln Arg Ile

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Trp Lys Gln Lys Ser Ala Glu Ile Tyr Pro Ile Met Asp Lys Ser Ser

Arg Thr Arg Leu Ala Leu Ile Ile Cys Asn Glu Glu Phe Asp Ser Ile

Pro Arg Arg Thr Gly Ala Glu Val Asp Ile Thr Gly Met Thr Met Leu 180

Leu Gln Asn Leu Gly Tyr Ser Val Asp Val Lys Lys Asn Leu Thr Ala

Ser Asp Met Thr Thr Glu Leu Glu Ala Phe Ala His Arg Pro Glu His 215

Lys Thr Ser Asp Ser Thr Phe Leu Val Phe Met Ser His Gly Ile Arg 225 230 Glu Gly Ile Cys Gly Lys Lys His Ser Glu Gln Val Pro Asp Ile Leu 250 Gln Leu Asn Ala Ile Phe Asn Met Leu Asn Thr Lys Asn Cys Pro Ser 265 Leu Lys Asp Lys Pro Lys Val Ile Ile Ile Gln Ala Cys Arg Gly Asp 275 280 Ser Pro Gly Val Val Trp Phe Lys Asp Ser Val Gly Val Ser Gly Asn Leu Ser Leu Pro Thr Thr Glu Glu Phe Glu Asp Asp Ala Ile Lys Lys 310 Ala His Ile Glu Lys Asp Phe Ile Ala Phe Cys Ser Ser Thr Pro Asp Asn Val Ser Trp Arg His Pro Thr Met Gly Ser Val Phe Ile Gly Arg Leu Ile Glu His Met Gln Glu Tyr Ala Cys Ser Cys Asp Val Glu Glu Ile Phe Arg Lys Val Arg Phe Ser Phe Glu Gln Pro Asp Gly Arg Ala 375 Gln Met Pro Thr Thr Glu Arg Val Thr Leu Thr Arg Cys Phe Tyr Leu 390 395 Phe Pro Gly His

- (2) INFORMATION FOR SEQ ID NO:5:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 505 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (ix) FEATURE:
 - (A) NAME/KEY: unsure
 - (B) LOCATION: at every Xaa
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Met Met Arg Gln Asp Arg Trp Leu Leu Glu Arg Asn Ile Leu Glu Phe 1 5 10 15

Ser Ser Lys Leu Gln Ala Asp Leu Ile Leu Asp Val Leu Ile Ala Lys 20 25 30

Gln Val Leu Asn Ser Asp Asn Gly Asp Val Ile Asn Ser Cys Arg Thr Glu Arg Asp Asn Glu Lys Glu Ile Val Lys Ala Val Gln Arg Arg Gly Asp Glu Ala Phe Asp Ala Phe Tyr Asp Ala Leu Arg Asp Thr Gly His Asn Asp Leu Ala Asp Val Leu Met Pro Leu Ser Arg Pro Xaa Xaa Xaa Asn Pro Val Pro Met Glu Cys Pro Met Ser Pro Ser Ser His Arg Arg 105 Ser Arg Ala Leu Ser Pro Pro Xaa Tyr Ala Ser Pro Thr Arg Val His Arg Asp Ser Ile Ser Ser Val Ser Ser Phe Thr Ser Thr Tyr Gln Asp Val Tyr Ser Arg Ala Arg Ser Ser Ser Arg Ser Ser Arg Pro Leu Gln Ser Ser Asp Arg His Asn Tyr Met Ser Ala Ala Thr Ser Phe Pro Ser Gln Pro Xaa Ser Ala Asn Ser Ser Phe Thr Gly Cys Ala Ser Leu Gly Tyr Ser Ser Ser Arg Asn Arg Ser Phe Ser Lys Thr Ser Ala Gln Ser Gln Tyr Ile Phe His Glu Glu Asp Met Asn Tyr Val Asp Ala Pro Thr Ile His Arg Val Phe Asp Glu Lys Thr Met Tyr Arg Asn Phe Ser Ser Pro Arg Gly Leu Cys Leu Ile Ile Asn Asn Glu His Phe Glu Gln Met Pro Thr Arg Asn Gly Thr Lys Ala Asp Lys Asp Asn Leu Thr Asn Ile Phe Arg Cys Met Gly Tyr Thr Val Ile Cys Lys Asp Asn Leu Thr Gly Arg Glu Met Leu Ser Thr Ile Arg Ser Phe Gly Arg Asn Asp Met His 295 Gly Asp Ser Ala Ile Leu Val Ile Leu Ser His Gly Glu Xaa Asn Val 305 Ile Ile Gly Val Asp Asp Val Ser Val Asn Val His Glu Ile Tyr Asp Leu Leu Asn Ala Ala Asn Ala Pro Arg Leu Ala Asn Lys Pro Lys Leu

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 340
 345
 345
 350
 350

 Val
 Phe
 Val
 Gln
 Ala
 Cys
 Arg
 Gly
 Glu
 Arg
 Arg
 Asp
 Asp
 Asp
 Gly
 Val
 Pro
 Ser
 Leu
 Ile
 Arg
 Arg
 Arg
 Arg
 Gly
 Trp

 Asp
 Asp
 Asp
 Gly
 Arg
 Leu
 Phe
 Asp
 Glu
 Leu
 Arg
 Gly
 Val
 Arg
 <t

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 480 amino acids

Glu Asp Arg Gly Arg Asn Ser Ala Val

- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (ix) FEATURE:
 - (A) NAME/KEY: unsure
 - (B) LOCATION: at every Xaa
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Thr Val Ser Ile Ser Leu Ile Ile Ala Arg Gln Val Leu Asn Ser Asp 1 10 15

Asn Xaa Xaa Met Ile Asn Ser Cys Gly Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa 20 25 30

105 Arg Ser Ser Arg Pro Leu His Thr Ser Asp Arg His Asn Tyr Val Ser Pro Ser Asn Ser Phe Gln Ser Gln Pro Ala Ser Ala Asn Ser Ser Phe Thr Gly Ser Ser Ser Leu Gly Tyr Ser Ser Ser Arg Thr Arg Ser Tyr 165 Ser Lys Ala Ser Ala His Ser Gln Tyr Ile Phe His Glu Glu Asp Met 185 Asn Tyr Val Asp Ala Pro Thr Ile His Arg Val Phe Asp Glu Lys Thr 200 Met Tyr Arg Asn Phe Ser Thr Pro Arg Gly Leu Cys Leu Ile Ile Asn 215 Asn Glu His Phe Glu Gln Met Pro Thr Arg Asn Gly Thr Lys Pro Asp 230 235 Lys Asp Asn Ile Ser Asn Leu Phe Arg Cys Met Gly Tyr Ile Val His Cys Lys Asp Asn Leu Thr Gly Arg Xaa Met Met Leu Thr Ile Arg Asp Phe Ala Lys Asn Glu Thr His Gly Asp Ser Ala Ile Leu Val Ile Xaa 280 Ser His Gly Glu Glu Asn Val Ile Ile Gly Val Asp Asp Val Ser Val 295 Asn Val His Glu Ile Tyr Xaa Leu Leu Asn Ala Ala Asn Ala Pro Arg 315 Leu Ala Asn Lys Pro Lys Leu Val Phe Val Gln Ala Cys Arg Gly Glu Arg Arg Asp Val Gly Phe Pro Val Leu Asp Ser Val Asp Gly Val Pro 345

Ala Leu Ile Arg Arg Gly Trp Asp Lys Gly Asp Gly Pro 365 Xaa Xaa 365 Phe Leu Gly Cys Val Arg Pro 375 Gln Ala Gln Gln Val Trp Arg Lys 380 Pro Ser Gln Ala Asp Ile Leu Ile Ala Tyr Ala Thr Thr Ala Glr 385 Thr Ala Glr 390 Val Ser Trp Arg Asn Ser Ala Arg Gly Ser Trp Phe Ile Gln Ala 405 Ser Ala Arg Gly Ser Trp Phe Ile Gln Ala 415 Cys Glu Val Phe Ser Leu His Ala Lys Asp Met Asp Val Val Gly 430 Leu Thr Glu Val Asn Lys Lys Val Ala Cys Gly Phe Gln Thr Ser 445 Gly Ala Asn Ile Leu Lys Gln Met Pro Xaa Leu Thr Ser Arg Leu 455 Lys Phe Tyr Phe Trp Pro Glu Asp Arg Asn Arg Ser Ser Ala 465									
Pro Ser Gln Ala Asp Ile Leu Ile Ala Tyr Ala Thr Thr Ala Glr 385 Val Ser Trp Arg Asn Ser Ala Arg Gly Ser Trp Phe Ile Gln Ala 410 Cys Glu Val Phe Ser Leu His Ala Lys Asp Met Asp Val Val Glu 420 Leu Thr Glu Val Asn Lys Lys Val Ala Cys Gly Phe Gln Thr Ser 435 Gly Ala Asn Ile Leu Lys Gln Met Pro Xaa Leu Thr Ser Arg Leu 450 Lys Lys Phe Tyr Phe Trp Pro Glu Asp Arg Asn Arg Ser Ser Ala	. Asr								
Val Ser Trp Arg Asn Ser Ala Arg Gly Ser Trp Phe Ile Gln Ala 410 Cys Glu Val Phe Ser Leu His Ala Lys Asp Met Asp Val Val Glu 420 Leu Thr Glu Val Asn Lys Lys Val Ala Cys Gly Phe Gln Thr Ser 435 Gly Ala Asn Ile Leu Lys Gln Met Pro Xaa Leu Thr Ser Arg Leu 450 Lys Lys Phe Tyr Phe Trp Pro Glu Asp Arg Asn Arg Ser Ser Ala	Lys								
Cys Glu Val Phe Ser Leu His Ala Lys Asp Met Asp Val Val Glu 420 Leu Thr Glu Val Asn Lys Lys Val Ala Cys Gly Phe Gln Thr Ser 435 Gly Ala Asn Ile Leu Lys Gln Met Pro Xaa Leu Thr Ser Arg Leu 450 Lys Lys Phe Tyr Phe Trp Pro Glu Asp Arg Asn Arg Ser Ser Ala	Tyr 400								
Leu Thr Glu Val Asn Lys Lys Val Ala Cys Gly Phe Gln Thr Ser 435 Gly Ala Asn Ile Leu Lys Gln Met Pro Xaa Leu Thr Ser Arg Leu 450 Lys Lys Phe Tyr Phe Trp Pro Glu Asp Arg Asn Arg Ser Ser Ala									
435 Gly Ala Asn Ile Leu Lys Gln Met Pro Xaa Leu Thr Ser Arg Leu 450 Lys Lys Phe Tyr Phe Trp Pro Glu Asp Arg Asn Arg Ser Ser Ala	. Leu								
450 455 460 Lys Lys Phe Tyr Phe Trp Pro Glu Asp Arg Asn Arg Ser Ser Ala	Glr								
	. Leu								
\cdot	Val 480								
INFORMATION FOR SEQ ID NO:7:									

- (2)
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 30 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single

 - (D) TOPOLOGY: linear
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

TCATCGACTT TTAGATGACT AGAGAACATC

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- (2) INFORMATION FOR SEQ ID NO:8:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 30 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single

 - (D) TOPOLOGY: linear
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

GTTGCACTGC TTTCACGATC TCCCGTCTCT

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- (2) INFORMATION FOR SEQ ID NO:9:
 - (i) SEQUENCE CHARACTERISTICS:
 - (\bar{A}) LENGTH: 21 base pairs

 - (B) TYPE: nucleic acid (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

_	7	0	-

	(xi)	SEQUE	NCE	DES	CRIPT	ion:	SEQ	ID	NO:	9:							
GTTT	AATTA	C CCA	AGTT	TGA	G												21
(2)	INFOR	MATIC	N FO	R SI	EQ ID	NO:	10:										
	(i)	(A) (B) (C)	LENG TYPE	TH: : ni NDE	21 b uclei DNESS	RIST ase p c ac : sin	air d										
	(xi)	SEQUE	ENCE	DES	CRIPI	CION:	SEQ	ID	NO:	10:							
GGTT	TTAAC	C AGI	TACT	CAA	G												21
(2)	INFOR	MATIC	ON FO	R S	EQ II	NO:	11:										
	(i)	(A) (B) (C)	LENG TYPE STRA	TH: : n NDE	19 h uclei DNESS	RIST: pase ; lc ac: s: si: lnear	pair id										
	(xi)	SEQUI	ENCE	DES	CRIPT	: NOI	SEQ	ID	NO:	11:							
CCGG	TGACA	T TGO	GACAC	TC													19
(2)	INFOR	DITAM	ON FO	R S	EQ II	NO:	12:			-							
	(i)	(A) (B) (C)	LENG TYPE STRA	TH: : n NDE	15 h ucle: DNES	ERIST base ic ac ic si inear	pair id	s									
	(xi)	SEQUI	ENCE	DES	CRIP	rion:	SEQ	ID	NO:	12:							
ACTA	ATTCA	C AC	ITG														15
(2)	INFO	ITAM	ON FO	R S	EQ II	o no:	13:										
	(i)	(A) (B)	LENG	TH:	171 mino	ERIST amin acid inear	ICS:	ids									
	(ii)	MOLE	CULE	TYP	E: p	rotei	n										
	(xi)	SEQU	ENCE	DES	CRIP:	rion:	SEC] ID	NO:	13:							
	Met 1	Leu '	Thr V	/al	Gln ' 5	Val T	yr A	irg :	Thr	Ser 10	Gln	Lys	Cys	Ser	Ser 15	Ser	

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 Lys
 His
 Val
 Val
 Glu
 Val
 Leu
 Leu
 Asp
 Pro
 Leu
 Glu
 Thr
 Ser
 Phe
 Cys

 Ser
 Leu
 Leu
 Leu
 Leu
 Tyr
 Glu
 Thr
 Asp
 Arg
 Gly
 Val

 Asp
 Gln
 Asp
 Gly
 Lys
 Asp
 His
 Thr
 Gln
 Ser
 Pro
 Gly
 Cys
 Glu
 Glu
 Gln
 Ser
 Pro
 Gly
 Cys
 Glu
 Glu
 Glu
 Met
 Lys
 Met
 Arg
 Glu
 Glu
 Leu
 Met
 Lys
 Gly
 Asp
 Met
 Met

(2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 402 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Met Ala Asp Lys Ile Leu Arg Ala Lys Arg Lys Gln Phe Ile Asn Ser 1 10 15

Val Ser Ile Gly Thr Ile Asn Gly Leu Leu Asp Glu Leu Leu Glu Lys 20 25 30

Arg Val Leu Asn Gln Glu Glu Met Asp. Lys Ile Lys Leu Ala Asn Ile 35 40 45

Thr Ala Met Asp Lys Ala Arg Asp Leu Cys Asp His Val Ser Lys Lys 50 60

Gly Pro Gln Ala Ser Gln Ile Phe Ile Thr Tyr Ile Cys Asn Glu Asp 65 70 75 80

Cys Tyr Leu Ala Gly Ile Leu Glu Leu Gln Ser Ala Pro Ser Ala Glu Thr Phe Val Ala Thr Glu Asp Ser Lys Gly Gly His Pro Ser Ser Ser Glu Thr Lys Glu Glu Gln Asn Lys Glu Asp Gly Thr Phe Pro Gly Leu Thr Gly Thr Leu Lys Phe Cys Pro Leu Glu Lys Ala Gln Lys Leu Trp Lys Glu Asn Pro Ser Glu Ile Tyr Pro Ile Met Asn Thr Thr Arg Thr Arg Leu Ala Leu Ile Ile Cys Asn Thr Glu Phe Gln His Leu Ser Pro Arg Val Gly Ala Gln Val Asp Leu Arg Glu Met Lys Leu Leu Leu 185 Glu Asp Leu Gly Tyr Thr Val Lys Val Lys Glu Asn Leu Thr Ala Leu Glu Met Val Lys Glu Val Lys Glu Phe Ala Ala Cys Pro Glu His Lys 210 220 Thr Ser Asp Ser Thr Phe Leu Val Phe Met Ser His Gly Ile Gln Glu Gly Ile Cys Gly Thr Thr Tyr Ser Asn Glu Val Ser Asp Ile Leu Lys Val Asp Thr Ile Phe Gln Met Met Asn Thr Leu Lys Cys Pro Ser Leu Lys Asp Lys Pro Lys Val Ile Ile Ile Gln Ala Cys Arg Gly Glu Lys 280 Gln Gly Val Val Leu Leu Lys Asp Ser Val Arg Asp Ser Glu Glu Asp Phe Leu Thr Asp Ala Ile Phe Glu Asp Asp Gly Ile Lys Lys Ala His Ile Glu Lys Asp Phe Ile Ala Phe Cys Ser Ser Thr Pro Asp Asn Val Ser Trp Arg His Pro Val Arg Gly Ser Leu Phe Ile Glu Ser Leu Ile Lys His Met Lys Glu Tyr Ala Trp Ser Cys Asp Leu Glu Asp Ile Phe Arg Lys Val Arg Phe Ser Phe Glu Gln Pro Glu Phe Arg Leu Gln Met 375 380

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Pro Thr Ala Asp Arg Val Thr Leu Thr Lys Arg Phe Tyr Leu Phe Pro 385 390 395

Gly His

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CLAIMS

- 1. An inhibitor of the activity of the ced-3 gene, comprising a portion of the ced-3 gene.
- 2. The inhibitor of Claim 1, wherein the gene portion is a portion of the nucleotide sequence of Figure 3 (SEQ ID NO:1), selected from the group consisting of:
 - a) nucleotides 1 to approximately 5850;
 - b) nucleotides 1 to approximately 3020; and
- 10 c) an inhibitory subportion (a) and (b).
 - 3. The inhibitor of Claim 1, wherein the gene portion encodes an amino acid sequence of the Ced-3 protein shown in Figures 6A-B (SEQ ID NO:2), selected from the group consisting of:
- 15 a) amino acids 1 to approximately 372;
 - b) amino acids 1 to approximately 149; and
 - c) an inhibitory subportion of (a) and (b).
- 4. The inhibitor of Claim 1, further comprising a heterologous structural gene fused 3' of the gene portion.
 - 5. The inhibitor of Claim 4, wherein the structural gene is E. coli lacz.
- 6. The inhibitor of Claim 1, further comprising a transcriptional signal and a translational signal suitable for expression of the gene portion in a host cell.

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- 7. The inhibitor of Claim 6, wherein the transcriptional signal and the translational signal are those of the ced-3 gene.
- 8. An inhibitor of the activity of the ced-3 gene,
 5 comprising RNA encoded by the sense strand of a
 nucleotide sequence of Figure 3 (SEQ ID NO:1), the
 nucleotide sequence selected from the group
 consisting of:
 - a) nucleotides 1 to approximately 5850;

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- b) nucleotides 1 to approximately 3020; and
- c) an inhibitory subportion of (a) and (b).
- 9. An inhibitor of the activity of the ced-3 gene, comprising protein having an amino acid sequence of the Ced-3 protein shown in Figures 6A-B (SEQ ID NO:2), selected from the group consisting of:
 - a) amino acids 1 to approximately 372;
 - b) amino acids 1 to approximately 149; and
 - c) an inhibitory subportion of (a) and (b).
- 10. An inhibitor of the activity of the ced-3 gene,
 20 comprising a non-peptide mimetic of the inhibitor of Claim 9.
 - 11. The inhibitor of Claim 1, consisting essentially of a construct selected from BGAFQ and PBA.
- 12. The inhibitor of Claim 1, comprising the encoded product of a construct selected from BGAFQ and PBA.

- 13. The inhibitor of Claim 1, comprising a non-peptide mimetic of the protein encoded by a construct selected from BGAFQ and PBA.
- 14. An inhibitor of the activity of the ced-3 gene,
 5 comprising protein having an amino acid sequence of
 ICE shown in Figures 6A-B (SEQ ID NO:4), selected
 from the group consisting of:
 - a) amino acids 1 to 298;
 - b) amino acids 1 to 111; and
- 10 c) an inhibitory subportion of (a) and (b).
 - 15. An inhibitor of the activity of the ced-3 gene, comprising a portion of the ICE gene which encodes the protein of Claim 14, or an inhibitory subportion of said gene.
- 15 16. An inhibitor of the activity of the ced-3 gene, comprising RNA encoded by the gene portion of Claim 15.
- 17. An inhibitor of the activity of the ced-3 gene, comprising a non-peptide mimetic of the protein of Claim 14.
 - 18. An inhibitor of the activity of the ced-3 gene, comprising a portion of the protein product of a gene which is structurally related to the ced-3 gene, said protein portion corresponding to an amino acid sequence of the Ced-3 protein shown in Figures 6A-B (SEQ ID NO:2), selected from the group consisting of:
 - a) amino acids 1 to approximately 372;

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- b) amino acids 1 to approximately 149; and
- c) an inhibitory subportion of (a) and (b).
- 19. An inhibitor of the activity of the ced-3 gene, comprising a portion of a gene which is structurally related to the ced-3 gene, said gene portion encoding the protein of Claim 18 or an inhibitory subsection of said gene portion.

- 20. An inhibitor of the activity of the ced-3 gene, comprising RNA encoded by the gene portion of Claim 10
 19.
 - 21. An inhibitor of the activity of the ced-3 gene, comprising a non-peptide mimetic of the protein portion of Claim 18.
- 22. An inhibitor of the activity of the ICE gene
 comprising a portion of said gene which encodes an
 amino sequence of ICE shown in Figures 6A-B (SEQ ID
 NO:4), selected from the group consisting of:
 - a) amino acids 1 to approximately 298;
 - b) amino acids 1 to approximately 111; and
- 20 c) an inhibitory subportion of (a) and (b).
 - 23. The inhibitor of Claim 22, further comprising a heterologous structural gene fused 3' of the gene portion.
- 24. The inhibitor of Claim 22, further comprising a transcriptional signal and a translational signal suitable for expression of the gene portion in a host cell.

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- 25. An inhibitor of the activity of the ICE gene, comprising RNA encoded by the gene portion of Claim 22.
- 26. An inhibitor of the activity of the ICE gene, comprising an amino acid sequence of ICE shown in Figures 6A-B (SEQ ID NO:4), selected from the group consisting of:
 - a) amino acids 1 to approximately 298;
 - b) amino acids 1 to approximately 111; and
- 10 c) an inhibitory subportion of (a) and (b).
 - 27. An inhibitor of the activity of the ICE gene, comprising a non-peptide mimetic of the protein of Claim 26.
- 28. An inhibitor of the activity of the ICE gene,
 15 comprising a portion of the ced-3 gene.
 - 29. The inhibitor of Claim 28, wherein said gene portion is a nucleotide sequence of Figure 3 (SEQ ID NO:1), selected from the group consisting of:
 - a) nucleotides 1 to approximately 5850;
- 20 b) nucleotides 1 to approximately 3020; and
 - c) an inhibitory subportion of (a) and (b).
 - 30. The inhibitor of Claim 28, wherein the gene portion encodes an amino acid sequence of the Ced-3 protein shown in Figures 6A-B (SEQ ID NO:2), selected from the group consisting of:
 - a) amino acids 1 to approximately 372;

- .b) amino acids 1 to approximately 149; and
- c) an inhibitory subportion of (a) and (b).

- 31. The inhibitor of Claim 28, further comprising a heterologous structural gene fused 3' of the gene portion.
- 32. The inhibitor of Claim 28, further comprising a transcriptional signal and a translational signal suitable for expression of the gene portion in a host cell.
- 33. An inhibitor of the activity of the ICE gene, comprising RNA encoded by the sense strand of a portion of the ced-3 gene, said gene portion which is a nucleotide sequence of Figure 3 (SEQ ID NO:1), selected from the group consisting of:
 - a) nucleotides 1 to approximately 5850;
 - b) nucleotides 1 to approximately 3020; and
- c) an inhibitory subportion of (a) and (b).
 - 34. An inhibitor of the activity of the ICE gene, comprising protein having an amino acid sequence of the Ced-3 protein shown in Figures 6A-B (SEQ ID NO:4), selected from the group consisting of:
- a) amino acids 1 to approximately 372;
 - b) amino acids 1 to approximately 149; and
 - c) an inhibitory subportion of (a) and (b).
- 35. An inhibitor of the activity of the ICE gene,comprising a non-peptide mimetic of the protein ofClaim 34.
 - 36. The inhibitor of Claim 28, consisting essentially of a construct selected from BGAFQ and PBA.

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- 37. An inhibitor of the activity of the ICE gene, comprising the encoded product of a construct selected from BGAFQ and PBA.
- 38. An inhibitor of the activity of the ICE gene
 comprising a portion of the protein product of a
 gene which is structurally related to said ICE
 gene, said protein portion corresponding to an
 amino acid sequence of the Ced-3 protein shown in
 Figures 6A-B (SEQ ID NO:2), selected from the group
 consisting of:
 - a) amino acids 1 to approximately 372;
 - b) amino acids 1 to approximately 149; and
 - c) an inhibitory subportion of (a) and (b).
- 39. An inhibitor of the activity of the ICE gene,
 15 comprising a portion of a gene which is
 structurally related to the ICE gene, said gene
 portion encoding the protein of Claim 38, or an
 inhibitory subsection of said gene portion.
- 40. An inhibitor of the activity of the ICE gene,
 20 comprising RNA encoded by the gene portion of Claim
 39.
 - 41. An inhibitor of the activity of the ICE gene, comprising a non-peptide mimetic of the protein of Claim 38.
- 25 42. An inhibitor of the activity of a gene belonging to the ced-3/ICE family of structurally related genes, comprising DNA selected from the group consisting of:

	a)	a portion of the nucleotide sequence of Figure						
		3 (SEQ ID NO:1), selected from the group						
		consisting of:						
		 nucleotides 1 to approximately 5850; 						
5		2) nucleotides 1 to approximately 3020; and						
		3) an inhibitory subportion of (a.1) and						
		(a.2);						
	b)	DNA encoding an amino acid sequence of the						
		Ced-3 protein shown in Figures 6A-B (SEQ ID						
10		NO:2), selected from the group consisting of:						
		 amino acids 1 to approximately 372; 						
		2) amino acids 1 to approximately 149; and						
		3) an inhibitory subportion of (b.1) and						
		(b.2);						
15	c)	a portion of the ICE gene which encodes an						
		amino acid sequence of ICE shown in Figures						
	•	6A-B (SEQ ID NO:4), selected from the group						
		consisting of:						
		 amino acids 1 to approximately 298; 						
20	**	2) amino acids 1 to approximately 111; and						
		3) an inhibitory subportion of (c.1) and						
		c.2);						
	đ)	a portion of said ced-3/ICE gene which encodes						
		an amino acid sequence corresponding to a						
25		portion of the Ced-3 protein shown in Figures						
		6A-B (SEQ ID NO:2), said Ced-3 portion						
		selected from the group consisting of:						
		1) amino acids 1 to approximately 372;						
		2) amino acids 1 to approximately 149; and						
30		3) an inhibitory subportion of (d.1) and						

(d.2); and

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- e) a portion of a ced-3/ICE gene other than said ced-3/ICE gene which encodes an amino acid sequence corresponding to a portion of the Ced-3 protein shown in Figures 6A-B (SEQ ID NO:2), said Ced-3 portion selected from the group consisting of:
 - 1) amino acids 1 to approximately 372;
 - 2) amino acids 1 to approximately 149; and
 - 3) an inhibitory subportion of (e.1) and
 (e.2).
- 43. An inhibitor of the activity of a gene belonging to the ced-3/ICE family of structurally related genes, comprising RNA encoded by the DNA of Claim 42.

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- 44. An inhibitor of the activity of a gene belonging to the ced-3/ICE family of structurally related genes, comprising protein encoded by the DNA of Claim 42.
- 45. An inhibitor of the activity of a gene belonging to the ced-3/ICE family of structurally related genes, comprising a non-peptide mimetic of the protein of Claim 44.
 - 46. A drug for reducing cell deaths, comprising an inhibitor of the activity of the ced-3 gene, selected from the group consisting of:
 - a) a portion of the ced-3 gene;
- b) a product encoded by a portion of
 the ced-3 gene;
 - c) a non-peptide mimetic of an inhibitory portion of the Ced-3 protein;
 - d) a portion of the ICE gene;

e)	a product encoded by a portion of	f
	the ICE gene;	

- f) a non-peptide mimetic of an inhibitory portion of the ICE protein;
- 5 g) a portion of a gene which is structurally related to the ced-3 gene;
 - h) a product encoded by the gene portion of (g); and
- i) a non-peptide mimetic of the protein encodedby the gene portion of (g).
 - 47. The drug of Claim 46, wherein the inhibitor is selected from the group consisting of:
 - a) DNA having a nucleotide sequence of Figure 3 (SEQ ID NO:1), selected from the group consisting of:
 - 1) nucleotides 1 to approximately 5850;
 - 2) nucleotides 1 to approximately 3020; and
 - 3) an inhibitory portion of (a.1) and (a.2);
 - b) DNA encoding an amino acid sequence of the Ced-3 protein shown in Figures 6A-B (SEQ ID NO:2), selected from:
 - 1) amino acids 1 to approximately 372;
 - 2) amino acids 1 to approximately 149; and
 - 3) an inhibitory portion of (b.1) and (b.2);
- 25 c) RNA encoded by DNA of (a);
 - d) RNA encoded by DNA of (b);
 - e) protein having an amino acid sequence of the Ced-3 protein shown in Figures 6A-B (SEQ ID NO:2), selected from the group consisting of:
- 30 1) amino acids 1 to approximately 372;
 - 2) amino acids 1 to approximately 149; and

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- 3) inhibitory portion of (e.1) and (e.2);
 and
- f) a non-peptide mimetic of the protein of e).
- 48. The drug of Claim 46, wherein the inhibitor is selected from the group consisting of:

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- a) DNA encoding an amino acid sequence of ICE shown in Figures 6A-B (SEQ ID NO:4), selected from the group consisting of:
 - 1) amino acids 1 to approximately 298;
 - 2) amino acids 1 to approximately 111; and
 - 3) an inhibitory portion of (a.1) and (a.2);
 - b) RNA encoded by DNA of a);
- c) protein having an amino acid sequence of ICE shown in Figures 6A-B (SEQ ID NO:4), selected from the group consisting of:
 - 1) amino acids 1 to approximately 298;
 - 2) amino acids 1 to approximately 111; and
 - an inhibitory portion of (c.1) and (c.2);
 and
- 20 d) a non-peptide mimetic of the protein of c).
 - 49. The drug of Claim 46, wherein the inhibitor is selected from the group consisting of:
 - a) protein encoded by a portion of a gene which is structurally related to the ced-3 gene, said protein portion corresponding to an amino acid sequence of the Ced-3 protein shown in Figures 6A-B (SEQ ID NO:2) selected from the group consisting of:
 - 1) amino acids 1 to approximately 372;
 - amino acids 1 to approximately 149; and
 - 3) an inhibitory portion of (a.1) and (a.2);

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- b) DNA encoding the protein of (a) or inhibitory subportion thereof;
- c) RNA encoding the protein of (a) or inhibitory subportion thereof; and
- 5 d) a non-peptide mimetic of the protein of (a).
 - 50. An inhibitor as defined in Claim 46, for use in therapy e.g. for treating a condition characterized by cell deaths.
- 51. A method for treating a condition characterized by cell deaths, comprising administering the drug of Claim 46.
 - 52. A drug for reducing cell deaths, comprising an inhibitor of the activity of the ICE gene, selected from the group consisting of:
- 15 a) a portion of the ICE gene;
 - b) a product encoded by a portion of the ICE gene;
 - c) a non-peptide mimetic of an inhibitory portion of the ICE protein;
- 20 d) a portion of the ced-3 gene;
 - e) a product encoded by a portion of the ced-3 gene;
 - f) a non-peptide mimetic of an inhibitory portion of the Ced-3 protein;
- 25 g) a portion of a gene which is structurally related to the ced-3 gene and the ICE gene;
 - h) a product encoded by the gene portion of (e); and
- i) a non-peptide mimetic of the protein encoded30 by (g).

- 53. A drug for reducing cell deaths, comprising an inhibitor of the activity of a gene which is structurally related to the ced-3 gene and ICE gene, selected from the group consisting of:
- 5 a) a portion of said related gene;
 - b) a product encoded by the gene portion of (e);
 - c) a non-peptide mimetic of the protein product encoded by (a);
 - d) a portion of the ICE gene;
 - e) a product encoded by the gene portion of (d);
 - f) a non-peptide mimetic of a protein product
 encoded by (d);
 - g) a portion of the ced-3 gene;
 - h) a product encoded by the gene portion of (g); and
 - i) a non-peptide mimetic of the protein product encoded by (g).
- 54. A drug for inhibiting the activity of a gene selected from the group consisting of ced-3 and a
 20 gene which belongs to the ced-3/ICE gene family, comprising an inhibitor of interleukin-1β convertase.
 - 55. The drug of Claim 54 which reduces cell deaths.
- 56. The drug of Claim 54 which is a peptide aldehyde

 25 containing the amino acid sequence Tyr-Val-X-Asp,

 wherein X is selected from Ala, His, Gln, Lys, Phe,

 Cha, and Asp.
 - 57. The drug of Claim 54 which is Ac-Tyr-Val-Ala-Asp-CHO, also referred to as inhibitor B.

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- 58. The drug of Claim 54 which is the cowpox virus CrmA protein.
- 59. An anti-inflammatory drug, comprising an inhibitor of the activity of the ICE gene, or inhibitory portion thereof, selected from the group consisting of:
 - a) a portion of the ICE gene;
 - b) a product encoded by a portion of the ICE gene;
- c) a portion of the ced-3 gene;

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- d) a product encoded by a portion of the ced-3 gene;
- e) a portion of a gene which is structurally related to the ced-3 gene and ICE gene; and
- 15 f) a product encoded by a portion of a gene which is structurally related to the ced-3 gene and the ICE gene.
 - 60. The anti-inflammatory drug of Claim 59, wherein the inhibitor is selected from the group consisting of:
- 20 a) DNA encoding an amino acid sequence of ICE shown in Figures 6A-B (SEQ ID NO:4), selected from the group consisting of:
 - 1) amino acids 1 to approximately 298;
 - 2) amino acids 1 to approximately 111; and
 - 3) an inhibitory portion of (a.1) and (a.2);
 - b) RNA encoded by DNA of (a) or an inhibitory subportion thereof;
 - c) protein having an amino acid sequence of ICE shown in Figures 6A-B (SEQ ID NO:4), selected from the group consisting of:
 - 1) amino acids 1 to approximately 298;

- 2) amino acids 1 to approximately 111; and
- 3) an inhibitory portion of (c.1) and (c.2);
- d) a non-peptide mimetic of the protein of (c).
- 61. The anti-inflammatory drug of Claim 59, wherein the inhibitor is selected from the group consisting of:
 - a) DNA having a nucleotide sequence of Figure 3 (SEQ ID NO:1), selected from the group consisting of:
 - nucleotides 1 to approximately 5850;
 - 2) nucleotides 1 to approximately 3020; and
 - 3) an inhibitory portion of (a.1) and (a.2);
 - b) DNA encoding an amino acid sequence of the Ced-3 protein shown in Figures 6A-B (SEQ ID NO:2), selected from the group consisting of:
 - amino acids 1 to approximately 372;
 - 2) amino acids 1 to approximately 149; and
 - 3) an inhibitory portion of (b.1) and (b.2);
 - c) RNA encoded by DNA of (a);
 - d) RNA encoded by DNA of (b);
- e) protein having an amino acid sequence of the Ced-3 protein shown in Figures 6A-B (SEQ ID NO:2), selected from the group consisting of:
 - 1) amino acids 1 to approximately 372;
 - 2) amino acids 1 to approximately 149; and
- 25 3) an inhibitory portion of (e.1) and (e.2); and
 - f) a non-peptide mimetic of the protein of (e).
 - 62. The anti-inflammatory drug of Claim 59, wherein the inhibitor is selected from the group consisting of:

- a) protein encoded by a portion of a gene which is structurally related to the ced-3 and ICE genes, said protein portion corresponding to an amino acid sequence of ICE shown in Figures 6A-B (SEQ ID NO:4), selected from the group consisting of:
 - 1) amino acids 1 to approximately 298;
 - 2) amino acids 1 to approximately 111; and
 - 3) an inhibitory portion of (a.1) and (a.2);
- 10 b) DNA encoding the protein of (a);
 - c) RNA encoding the protein of (a); and
 - d) a non-peptide mimetic of the protein of (a).
 - 63. A method for treating inflammation, comprising administering the drug of Claim 59.
- 15 64. An inhibitor or inhibitory portion as defined in Claim 59, for use in therapy e.g. for treating inflammation.
- 65. A method for altering the occurrence of cell death, comprising altering the activity of a cell death gene which is structurally related to ced-3.
 - 66. The method of Claim 65, wherein the structurally related gene is ICE.
 - 67. A drug for increasing cell deaths, comprising a molecule, of active portion thereof, selected from:
- 25 a) DNA comprising a gene which belongs to the ced-3/ICE gene family;
 - b) RNA encoded by the DNA of (a);
 - c) protein encoded by the DNA of (a);

- d) an agent which is structurally similar to and mimics the activity of the protein of (c);
- e) an agonist of the activity of a gene which belongs to the ced-3/ICE gene family;
- 5 f) DNA comprising a constitutively activated mutated form of a gene which belongs to the ced-3/ICE gene family;
 - g) RNA encoded by the DNA of (e);
 - h) protein encoded by the DNA of (e);
- i) an agent which is structurally similar to and mimics the activity of a protein encoded by the DNA of (e); and
 - j) an agonist of the activity of a constitutively activated mutated form of a gene which belongs to the ced-3/ICE gene family.
 - 68. The drug of Claim 67, wherein the gene which belongs to the ced-3/ICE gene family is ICE.

- 69. The drug of Claim 67(f), wherein the constitutively activated mutated form of the gene which belongs to the ced-3/ICE gene family encodes a carboxylterminal portion of a protein product of the wild-type gene, said carboxyl-terminal portion having a deletion of an amino-terminal portion which corresponds to an amino acid sequence of the Ced-3 protein shown in Figures 6A-B (SEQ ID NO:2), selected from the group consisting of:
 - a) amino acids 1 to approximately 372;
 - b) amino acids 1 to approximately 149; and
 - c) an inhibitory subportion of (a) and (b).

- 70. The drug of Claim 69, wherein the protein product of the wild-type gene has sequences corresponding to the autocleavage sites of ICE and the protein product of the wild-type gene is selected from the group consisting of:
 - a) the uncleaved form of the protein product; and
 - b) the subunits corresponding to the active subunits of ICE.
- 71. A method for reducing the proliferative capacity or size of a population of cells, comprising contacting the cells with the drug of Claim 67 under conditions suitable for activity of the drug.
 - 72. The method of Claim 71, wherein the population of cells is selected from the group consisting of:
- a) cancerous cells;
 - b) cells which produce autoreactive antibodies;
 - c) infected cells;
 - d) hair follicle cells;
 - e) cells which are critical to the life of a parasite;
 - f) cells which are critical to the life of a pest; and
 - g) cells which are critical to the life of a recombinant organism.
- 25 73. A molecule or active portions thereof as defined in Claim 67, for use in therapy e.g. for reducing the proliferative capacity or size of population of cells, selected for example from the group consisting of:
- 30 a) cancerous cells;

-92-

- b) cells which produce autoreactive antibodies;
- c) infected cells;

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- d) hair follicle cells;
- e) cells which are critical to the life of a parasite;
- f) cells which are critical to the life of a pest; and
- g) cells which are critical to the life of a recombinant organism.
- 10 74. A drug for decreasing cell deaths comprising a molecule selected from the group consisting of:
 - a) single stranded nucleic acid having all or a portion of the antisense sequence of a gene which is structurally related to ced-3, said nucleic acid which is complementary to the mRNA of the gene;
 - b) DNA which directs the expression of (a);
 - c) a mutated form of a gene which is structurally related to ced-3, does not cause cell death and antagonizes the activity of the wild-type gene; and
 - d) an antagonist of the activity of a gene which is structurally related to ced-3.
- 75. The drug of Claim 74, wherein the structurally related gene is ICE.
 - 76. A molecule as defined in Claim 74, for use in therapy e.g. for treating, in a human or other animal, a condition characterized by cell deaths, and wherein for example the condition is selected from the group consisting of:

- a) myocardial infarction;
- b) stroke;
- c) degenerative disease;
- d) traumatic brain injury;
- 5 e) hypoxia;
 - f) pathogenic infection; and
 - g) hair loss.
- 77. A method for treating, in a human or other animal, a condition characterized by cell deaths,
- comprising administering the drug of Claim 74 to the human or other animal under conditions suitable for activity of the drug.
 - 78. The method of Claim 77, wherein the condition is selected from the group consisting of:
- a) myocardial infarction;
 - b) stroke;
 - c) degenerative disease;
 - d) traumatic brain injury;
 - e) hypoxia;
- 20 f) pathogenic infection; and
 - g) hair loss.
 - 79. A constitutively activated cell death protein comprising an amino acid sequence of the Ced-3 protein shown in Figures 6A-B (SEQ ID NO:2),
- selected from the group consisting of:
 - a) the amino acids from approximately 150 to 503;
 - b) the amino acids from approximately 373 to 503;
 - c) the amino acids from approximately 150 to 372;
 - d) (b) and (c) together;
- e) an active subportion of (a), (b), and (c); and

-94-

f) combinations of these.

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- 80. The constitutively activated cell death protein of Claim 79, further comprising a subportion of the region of Ced-3 from amino acids 1 to 149, as shown in Figures 6A-B (SEQ ID NO:2), said subportion which enhances and does not inhibit the activity of the protein.
- 81. Isolated nucleic acid encoding the protein of Claim 79.
- 10 82. A constitutively activated cell death protein having an amino acid sequence of ICE shown in Figures 6A-B (SEQ ID NO:4), selected from the group consisting of:
 - a) the amino acids from approximately 111 to 404;
 - b) the amino acids from approximately 298 to 404;
 - c) the amino acids from approximately 111 to 297;
 - d) (b) and (c) together;
 - e) an active subportion of (a), (b), and (c); and
 - f) combinations of these.
- 20 83. Isolated nucleic acid encoding the protein of Claim 82.
 - 84. A drug for increasing cell deaths, comprising a molecule selected from the protein of Claim 79 or a nucleic acid encoding said protein.
- 25 85. A drug for increasing cell deaths, comprising a molecule selected from the protein of Claim 79 or a nucleic acid encoding said protein.

- 86. Isolated protein having cell death activity and the amino acid sequence of the NEDD-2 protein shown in Figure 7 (SEQ ID NO:13), or an active portion thereof.
- 5 87. Isolated nucleic acid encoding the protein of Claim 86.
 - 88. An isolated substrate-specific protease having the amino acid sequence of the Ced-3 protein shown in Figures 6A-B (SEQ ID NO:2).
- 10 89. An isolated substrate-specific protease, consisting essentially of a protein product of a gene which is structurally related to the ced-3 and ICE genes.
 - 90. The protease of Claim 89 which cleaves after aspartate residues.
- 15 91. The protease of Claim 89 which is a cysteine protease.
 - 92. Isolated ICE having an alteration which reduces the activity of the enzyme, the alteration selected from the group consisting of:
- 20 a) L to F at amino acid 26;
 - b) G to R at amino acid 65;
 - c) G to S at amino acid 287;
 - d) E to termination at amino acid 324;
 - e) W to termination at amino acid 340;
- 25 f) A to V at amino acid 361;
 - g) E to K at amino acid 390; and
 - h) T to F at amino acid 393.

- 93. Isolated DNA which is a mutated ICE gene encoding the altered enzyme of Claim 92.
- 94. Isolated RNA encoded by the DNA of Claim 93.
- 95. An isolated gene belonging to the ced-3/ICE family of structurally related genes which has a mutation which reduces the activity of the gene, said mutation resulting in an amino acid alteration corresponding to an amino acid alteration of the Ced-3 protein which inactivates the Ced-3 protein.
- 10 96. A product of the gene of Claim 95 selected from RNA and protein.
 - 97. Isolated protein which is the NEDD-2 protein having an alteration which inactivates the protein, said alteration selected from the group consisting of:
- a) A to V at amino acid 117;
 - b) E to K at amino acid 483; and
 - c) S to F at amino acid 486.
 - 98. Isolated nucleic acid encoding the protein of Claim 97.
- 20 99. Isolated protein which is structurally similar to Ced-3 and has an alteration at a conserved amino acid corresponding to an amino acid of the Ced-3 protein selected from the group consisting of:
 - a) Ser 183;
- 25 b) Met 234;
 - c) Arg 242;
 - d) Leu 246;

```
e) Ile 247;
           f) Ile 248;
           g) Asn 250;
           h) Phe 253;
           i) Arg 259;
5
           j) Gly 261;
           k) Asp 265;
           1) Gly 277;
           m) Tyr 278;
           n) Val 280;
10
           o) Lys 283;
           p) Asn 285;
           q) Leu 286;
           r) Thr 287;
15
           s) Met 291;
           t) Phe 298;
           u) His 304;
           v) Asp 306;
           w) Ser 307;
           x) Leu 310;
20
           y) Val 311;
           z) Ser 314;
          aa) His 315;
          .bb) Gly 316;
          cc) Ile 321;
25
          dd) Gly 323;
          ee) Ile 334;
          ff) Asn 339;
          gg) Pro 344;
30
          hh) Leu 346;
          ii) Lys 349;
          jj) Pro 350;
          kk) Lys 351;
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-98-

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11) Gln 356;
          mm) Ala 357;
          nn) Cys 358;
          oo) Arg 359;
5
          pp) Gly 360;
          qq) Asp 371;
          rr) Asp 414;
          ss) Arg 429;
          tt) Gly 434;
        · uu) Ser 435;
10
          vv) Ile 438;
          ww) Ala 449;
          xx) Val 452;
          yy) Leu 488;
15
          aa) Tyr 493; and
         aaa) Pro 496.
```

- 100. Isolated nucleic acid encoding the protein of Claim 99.
- 101. A method for identifying a portion of the ced-3

 gene which inhibits the activity of the ced-3

 gene, comprising the steps of:
 - a) injecting wild-type nematodes with a portion of the ced-3 gene under conditions suitable for expression of said gene portion; and
- 25 b) detecting a decrease in programmed cell deaths,

whereby a decrease in programmed cell deaths is indicative of a portion of the ced-3 gene which inhibits the activity of said gene.

- 102. Isolated DNA comprising the inhibitory portion of the ced-3 gene identified by the method of Claim 101.
- 103. A method for identifying a portion of a gene which is structurally related to the ced-3 gene which inhibits the activity of the ced-3 gene, comprising the steps of:
 - a) injecting wild-type nematodes with a portion of the structurally related gene under conditions suitable for expression of said gene portion; and
 - b) detecting a decrease in programmed cell deaths.

- whereby a decrease in programmed cell deaths is indicative of a portion of the structurally related gene which inhibits the activity of the ced-3 gene.
 - 104. Isolated DNA comprising the inhibitory portion of the structurally related gene identified by the method of Claim 103.
- 20 105. The DNA of Claim 104, wherein the structurally related gene is the ICE gene.
 - 106. A method for identifying a portion of ICE which inhibits the activity of said enzyme, comprising the steps of:
- 25 a) combining a portion of ICE with ICE and a substrate of ICE under conditions suitable for cleavage of the substrate by ICE; and
 - b) detecting a decrease in cleavage of the substrate.

-100-

whereby a decrease in cleavage of the substrate is indicative of a portion of ICE which inhibits the activity of said enzyme.

- 107. An isolated inhibitory portion of the ICE proteinidentified by the method of Claim 106.
 - 108. Isolated nucleic acid encoding the inhibitory portion of Claim 107.
- 109. A method for identifying a portion of the protein product of a gene which is structurally related to the ced-3 and ICE genes, and inhibits the activity of ICE, comprising the steps of:
 - a) combining a portion of the protein product of a gene which is structurally related to the ced-3 and ICE genes with ICE and a substrate of ICE under conditions suitable for cleavage of the substrate by ICE; and
 - detecting a decrease in cleavage of the substrate,

- whereby a decrease in cleavage of the substrate is indicative of a portion of the protein product of a gene which is structurally related to the ced-3 and ICE genes and inhibits the activity of ICE.
 - 110. An isolated inhibitory portion identified by the method of Claim 109.
- 25 111. Isolated nucleic acid encoding the inhibitory portion of Claim 110.

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- 112. A method for identifying a gene which is structurally related to the ced-3 gene and the ICE gene, comprising detecting a gene with:
 - a) a probe derived from the ced-3 gene or a product encoded by the ced-3 gene; and
 - b) a probe derived from the ICE gene or a product encoded by the ICE gene.
- 113. An isolated gene identified by the method of Claim 112.

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- 10 114. A method for identifying a gene which belongs to the ced-3/ICE family of structurally related genes, comprising detecting a gene with a probe selected from the group consisting of:
- a) a probe derived from a gene which is structurally related to the ced-3 gene and the ICE gene; and
 - b) a probe derived from the consensus sequence of a conserved region in genes belonging to the ced-3/ICE gene family.
 - 115. An isolated gene identified by the method of Claim-114 which has an activity selected from cell death activity and protease activity.
- 116. Isolated DNA selected from the group consisting of:

 25
 a) a region of a gene belonging to the ced-3/ICE family of structurally related genes which is

conserved among two or more family members; and

b) the consensus sequence of a conserved region in genes belonging to the ced-3/ICE gene family,

or encoded product thereof.

- 5 117. A method for identifying a gene which interacts with a ced-3/ICE gene belonging to said family, comprising identifying a mutation which enhances or suppresses the activity of a ced-3/ICE gene in a nematode, wherein the enhancing or suppressing mutation is indicative of a gene which interacts with the ced-3/ICE gene.
 - 118. The method of Claim 117, wherein the ced-3/ICE gene is selected from the group consisting of:
 - a) a wild-type ced-3 gene;
- b) a mutated ced-3 gene, the nematode being a mutant nematode;
 - c) a transgene which is a wild-type form of said ced-3/ICE gene, the nematode being a transgenic nematode having an inactivated endogenous ced-3 gene; and
 - d) a transgene which is a mutated form of said ced-3/ICE gene, the nematode being a transgenic nematode having an inactivated endogenous ced-3 gene.
- 25 119. An isolated gene identified by the method of Claim 117, or an encoded product thereof.

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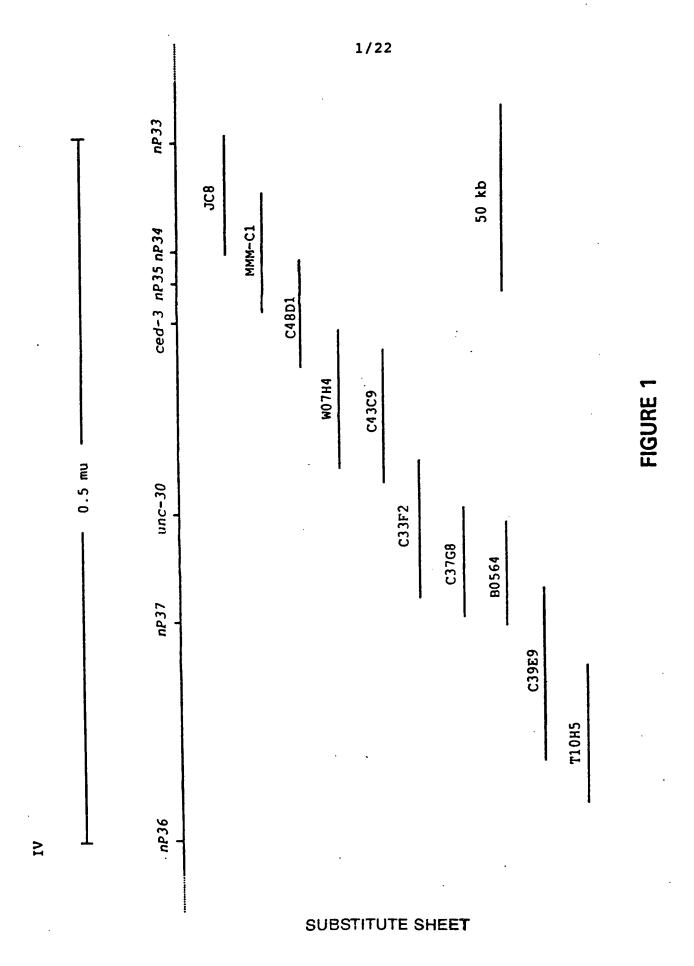
- 120. A bioassay for identifying an agent which affects the activity of a gene belonging to the ced-3/ICE family of structurally related genes, comprising the steps of:
- a) introducing an agent into a transgenic
 nematode which expresses a ced-3/ICE gene; and
 - b) detecting an alteration in the occurrence of cell deaths in the transgenic nematode, wherein an alteration indicates that the agent affects the activity of the ced-3/ICE gene.
 - 121. The method of Claim 120, wherein the ced-3/ICE gene is selected from a wild-type gene and a mutated gene.
 - 122. An agent identified by the method of Claim 120.
- 15 123. A diagnostic probe for a disease characterized by cell deaths, comprising a molecule selected from the group consisting of:
 - a) all or a portion of the ced-3 gene (SEQ ID NO:1) which is specific to said ced-3 gene;
- 20 b) RNA encoded by the ced-3 gene;
 - c) degenerate oligonucleotides derived from the amino acid sequence of the Ced-3 protein (SEQ ID NO:2);
 - d) an antibody directed against the Ced-3 protein;
 - e) all or a portion of the ICE gene (SEQ ID NO:3) which is specific to said ICE gene;
 - f) RNA encoded by the ICE gene;
- g) degenerate oligonucleotides derived from the amino acid sequence of ICE (SEQ ID NO:4);

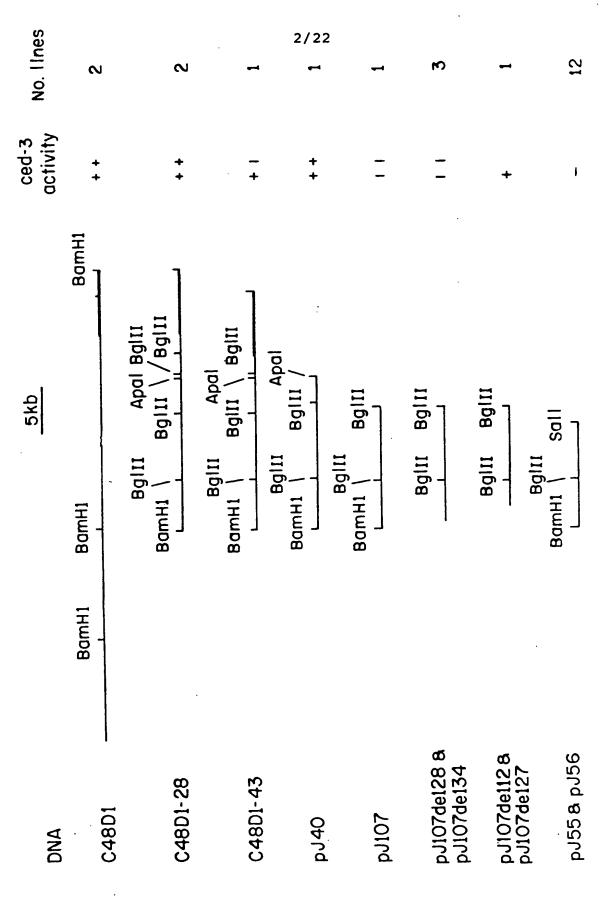
-104-

- h) an antibody directed against ICE;
- i) a gene which is structurally related to the ced-3 gene, or portion thereof specific to said structurally related gene;
- 5 j) RNA encoded by the structurally related gene;
 - k) degenerate oligonucleotides derived from the amino acid sequence of the protein product of a gene which is structurally related to ced-3; and
- 10 d) an antibody directed against the protein product of a gene which is structurally related to ced-3.
 - 124. A method for diagnosis of a disease characterized by cell deaths, comprising detecting an abnormality in the sequence of a gene which is structurally related to ced-3.
 - 125. The method of Claim 124, wherein the structurally related gene is ICE.

- 126. A method for diagnosis of a disease characterized 20 by cell deaths, comprising detecting an abnormality in the activity of a gene which is structurally related to ced-3.
 - 127. The method of Claim 126, wherein the structurally related gene is ICE.
- 25 128. A diagnostic probe for an inflammatory disease, comprising a molecule selected from the group consisting of:

- a) all or a portion of the ced-3 gene shown in Figure 3 (SEQ ID NO:1) which is specific to the ced-3 gene;
- b) RNA encoded by (a);
- 5 c) degenerate oligonucleotides derived from the amino acid sequence of the Ced-3 protein as shown in Figures 6A-B (SEQ ID NO:2);
 - d) an antibody directed against the Ced-3 protein;
- e) a gene which is structurally related to the ced-3 and ICE genes, or portion thereof which is specific for said related gene;
 - f) RNA encoded by (a);
- g) degenerate oligonucleotides derived from the
 amino acid sequence of the protein encoded by
 (e); and
 - h) an antibody directed against the protein encoded by (e).
- 129. A method for diagnosis of an inflammatory disease,
 20 comprising detecting an abnormality in the sequence
 of a gene which is a member of the ced-3/ICE gene
 family.
 - 130. The method of Claim 129, wherein the gene is ced-3.
- 131. A method for diagnosis of an inflammatory disease,
 25 comprising detecting an abnormality in the activity
 of a gene which belongs to the ced-3/ICE gene
 family, or an encoded product thereof.
 - 132. The method of Claim 131, wherein the gene is ced-3.





SUBSTITUTE SHEET

3/22 ced-3 Genomic Sequence

	AGATCTGAAATAAGGTGATAAATTAATTAAGTGTATTTCTGAGGAAATTAA	60
1	TTTAGCACAATTAATCTTGTTTCAGAAAAAAAGTCCAGTTTTCTAGATTTTTCCGTCTTA	120
61	TATEL TO THE TOTAL TENT OF THE TENT OF THE TOTAL CONTROL OF THE TOTAL CO	180
121	A B B C B A TTGTGAGAGCAAA CGCGCTCCCATTGACCTCCACACTCAGCCGCCAAAA CAAA C	
181		240
241	TTTTCTTTGTTCTTTTTGTTGAACGTGTTGCTAAGCAATTATTACATCAATTGAAGAAAA	300
301	·	360
361	GGCTCGCCGATTTATTGTTGCCAGAAAGATTCTGAGATTCTCGAAGTCGATTTTATAATA	420
421	TTTAACCTTGGTTTTTGCATTGTTTCGTTTAAAAAAACCACTGTTTATGTGAAAAACGAT	480
481	TAGTTTACTAATAAAACTACTTTTAAACCTTTACCTTTACCTCACCGCTCCGTGTTCATG	540
	GCTCATAGATTTTCGATACTCAAATCCAAAAATAAATTTACGAGGGCAATTAATGTGAAA	600
541	CAAAAACAATCCTAAGATTTCCACATGTTTGACCTCTCCGGCACCTTCTTCCTTAGCCCC	660
601	ACCACTCCATCACCTCTTTGGCGGTGTTCTTCGAAACCCACTTAGGAAAGCAGTGTGTAT	720
661	THE TRUE TO THE TATA CONTINUE TO THE TAXABLE TO THE TAXABLE TO TAX	760
721	AATCCAAATCGCATTATATTTGTGCATGGAGGCAAATGACGGGGTTGGAATCTTAGATGA	840
781	CATCA CONCETT CACGGT A A CCCCCGGT CATTTTGT A CCACATTTCATCATTTTCCT	
841	CTCCTCCTATCCTCAACTTGTCCCGGTTTTGTTTTCGGTACACTCTTCCGTGATGC	900
901	CACCTETCTCCGTCTCAATTATCGTTTAGAAATGTGAACTGTCCAGATGGGTGACTCATA	960
961	TIGCTGCTGCTACAATCCACTTTCTTTTCTCATCGGCAGTCTTACGAGCCCATCATAAAC	1020
1021	TTTTTTTTCCGCGAAATTTGCAATAAACCGGCCAAAAACTTTCTCCAAATTGTTACGCAA	1080
1081	TATATACATCCATAAGAATATCTTCTCAATGTTTATGATTTCTTCGCAGCACTTTCTCT	1140
1141		1200
1201	TCGTGTGCTAACATCTTATTTTTATAATATTTCCGCTAAAATTCCGATTTTTGAGTATTA	1260
1261	ATTTATCGTAAAATTATCATAATAGCACCGAAAACTACTAAAAATGGTAAAAGCTCCTTT	132
	Repeat 1	
	TAAATCGGCTCGACATTATCGTATTAAGGAATCACAAAATTCTGAGAATGCGTACTGCGC	
1321		138
	AACATATTTGACGGCAAAATATCTCGTAGCGAAAACTACAGTAATTCTTTAAATGACTAC	144
138	Repeat 1	
	<pre></pre>	
	#CTNCCCCTTCTCTCCNTTTACGGGCTCAATTTTTGAAAATAATTTTTTTTTT	_
	101A00001101010000011100000110000000000	, 150

FIGURE 3(a)

	4/22			_	
	TGATAACCCGTAAATCGTCACAACGCTACAGTAGTCATTTAAAGGATTACTGT	AGI	TCT	- A +	1560
1501					
				_	
	GCTACGAGATATTTTGCGCGCCAAATATGACTGTAATACGCATTCTCTGAAT			-	1620
1561	TCCGTAATAATTTCACAAGATTTTGGCATTCCACTTTAAAGGCGCACAGGAT	TTA:	TTCC	А	1680
1621	ATGGGTCTCGGCACGCAAAAAGTTTGATAGACTTTTAAATTCTCCTTGCATT	TTI	LTA	C	1740
1681	**************************************	TA	ATA7	T	1800
1741	THE CONCENT OF THE CONTRACT OF	TTT		-	
1801	· · · · · · · · · · · · · · · · · · ·			•	1860
1861				•	1920
1921	GATTCAAAACAATTTTAACAAAAAAAAACCCCAAAATTCGCCAGAAATCAAG			•	1980
_	TTCAAGAGGGTCAAAATTTTCCGATTTTACTGACTTTCACCTTTTTTCGT			-+	2040
1981	GCAGTTGTTGGAGTTTTTGACGAAAACTAGGAAAAAAATCGATAAAAATTAC	TCA	AAT	CG -+	2100
2041	* PECTER ATTTCAGGACA ATGTTTAAAAAAAAAACACTATTTTTCCAATAATT	TCA	CTC	AT	2160
2101	1			•	
	TTTCAGACTAAATCGAAAATCAAATCGTACTCTGACTACGGGTCAGTAGAGA	GGI	CAA	СС	
				-+	2220
2161	ATCAGCCGAAGATGATGCGTCAAGATAGAAGGAGCTTGCTAGAGAGGAACAT	AT1	TGAT	GT	
2221]			- •	2280
	MMRQDRRSLLERNI 1 10	Ε,	**	•	
	T (n1040)				
	TCTCTAGTCATCTAAAAGTCGATGAAATTCTCGAAAGTTCTCATCGCAAAAC	AAG'	IGTI	GA +	2340
2281	S S H L K V D E I L E V L I A K Q	V	L	N	
	Lintron 1	TTN			
2341	ATAGTGATAATGGAGATATGATTAATGTGAGTTTTTAATCGAATAATAATT			+	2400
2011	SDNGDMIN 40				
	 AATTGATAATATAAAGAATATTTTTGCAGTCATGTGGAACGGTTCGCGAGA	AGA	GAC	GGG	
240				+	2460
	50	•		-	
	እ (ກ718) 1				
	AGATCGTGAAAGCAGTGCAACGACGGGGAGATGTGGCGTTTCGACGCGTTTT	ATC	ATG	CTC +	2520
246	IVKAVQRRGDVAFDAFI) A	L	
				tron	2
252	TTCGCTCTACGGGACACGAAGGACTTGCTGAAGTTCTTGAACCTCTCGCCA			4	2580
	RSTGHEGLAEVLEPLAI	? :	S		

FIGURE 3(b)

												5/	22								
T	ITI	TA	AA	GT	TC	GCC	CAJ	AAAGC	AAG	GGT	CT	CAC	GGZ	w	AA.	SAG	3CG(SAT(-+-	GT	AT:	TTT +
G	CAP	~~	~ »	~	·cc	-20	COT	TTTT	CCT	cco	:AA	AAT	CGG	SAAJ	TT	ATG	CAC	ITT	CCC	AAA.	TAT
_				-+				-+ Tatti			- + -				+ – – ·			-+			
_				4				_ +			-+-				+			-+-			+
A'	TT	TC	GT	`GG	CG	CAA	AAG	SCCAT	TTT	`GT	\GA -+-	TTI	rgc	CGA	√ΑΑ. +	TAC	TTG	TCA(CAC.	ACA	+
_				+							•				•						
_								TATCO			*^^	. N G 1	rcT'	TGA	CTC	C A A	TGC	TGT	CGA	ATT	CGA
A -	CAG	CAC	TA.	1	, CC			-+			-+-				+			-+-			+
													V	D	S	N	λ	V 10		F	E
_				4	٠			AAGC			-+-				+			-+-			+
	С	P	1	4	S	P	A	S 1	4 1	R 1	R	S	R	λ	L	S	P	12	0	Y	T
1	TC	ACC	:GZ	A C	CCG	AGT	TCA	CCGT	GAC	AGC	GT	CTC	TIC	AGT	GTC	ATC	ATT	CAC	TTC	TTA	TCA
-	 s	 P		· T	* R	v	H	R	D :	s ·	V	s	S	v	s	s	F	T	s	Y	Q
								130										14	0		
Ç	SG A	TA?	LC,	TA	стс	AAC	SAGC	AAGA	TCT	CGT	TC	TCG	ATC	:GCG	TGC	ACT	TCA	TTC	ATC	GGA	TCG
-					+	 D		-+ R	 s	 R	-+ S	 R	 s	R	+	ı	н	-+- S	s	D	R
	ט	1		•			•	1.50		•	_	•		•				16	0		
					+			P 170	 v		-+				+		TTC	int STAT	GT	GAT	GCG
					1	Rep	eat	1													
								ATGCG						2 ·			CON	TAA	ATC'	rcg'	TAGO
	AAC	CAC	TA		.+-		AGA	+			+				-+-			+			+
	GA)	raa 	A T	AC.	:AG	TAA 	ccc.	TTTA	VATO	AC.		TG	TAG	TG1	-+ <i>-</i>			+			
	:	>						ኢ አ ፕፕር				- 2 2	~~~	T 3 3	~~~	CTC	>- +	ጥ ጉጉ	CTC	ተ ተጉ	TOT
	AA	ACC	;A,	AT.	4TA -+-	TGC	TCG	+-				>AA +			-+-			+			
	TT 	GA1	[A]	TT'	ITT -+-	GAT	CXA	AATT.	TAA:	TTA	AT'	TTC +	CG1	AAA	-+-	ACA	CCA	.GCG +	CTA	CAG	TAC:
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	TG	ኢኢ	AA	AA	AAI	CA7	CTA	LACAT	GTG	CCA	AA	ACG	- -			WG1		.GCA	CAI	111	- 1 6

FIGURE 3(c)

	Repeat 2	
	THE THE PARTY OF THE PROPERTY	3540
3481		3540
	•	
	TAGAAATTTTGGGCTTTTCGTTCTAGTATGCTCTACTTTTGAAATTGCTCAACGAAAAA	
	TAGAAATTTTGGGCTTTTCGTTCTAGTATGCTCTAGTTTTGGGCTTTTCGTTTTGGGCTTTTCGTTCTAGTATGCTCTAGTTTTGGGCTTTTTGGGCTTTTTGGGCTTTTTGGGCTTTTTGGGCTTTTTGGGCTTTTTGGGCTTTTTGGGCTTTTTGGGCTTTTTGGGCTTTTTGGGCTTTTTGGGCTTTTTGGGCTTTTTGGGCTTTTTGGGCTTTTTGGGGCTTTTTGGGGCTTTTTGGGGCTTTTTGGGGCTTTTTGGGGCTTTTTGGGGCTTTTTGGGGGTTTTGGGTTTTGGGGTTTTGGGTTTTGGGG	3600
3541	******	
	TO TOTO TITO TO TATO A TOTAL TO A TOTAL A TOTA	3660
3601	1CA10100111011CA1A1010101010101010101010	3000
	TO THE TOTAL COLOR DE LA COLOR	
3661	TCATGTTGTGCAGAAAAATAGTAAAAAAGCCCAT	3720
3001		
	ACGACAGCTCACTTCACATGCTGAAGACGAGAGAGACGCGGGAGAAATACCACACATCTTTCT	
	ACGACAGCTCACTTCACATGCTGAAGACGAGAGACGCGGGAGGGTTTTTTTT	3780
3721		
	Repeat 2	
•		
	GCGTCTCTCGTCTTCAGCATGTGAAATGGGATCTCGGTCGATGTAAAAAAATGTCGAATA	3840
3781	GCG1C1CCG1C11CAGCA101GCGCGCGCGCGCGCGCGCGCGCGCGCGCGCGCGCGCG	3640
	A TOTA A A A A TOCA TOCOTT TITTIA CACTTITCTGCACAAATGAATAGGGGGGAAAATGT	
3841	Williamoratoriocontrational and a second sec	3900
•		
	ATTAAAATACATTTTTTGTATTTTTCAACATCACATGATTAACCCCATTATTTTTTCGTT	
3901	ATTRACATACATTTTTTTTTTTTTTTTTTTTTTTTTTTT	3960
3901		
	GAGCAACTTAAAAGTAGAGAATATTAGAGCGAAAACCAAAATTTCTTCAAGATATTACC	4020
3961		
	TITATTGATAATTATAGATGTTAATAAGCATATCTTGAATGAA	4083
4021	GCGAAACACCTGAAAAAATCAAAAATTCTGCGAAAATTGAAAAAATGCATTAAAATACA	400.
	·	4140
4081	TTTTTGCATTTTTCTACATCACATGAATGTAGAAAATTAAAAGGGAAATCAAAATTTCTA	
4141		420
	CA CONTATA A TTOA A TOA A A CATTOCOA A A TTAA A A TOTOCOA A A A A A A A A A A A A A A A A A A	135
4201	0.000.1.1.1.1.1.1.1.1.1.1.1.1.1.1.1.1.1	426
	1	
	AATTTGGGTATCAAAATCGATCCTAAAACCAACACTTTCAGCATCCGCCAACTCTTCAT	
4263		432
4201	SANSSF	
	180	

FIGURE 3(d)

											7/.					_					
	TCAC	CGG	ATG	CTC	TTC	TCT	CGG	ATA	CAG'	TTC	AAG	TCG	TAA	TCG	272	ATT	CAG	CAA	AGC	11	4380
4321	T	G	+ C	s	5 19	_	G	Y	S	-+- \$	s	R	N	Ŗ	s 20		5	K	λ	S	4300
	CTGG.	ACC	AAC	TCA	ATA	CAT	ATT	CCA	ΤGλ	AGA	GGA	TAT	GAA	C. T	1G:	CGA	TGC	ACC	λλC	CA	
4381			+				+			-+-			+				+			-+	4440
	.G	P	T	6	Y 21	O	F	H	E	E	D	M	N		22		^	•	•	•	
	TAAG	ccc	TGT	TTI	CGA	CGA	GAA	AAC	CAT	GTA	CAG	AAA	CTI	CTC	GAS	tcc	TCG	TGG	XXT	GT	4500
4441	\$	R	v	F	D 23	E	+ K	T	H	-+- Y	R	N	F	S	5 24		R	G	М	c	4500
	GCCT	CA7	rcat	:AA	(ATA	TGA	ACA	CTI	TGA	.GC#	GAT	GCC	AAC	ACG	GXX	TGG	TAC	CAA	GGC	:CG	
4501							+			-+-			+ T				+			-+	4560
	L	I	1	N		50	n	2	L	v	r,	•	•	•	26		•	••	••		
	ACAP	\GG/	ACAJ	ATC:	AT1	CAA	TTI	GTI	CAC	ATC	CA1	rGG(CTA	TAC	GGT	TAT	TIC	CN	GGA	CA -+	4620
4561	K	D	N	L	_	N 70	L	F	R	c	M	G	Y	7	V 26		С	K	D	N	
	ATC				1 :	inti	on	4			-						rcc			:ca	
4621	ATC	rga:		3AA:		TACC											-+			+	4680
	ı	Ţ	G	R																	
				P.o.																	
				Re									>						- >		
4681	CCG					CCG		rcg/	ACA:	CGA:	CAA	:II	GIG	17 <i>6</i>) +	W		~~~.			+	4740
4001	TTT	GCA	AAA	AAC	AAA	ATT:	TTG	AAC'	TTC	CGC	GAA	AAT	GAT'	TTA:	CT	NGT'	TTC	SAN	KIT.	TTC	4800
4741	GII	 	 TCC	4	TAC	ATT	-+- ATG	TGT	TTT	+ TTC	TTA	GTT	TIT	t CTA:	. AA		TTG	ATG	. AA	NAA	4600
4801				+			-+-			+				+			-+-			+	4860
4063	ACC	GTT 	TGT	888 	TTT	TCA	GAC.	AAT'	777	CCG +	CAT	АСА 	AAA 	CTI:	3A7	AGC.	AC 0			+	4920
4861	TTC	TGA	TTA	TTC	AAA	ATT.	ATC	CAA										TTG	GCA	AAC	
4921		CTT		4	TGA	AAT	-+- GTA	 TTT					AAA					AAG	CAN.	TAA	4980
4981				+			-+-			+				+			-+-			+	5040
5041	AAA	TCA	AXA	.CAA	CGI	CAC							A11			A 	-4-			+	5100
3041	CAA	AAI	TTG	w	aa.	ATCA	TGA	AGG	TTA	TAG	AAA	AGT	TTT	ATA	ACA	:::	TIT	CTA	GAT	TTT	6361
5101	TC3		 \TTT	·+ `TTI	TTZ	ACA	-+- Aat	CGA	GAA	AAA	GAG	AAT	'GAA	* ***	TCG	λ 	TTA	አጹኢ	ATA	TCC	5160
5161				+			-+-			+				+			-+-			+	5220
			pes																		
	AC1	\GC1	TCC	:AGJ	AGT	TGA	AAT	TAC	AGT	'ACI	CCI	TAP	LAGG	ccc	ACA	ccc	CAT	TIG	CAT	TGG	
5221				-+			-+-							+			-4-			+	528
	200			 	ane:	GTGI	CG	GAC	CAC	GGT/	ACC	STAC	JTT1	TTG		 ::\.		.770	CAC	CAT	
5283	1			-4			4-				+			-+			-+-				534
•	TG	GAC	LAAT	NA.	CCI,	TCCI	TAAT	CAC	CAJ	AAA	AGT	WW	ATTO	دىد	.TC	TC	JAAJ	J.GC	CA	AAA	

FIGURE 3(e)

											8/	22						A A C	- 2 2 1	4	
	TTCA																				5460
,	LAATO	:AAT	TTT	CTO	CAA	AAT	ACC		AAG	:XXI	ACC	CGA	W	TKA	TTC	CCA	\$CC'	TTG:	TTC	-+	5520
1	LATG:	raa:	-		TTI	TAA1	TTC	l CAG	GG?)TA						A GA	CTT	TGC	CAN	AC -+	5580
			+-			+				M	L	L	T	I	R	D	F	λ	Ж 30		
	ACGA	ATC	ACA(cGG	AGA:	TTC	rgc	GATA	CT	CGT	GAT	TCT	ATC	λCλ	ccc	AGA	AGA	GAA	TGT	GA -+	5640
	E	S	H	G	D	S	λ	1	L 31		1	ı	s	н	G	E	E	N	V 32	0	
	TTAT	TGG	AGT	TGA	TGA	TAT	ACC	GAT?	rag	TAC	ХСХ	CGA	GAT	KTA'	TGA	TCI	TCI	CYY	ccc	GG -+	5700
	<u>-</u>		+ V	D	D	1	+ P	1	s 33		н	E	I	¥	D	L	L	N	λ 34	A 0	
														r a c'	raci	, cc	-TT	STC	1	1	33) intron !
	CAAA																				57€0
	N	λ		R	L	λ	N	K	9 35	, K	1	V		V	V	•		•	30	50	
	GTT	:GT		ΓΤΑ: •	TTT	AA7	TT:	TAAT	ATA		IAT	111 	AAA	TAA +	ATT	CAT	TTT -+-	l CAG		•	5820
				-										•					R	R	
	GTG	ACA	ATG	GAT	TCC					+				-							5880
1	D	N	G	F	P	ν		70	s	ν	D	• 6	V	F	· >	F	3	. R 80	R		
																				1	(n1165)
_	GAT	GGG	ACA	ATC	GAG	ACG	GGC	CAT	TGT	TCA	AT1			-+							5740
l	V) N	F	D	G	-	90 90	F	•	1	7 1	L (3 (٠ :	, 1		P (, ,	, 0	!
•	AG	STTC		XTT1				TAAE													
1	AT:	TCC	CAI	AAG	rccc	GAT	CG	AAAA	ATI	CCC	SAT	ATA	ATT.	ACG	***	TTT:	GTG +	ATA	AAA:	rgac	
•	•				4																•
61		ACC.	TAA	CAG-+-	CAT	CGT	CGA	TCTC	CGG	ccc	ACT +	TCA		GAT +-	TGG		+	AĢT			6120
						GGT		AGT:	TT	CAG	TTT	AGA	.GGC	AA T	TTA	AAA	ATC	GCC	TTT	TCG	A + 6180
121	AA		AAA	-+- Taa	TGA	777	+ TTT	CAA'	TTT	TTI	CGA	AA	JAT!	111	CG	ATT.	TTI	TAT	ATT	CTT	

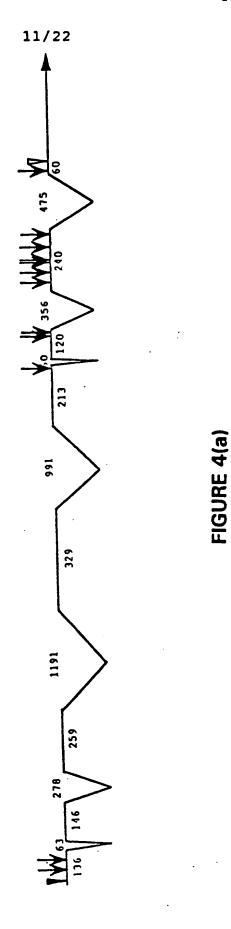
FIGURE 3(f)

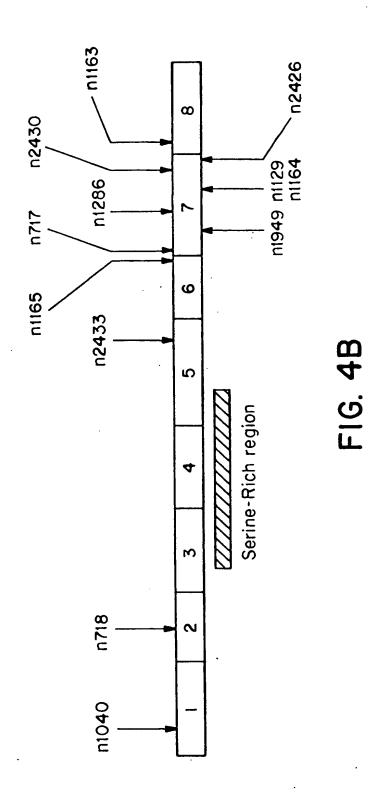
	9/22 A(n717)													
6241	GGAGCGAAAGCCCCGTCCTGTAAACATTTTTAAATGATAATTAAT	6300												
	T (n1949)													
6301	GTGTGGAGAAAGAAGCCGAGCCAAGCTGACATCTGATTCGATACGCAACGACAGCTCAA V W R K K P S Q A D I L I R Y A T T A Q	6360												
	410 420 A (n1286)													
6262	TATGTTTCGTGGAGAAACAGTGCTCGTGGATCATGGTTCATTCA	6420												
6361	Y V S W R N S A R G S W F I Q A V C E V 430 440													
	T (n1129, n1164) I TTCTCGACACGCAAAGGATATGGATGTTGTTGAGCTGCTGACTGA													
6421	F S T H A K D M D V V E L L T E V N K K	6480												
	T (n2430) (n2426) (introduced the control of t	on 7												
6481	V A C G F C T S Q G S N I L K Q M P E	6540												
	Repeat 5													
6541	CTTGANACANACANTGCATGTCTANCTTTTANGGACACAGANAAATAGGCAGAGGCTCCT	6600												
	TTTGCAAGCCTGCCGCGCTCAACCTAGAATTTTAGTTTTTAGCTAAAATGATTGAT													
6601		6660 6720												
6661	CCAGTAAAAATGTTTATTAGCCATTGGATTTTACTGAAAACGAAAATTTGTAGTTTTTC	678C												
6721 6781	AACGAAATTTATCGATTTTTAAATGTAAAAAAAAAATTACGCGAAAATTACATCAACCATCAA	6840												
6841	GCATTTAAGCCAAAATTGTTAACTCATTTAAAAATTAATT	6900												
	Repeat 5 ACACGGTTGGCGCGCGCAAGTTTGCAAAACGACGCTCCGCCTCTTTTTCTGTGCGGCTT													
6901		6960												
		n1163)												
696	GAAAACAAGGGATCGGTTTAGATTTTTCCCCAAAATTTAAATTAAATTTCAGATGACATC	7020												

FIGURE 3(g)

K K 490	F	Y	F	W	P	Ε	λ	R		_	λ	V	•	
TTAA	ACC	ATGT	GTA	TAT	TGI	TAT	CCI	ATA	CTC	TA	TC	CTI	TAT.	CATTCI
·+	CAT	+- rttc	:ACA	CAT	+ TTC	CAT	TIC	TCT	ACC	ATA	LATO	-+- :TAP	LAA7	TATGAC
TCGA	CGC	-+- Kata	TAP	TTI	אני	TA	CTC	GTI	+	AA?	TTC	AT7		+
TATAT	TAT	STAC	TAT	rGC1	TCI	ATC	:AAC	AAA:	ATA	GT	CIC	TAC	ATC	ATCACC
													ငလ	TTAATT
-+ TTTT	CGC	CAC)	LAAJ	VAA T	CTA	LATA	TT	CN	TT	AC	JAX:	RAGO	AT	CCCATC
cccci														
AGGTC		•												
-+		+-							+			+-		+
	K K 490 TCATT TTTAA TCTTCC TCGAI TATATC TCTTCT TCGAI TATATC TCGAI TATATC TCGAI TTTTT TCTCAI TCTTCC TCGAI TCTTCC TCGAI TCTTCC TCTCCAI TCTTCC TCTCC TCTCCAI TCTTCC TCTCC TCTC TCT TCTC TCTC TCT	K K F 490 TCATTGCCC TTTAAAACCI TCTTCCCATT TCGAACGCI TATATGTATC TTTTTTCGCC GCCGGAATGC	K K F Y 490 TCATTGCCCAAT TTAAAACCATGT TCTTCCCATTTTC TCGAACGCATAA TATATGTATGTAC TTTTTTCGCCACA TTTTTTCGCCACA GCCGGAATGCCTACGAACCCCCCCA	K K F Y F 490 TCATTGCCCAATTGA TTAAAACCATGTGTA TCTTCCCATTTTCACA TCGAACGCATAATAA TATATGTATGTACTAC TTTTTTCGCCACAAAA GCCGGAATGCCTCCCC	K K F Y F W 490 TCATTGCCCAATTGATAA TTAAAACCATGTGTATAT TCTTCCCATTTTCACACAT TCGAACGCATAATAATTT ACCAACCTACCGTACCATA TTTTTTCGCCACAAAAAAT GCCGGAATGCCTCCCGGCC	K K F Y F W P 490 TCATTGCCCAATTGATAATTG TTAAAACCATGTGTATATTGT TCTTCCCATTTTCACACATTTC TCGAACGCATAATAATTTTAA TATATGTATGTACTATGCTTCT ACCAACCTACCGTACCATATTC TTTTTTCGCCACAAAAAATCTA GCCGGAATGCCTCCCGGCCTTT	K K F Y F W P E 490 TCATTGCCCAATTGATAATTGTCT TTAAAACCATGTGTATATTGTTAT TCTTCCCATTTTCACACATTTCCAT TCGAACGCATAATAATTTTAATA TATATGTATGTACTACGCTTCTATC TTTTTTCGCCACAAAAAATCTAATA GCCGGAATGCCTCCCGGCCTTTTAA	K K F Y F W P E A 490 TCATTGCCCAATTGATAATTGTTATCCT TTTAAAACCATGTGTATATTGTTATCCT TCTTCCCATTTTCACACATTTCCATTTC TCGAACGCATAATAATTTTAATAACTC TATATGTATGTACTATCATCTTTTTT TTTTTTCGCCACAAAAAATCTAATATTT GCCGGAATGCCTCCCGGCCTTTTAAAGT	K K F Y F W P E À R 490 TCATTGCCCAATTGATAATTGTCTGTATCT TTAAAACCATGTGTATATTGTTATCCTATA TCTTCCCATTTTCACACATTTCCATTTCTCT TATATGTATGTACTATGCTTCTATCAACAAA ACCAACCTACCGTACCATATTCATTTTTGCC TTTTTTCGCCACAAAAAATCTAATATTTGAA GCCGGAATGCCTCCCGGCCTTTTAAAGTTCC	K K F Y F W P E A R N 490 50 TCATTGCCCAATTGATAATTGTCTGTATCTTCT TTAAAACCATGTGTATATTGTTATCCTATACTC TCTTCCCATTTTCACACATTTCCATTTCTCTACG TCGAACGCATAATAATTTTAATAACTCGTTTTC TATATGTATGTACTATGCTTCTATCAACAAAATA ACCAACCTACCGTACCATATTCATTTTTGCCGGC TTTTTTCGCCACAAAAAATCTAATATTTGAATTA GCCGGAATGCCTCCCGGCCTTTTAAAGTTCGGAAAAGGCCCCCCCC	K K F Y F W P E A R N S 490 500 TCATTGCCCAATTGATAATTGTTATCCTATACTCATT TCTTCCCATTTCACACATTTCCATTTCTCTACGATA TCTTCCCATTTTCACACATTTCCATTTCTCTACGATA TCGAACGCATAATAATTTTAATAACTCGTTTTGAAT TATATGTATGTACTATGCTTCTATCAACAAAATAGTT TCCAACCTACCGTACCATATTCATTTTTGCCGGGAAT TTTTTTCGCCACAAAAAATCTAATATTTGAATTAACG TCCGGAATGCCTCCCGGCCTTTTAAAGTTCGGAACA	K K F Y F W P E A R N S A 490 500 TCATTGCCCAATTGATAATTGTCTGTATCTTCTCCCCCA TTAAAACCATGTGTATATTGTTATCCTATACTCATTTCA TCTTCCCATTTCACACATTTCCATTTCTCTACGATAATC TCGAACGCATAATAATTTTAATAACTCGTTTTGAATTC TATATGTATGTACTATGCTTCTATCAACAAAATAGTTTCA TCCAACCTACCGTACCATATTCATTTTTTTTTCCCGGGAATCAA TTTTTTCGCCACAAAAAATCTAATATTTGAATTAACGAAT TCCGGGAATGCCTCCCGGCCTTTTAAAGTTCGGAACATTTC	K K F Y F W P E A R N S A V 490 500 TCATTGCCCAATTGATAATTGTCTGTATCTTCTCCCCCAGTT TTAAAACCATGTGTATATTGTTATCCTATACTCATTTCACTT TCTTCCCATTTTCACACATTTCCATTTCTCTACGATAATCTAA TCGAACGCATAATAATTTTAATAACTCGTTTTGAATTTGATT TATATGTATGTACTATGCTTCTATCAACAAAATAGTTTCATAC ACCAACCTACCGTACCATATTCATTTTTGCCGGGAATCAATTT TTTTTTCGCCACAAAAAATCTAATATTTGAATTAACGAATAGC GCCGGAATGCCTCCCGGCCTTTTAAAGTTCGGAACATTTGGCAAGGTCCCCCCCATCATTTCCCGCCCATCATCTCAAATTGCATT	TCATTGCCCAATTGATAATTGTCTGTATCTTCTCCCCCAGTTCTC TTAAAACCATGTGTATATTGTTATCCTATACTCATTTCACTTTATCTTCCCCCATTTTCACTTTATCTCTCCCATTTCACATTTCACTTTATCTCCCAACCATAATCTAAAATTTTAATAA

FIGURE 3(h)





SUBSTITUTE SHEET

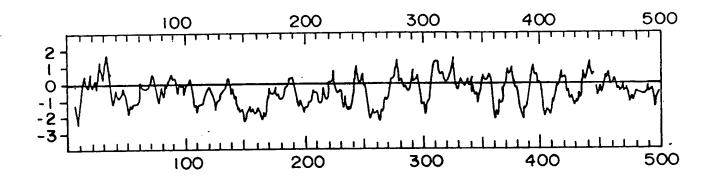


FIG. 5

Alignment of Ced-3 and Human Interleukin-1eta Convertase

ICE	1	MADKVLKEKRKLFIRSMGEGTINGLLDELLQTRVLNKEEMEKVKRE
Ced-3	1	.: :: . : .: :::: : : .:: :.:MMRQDRRSLLERNIMMFSSHLKVDEILEVLIAKQVLNSDNGDMIN.S
BGAFQ PBA		. F ====================================
	47 47	NATVMDKTRALIDSVIPKGAQACQ.ICITYICEEDSYLAGTLGLSADQTS : . : . : : CGTVREKRREIVKAVQRPGDVAFDAFYDALRSTGHEGLAEVLEPLARSVD
BFAFQ PBA		R ====================================
BGAFQ	96 97	autocleavage site GNYLNMQDSQGVLSSF : :::: :: SNAVEFECPMSPASHRRSRALSPAGYTSPTRVHRDSVSSVSSFTSYQDIY Berine-rich region
PBA		
	112 147	
BGAFQ PBA		====================================
	140 197	EAQRIWKQKSAEIYPIMDKSSRTRLAL: :: ::: ::: RNRSFSKASGPTQYIFHEEDMNFVDAPTISRVFDEKTMYRNFSSPRGMCL
BGAFO	131	ANASI SAASGI IQI I I REEDMNI VDAPI I SKVIDEKI MYRNI SSPRGMCL

FIGURE 6(a)

ICE	167	
Ced-3	247	: : . : . : . .
BGAFQ		
	217	AFAHRPEHKTSDSTFLVFMSHGIREGICGKKHSEQVPDI.LQLNAIFNML
	297	. .: : .: :. . . : :. ::: DFAKHESHGDSAILVILSHGEENVIIGVDDIPISTHEIYDLL
BGAFQ		######################################
:	266	The state of the s
	339	- - - : :::: :: ::::::::::
BGAFO		\$ ====================================
	315	- The control of the
•	389	::::::::::::::::::::::::::::::::::::::
		stop stop
	351	GRI.I FHMOFVACSCOVERIE BUIDDERDORDED CONTROL
	439	GRLIEHMQEYACSCDVEEIFRKVRFSFEQPDGRAQMPTT.ERVT.L .:. : . :: : : : : ::
	433	QAVCEVFSTHAKDMDVVELLTEVNKKVACGFQTSQGSNILKQMPEMTSRL
		V K F
	395	TRCFYLFPGH* 404 : :: :
	489	LKKFYFWPEARNSAV 503

FIGURE 6(b)

251	EHFEOMPTRNGTKADKDNLTNLFRCMGYTVICKDNLTGRGMLLTIRDFAK	300
1		9
301	HESHGDSAILVILSHGEENVIIGVDDIPISTHEIYDLLNAANAPRLANKP	350
10	::: : : ::: :: :: :: :: :: :: :: :: ::	
351	KIVFVQACRGERRDNGFPVLDSVDGVPAFLRRGWDNRDGPLFNFLGCVRP	400
38	PLLLYETDRGVDQQDGKNHTQSPGCEESDAGKEELM	73
401	QVQQVWRKKPSQADILIRYATTAQYVSWRNSARGSWFIQAVCEVFSTHAK	450
74	KMRLPTRSDMICGYACLKGNAAMRNTKRGSWYIEALTQVFSEREC	118
451	DMDVVELLTEVNKKVACGFQTSQGSNILKQMPEMTSRLLKKFYFWPEA	498
119	DMDVVELLIEVNR.: KVACGIQISQOSHIEV II.	168
499	RNSAV 503	
160	 ppm* 172	

FIGURE 7

--voovs--..vheffdulyanabilyankeki vevdacrg

Consensus

Alignment of N-terminal regions of ced-3/ICE- related proteins

_			m	
ed-) marodrwsllernilefsski gadlildvijakovlasdagdvinscrterdneke i vkavorrodeafdafydalrotorindladvimfi.srfnpv marodrrsllernimmfsshlkvde i Levliakovlasdagdminscotvrekrre i vkavorpodafydalrstoheglaevleplarsvdsnav 100	MADKIL	ed-3 EFECPMSPSSHRRSRALSPPGYASPTRVHRDSISSVSSFTSTYQDVYSRARBSSRSSRPLQBSDRHNYSBPVNAFPSQPSBANSFTGCASLGYSSSRN 198 EFECPMSPASHRRSRALSPAGYTSPTRVHRDSVSSVSSFTSYQD-IYSRARBRSRS-RALHBSDRHNYSBPVNAFPSQPSBANSSFTGCSSLGYSSSRN 198 ed-3 KLANITAMDKARDLCDHVSKKGPQASQIFITYICNEDCYLAGILELQSAPSAETFVATEDSKGCHFSFSETKEEQNKED-G KRENATVMDKTRALIDSVIPKGAQACQICITYICEEDSYLAGILGLSADQTSGNYLWMQDSQCVLSSFPAPQAVQDNPAMP KRENATVMDKTRALIDSVIPKGAQACQICITYICEEDSYL	RSFSKTSAGSQYIFHEEDMNYVDAPTIHRVPDEKTBYRNFSSPRIGGII WENFEQHPTRIGHKADKONLTNIFRCHGYNVLCKINLTDREMLSTIRSF 298 RSFSKASGPTQYIFHEEDMNYVDAPTISRVFDEKTMYRNFSSPRIMGII WENFEQMPTRIGHKADKONLTNLFRCHGYNVLCKINLTGRCMLLTIRGF 298 RSYSKASAHSQYIFHEEDMNYVDAPTHRVPDEKTMYRNFSTPRIGHII WENFEQMPTRIGHKADKONISNLFRCHGYNVCKINLTGRAML-TIRGF TFPGLTGTLKFCPLEKAQKLWKENP9EIYPIMUNT-TRTR-LALII WENFFDHLSFRIGHQNDAREMKLLEDLGYNVKVKFNLTNLEMLKEVKEF TSSGSEGNVKLCSLEEAQRIWKQKSAEIYPIMUKS-SRIR-LALII WENFEDSIPRNIGHENTHLLQNLGYNVKVLTNSCMITELEAF RS.SK.BQYIFHEEDMN.VDAPTI.RVFDEKTMYRNFS.FRSICLII WENCOMPTRIGHK.DKON.TNLFRCHGYNVCKLUTGR.ML.TIR.E	ed-3 GRNDMH-CDSKILVILSHGEENVINGVDDVSVNVHEI VDLINAMAPALANKPKIVFVQACRG 360 ÄXHESHCDSAILVILSHGEENVINGVDDVSISTHEI VDLINAMAPALANKPKIVFVQACRG 360 ÄXNETHCDSAILVILSHGEENVINGVDDVSVNVHEI VDLINAMAPALANKPKIVFVQACRG AACPEHKTSDSIFLVFYSHGIQEGI ISTTYSNEVSDILKVDTI FOMMITIKOPILKKPKVIIIQACRG FIGORETICKKHSEQVPDILQLMAI FNMLNTKNOPILKKPKVIIIQACRG FIGORETICKKHSEQVPDILQLMAI FNMLNTKNOPILKKPKVIIIQACRG FIGORETICKKHSEQVPDILQLMAI FNMLNTKNOPILKKPKVIIIQACRG FIGORETICKKHSEQVPDILQLMAI FNMLNTKNOPILKKPKVIIIQACRG
C. briggsae ced-1 ced-3 protein	C. vulgaris ced-3 Mouse ICE.gw Kuman ICE.GW Consensus	C. briggsae ced-3 ced-3 protein C. vulgaris ced-3 Mouse ICE.gw Auman ICE.GW	C. briggsae ced-3 ced-3 protein C. vulgaris ced-3 Mouse ICE.gw Human ICE.GW	C. briggsae ced-3 ced-3 protein C. vulgaris ced-3 Mouse ICE.gw Human ICE.GW

SI	3 8 · 8 ·	432	480	
ted proteir	RRGWDNRRGWDNRRGWDK LYETDRGVDQ	PDNVŠMRHPT PDNVSWRHPV AQYVSWRNSA AQYVSWRNSA KGNAAMRNTK	QPDGRAQMPT QPEFRLQMPT TSQGSNILKQ TSQGSNILKQ TSQGANILKQ PGTEFHRCKE	FIGURE 8(b)
D-2 - rela		KDFIAFCSST PDNVSWRHPV KDFIAFCSST PDNVSWRHPV ADMLIAYATT AQYVSWRNSA ADMLIAYATT AQYVSWRNSA ADMICGYACT KGNAAWRNTK DYA.TVSWRN	-RFSFE -RKVACGFQ KKVACGFQ KKVACGFQ ALIKEREGYA	FIGU
3-3/ICE/NE	FrdbygLkdbyrgVLdbydgVLdbydgVLdbydg- EVLLdplgTSLdby	TTEEFED DAIKKA-HIE GCVRPQVQ QVWRKK-PSQ GCVRPQVQ QVWRKK-PSQ GCVRPQAQ QVWRKK-PSQ GCVSDAGKE ELMKMRLPTR GCKP	DVEEIFRKV- DLEDIFRKV- DVVELLTEVN DVVELLTEVN IIVADMLVKVN DV.E.LKN	 RNSAV -NSAV 503 RSSAV -YPPT
ons of ced	DSPGVVWFKDSVG ERQCVVLLKDSVR ERRDNGFPVLDSVDG- ERRDVGFPVLDSVDG- ERRDVGFPVLDSVDG- MLTVQVYRTS QKCSSSKHVV EVLLDPLGTS E LDSV		EHMQEYALSC KHMKEYAWSC EVFSLHAKDM EVFSLHAKDM QVFSERA;DM EVFSLHAKDM	CFY LFP3H RFY LFP3H KFY FWP3DRG KFY FWP3AR- CLY JFP3
erminal regi	DSPGVVW EKQGVVL ERRDNGFP ERRDVGFP ERRDVGFP	SGNLSLP SEEDFL RDG-PLFNFL RDG-PLFNFL GDG-PNFL QDGKNHTQSP	MGS/FIGRLI RGS/FIGSLI RGS/VFIGAVC RGS/VFIGAVC RGS/VFIGAVC RGS/VFIGAVC RGS/VFIGAVC	T_ERVT-LITK A_DRVT-LITK MPELTSRLLK MPELTSRLLK MPELTSRLLK MSEYCSTLCQ MSEYCSTLCQ
Alignment of the C-terminal regions of ced-3/ICE/NEDD-2 - related proteins	ICE C-terminus Mouse ICE C-ter C.briggsae C-ter ced-3 Cterminus C. vulgaris C-terminus nedd-2 protein.gw Consensus	ICE C-terminus Mouse ICE C-ter C.briggsae C-ter ced-3 Cterminus C. vulgaris C-terminus nedd-2 protein.gw	ICE C-terminus Mouse ICE C-ter C.briggsae C-ter ced-3 Cterminus C. vulgaris C-terminus nedd-2 protein.gw	ICE C-terminus Mouse ICE C-ter C.briggsae C-ter ced-3 Cterminus C. vulgaris C-terminus nedd-2 protein.gw

Lines		
1 2 3	01	MMRQDRRSLLERNIMMESSHLKVDEILEVLIAKQVLNSDNGDMINSCGTV 50WLEK.QA.LDVR.E TVSISLJRM
1 2 3	51	REKRREIVKAVQRPGDVAFDAFYDALRSTGHEGLAEVLEPLARSVDSNAV 100 .DNEKREDNDDMS.PP.
1 2 3		EFECPMSPASHRRSRALSPAGYTSPTRVHRDSVSSVSSFTS_YQDIYSRA 149 PMSP.AITV S
1 2 3	150	RSRSR_SRAIHSSDRHNYSSPPVNAFPSQPSSANSSFTGCSSLGYSSSRN 198ssp.QM.AA_TSA
1 2 3	199	RSFSKASGPTQYIFHEEDMNFVDAPTISRVFDEKTMYRNFSSPRGMCLI 247T.AQSYH
1 2 3	248	INNEHFECMPTRNGTKADKDNLTNLFRCMGYTVICKDNLTGRGMLLTIRD 297
1 2 3	298	FAKHESHGDSAILVILSHGEENVIIGVDDIPISTHEIYDLLNAANAFRLA 347 .GRNDMVSVNV
1 2 3	348	NKPKIVEVCACRGERRONGEPVIDS DGVPAFLRRGWONROGPLENFLGC 397
1 2 3	398	S VRPQVQQVWRKKPSQADILIRYATTAQYVSWRNSARGSWFIQAVCEVFST 447
1 2 3	44	8 HAKDMDVVELLTEVNKKVACGFQTSQGSNILKQMPEMTSRLLKKFYFWPE 497
1 2 3	49	8 ARN SAV 503 DRG DRS

FIGURE 9

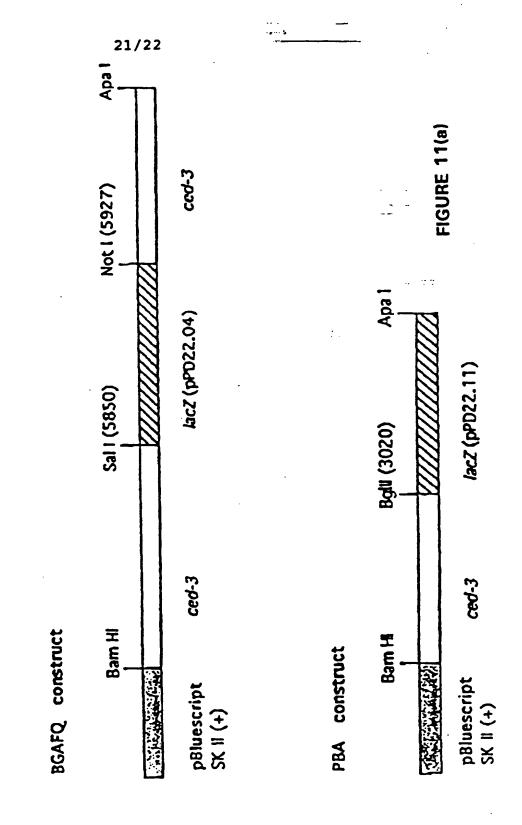
Interleukin-1 β convertase cDNA sequence

1	AAAAGGAGAG	AAAAGCCATG	GCCGACAAGG	TCCTGAAGGA	GAAGAGAAA
51	CTGTTTATCC	GTTCCATGGG	TGAAGGTACA	ATAAATGGCT	TACTGGATGA
101	ATTATTACAG	ACAAGGGTGC	TGAACAAGGA	AGAGATGGAG	AAAGTAAAAC
151	GTGAAAATGC	TACAGTTATG	GATAAGACCC	GAGCTTTGAT	TGACTCCGTT
201	ATTCCGAAAG	GGGCACAGGC	ATGCCAAATT	TGCATCACAT	ACATTTGTGA
251	AGAAGACAGT	TACCTGGCAG	GGACGCTGGG	ACTCTCAGCA	GATCAAACAT
301	CTGGAAATTA	CCTTAATATG	CAAGACTCTC	AAGGAGTACT	TTCTTCCTTT
351	CCAGCTCCTC	AGGCAGTGCA	GGACAACCCA	GCTATGCCCA	CATCCTCAGG
401	CTCAGAAGGG	AATGTCAAGC	TTTGCTCCCT	AGAAGAAGCT	CAAAGGATAT
451	GGAAACAAAA	GTCGGCAGAG	ATTTATCCAA	TAATGGACAA	GTCAAGCCGC
501	ACACGTCTTG	CTCTCATTAT	CTGCAATGAA	GAATTTGACA	GTATTCCTAG
551	AAGAACTGGA	GCTGAGGTTG	ACATCACAGG	CATGACAATG	CTGCTACAAA
601	ATCTGGGGTA	CAGCGTAGAT	GTGAAAAAAA	ATCTCACTGC	TTCGGACATG
651	ACTACAGAGC	TGGAGGCATT	TGCACACCGC	CCAGAGCACA	AGACCTCTGA
701	CAGCACGTTC	CTGGTGTTCA	TGTCTCATGG	TATTCGGGAA	GGCATTTGTG
751	GGAAGAAACA	CTCTGAGCAA	GTCCCAGATA	TACTACAACT	CAATGCAATC
801	TTTAACATGT	TGAATACCAA	GAACTGCCCA	AGTTTGAAGG	ACAAACCGAA
851	GGTGATCATC	ATCCAGGCCT	GCCGTGGTGA	CAGCCCTGGT	GTGGTGTGGT
901	TTAAAGATTC	AGTAGGAGTT	TCTGGAAACC	TATCTTTACC	AACTACAGAA
951	GAGTTTGAGG	ATGATGCTAT	TAAGAAAGCC	CACATAGAGA	AGGATTTTAT
001	CGCTTTCTGC	TCTTCCACAC	CAGATAATGT	TTCTTGGAGA	CATCCCACAA
051	TGGGCTCTGT	TTTTATTGGA	AGACTCATTG	AACATATGCA	AGAATATGCC
101	TGTTCCTGTG				
151	GCAGCCAGAT				
201	CAAGATGTTT				
251	ATGTCTGCGG				
301	TTATGTCTGC	TGAATAATAA	ACTTTTTTG	AAATAATAAA	TCTGGTAGAA
351	ABBTCBBBB	*****			

FIGURE 10

Constructs that Prevent Programmed Cell Death in C. elegans

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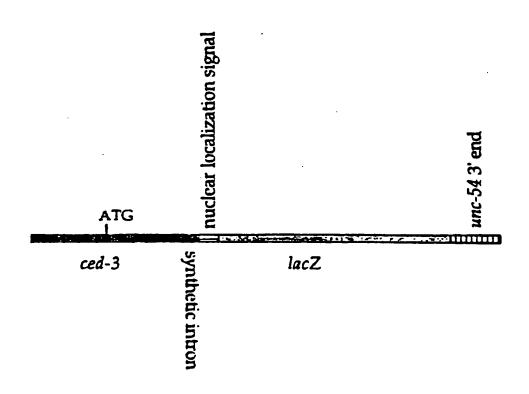


FIGURE 11(b)

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 93/05705

	CT MATTER (if several classification syr		
·	Classification (IPC) or to both National Cla		101101 170
Int.Cl. 5 C12N15/57 A61K37/02			A61K31/70 G01N33/68
II. FIELDS SEARCHED			
	Minimum Documen	tation Searched?	
Classification System	C	lassification Symbols	
Int.Cl. 5	C12N ; A61K ;	C12Q ; G01N	
	Documentation Searched other the to the Extent that such Documents as		
III. DOCUMENTS CONSIDERE	D TO BE DEI EVANT 9		
	ocument, 11 with indication, where appropriat	of the palgraph personne 12	Relevant to Claim No.13
Category Citation of Do	ocument, with indication, where appropriate	ie, of the relevant passages	RESEVANT TO CLAIM INC.
vol. 50, page 443 J. YUAN ced-3 ar program	'Genetic and molecular nd ced-4: Two genes that ned cell death' e 4431-B, left column, p	studies of control	1-13, 16-21, 28-45, 101-122
THE UNIV	e 6, line 1 - page 7, li		1-45, 101-122
"E" earlier document but publifiling date "L" document which may throw which is cited to establish citation or other special re "O" document referring to an other means	neral state of the art which is not ular relevance ished on or after the international w doubts on priority claim(s) or the publication date of another ason (as specified) oral disciosure, use, exhibition or to the international filing date but	"T" later document published after the int or priority date and not in conflict wincited to understand the principle or the invention "X" document of particular relevance; the cannot be considered novel or cannot involve an inventive step "Y" document of particular relevance; the cannot be considered to involve an involve and involved with one or moments, such combined with one or moments, such combination being obvious in the art. "&" document member of the same patent	th the application but leave underlying the claimed invention be considered to claimed invention ventive step when the tre other such docu- is to a person skilled
Date of the Actual Completion of t	the International Search	Date of Mailing of this International S	iearch Report
05 OCTOR		1 5 -10- 199	•
International Searching Authority EUROPEA	AN PATENT OFFICE	Signature of Authorized Officer HORNIG H.	

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Category °	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No.
X	WO,A,9 115 577 (IMMUNEX CORPORATION) 17 October 1991	54-56
Y	see page 3, line 25 - page 6, line 10; claims 1-30	14,15, 22-27, 101-122
	MATURE	54-57
X	NATURE vol. 356, 30 April 1992, MACMILLAN JOURNALS LTD., LONDON,UK;	54-57
	pages 768 - 774 N.A. THORNBERRY ET AL. 'A novel heterodimeric cystein protease is required	
	for interleukin-1-beta processing in monocytes'	
γ	cited in the application see page 770, right column, paragraph 2	14,15,
1	see page 770, right column, paragraph 2	22-27,
		101-122
	see page 773, right column, paragraph 2	
x	CELL	54,58
^	vol. 69, no. 4, 15 May 1992, CELL PRESS,	31,00
	CAMBRIDGE, MA, US;	
	pages 597 - 604	
	C.A. RAY ET AL. 'Viral inhibition of inflammation: Cowpox virus encodes an	
	inhibitor of the interleukin-1-beta	
	converting enzyme'	
	cited in the application see page 599, right column, paragraph 2 -	
	page 600, left column, paragraph 1	
A	ANNUAL REVIEW OF CELL BIOLOGY	-
	vol. 7, 1991, ANNUAL REVIEW INC., PALO	
	ALTO,CA,US; pages 663 - 698	
	R.E. ELLIS ET AL. 'Mechanisms and	
	functions of cell death'	
	cited in the application	
	the whole document	
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	vol. 185, no. 3, 30 June 1992, ACADEMIC	
	PRESS, N.Y., US;	
	pages 1155 - 1161 S. KUMAR ET AL. 'Identification of a set	
	of genes with developmentally	
	down-regulated expression in the mouse	
	brain'	
	cited in the application see page 1161,	
	paragraph 1; table 1	
	-/	
	•	

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)					
Category o	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No			
, х	DATABASE WPI	128			
	Week 9318, Derwent Publications Ltd., London, GB; AN 93-144400				
	& CA,A,2 076 159 (MERCK & CO. INC.) 17 February 1993 see abstract	·			
,х	EP,A,O 533 350 (MERCK & CO. INC.) 24 March 1993	16,15, 24, 30,42, 60,128			
	abstract	00,120			
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	<pre>page S47 S. LEDOUX AND J. YUAN 'Isolation of nematode homologs of the C. elegans cell</pre>				
	death gene ced-3' Third international conference on Alzheimer's disease and related disorders, Abano terme, Italy, July 12-17, 1992; Abstract po 183:				
	Abstract no. 183; abstract				
	•				
	•				
}					

INTERNATIONAL SEARCH REPORT

national application No.

PCT/US 93/05705

Box I	Observations where certain claims were found unscarchable (Continuation of item 1 of first sheet)					
This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:						
ı. [X]	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely: Remark . Although claims 65,66,71,72 (as far as they concern in vivo treatment of human or animals) and claims 51,63,77,78 are directed to a method of treatment of the human/animal body the search has been carried out and based on the alleged effects of the compound/composition.					
2.	Clams Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:					
3.	Claims Nos because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).					
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)					
This Int	ernational Searching Authority found multiple inventions in this international application, as follows:					
1.	As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.					
2. [As all searchable claims could be searches without effort justifying an additional fee, this Authority did not invite payment of any additional fee.					
3. []	As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:					
4. []	No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:					
Remark	on Protest The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.					

ANNEX TO THE INTERNATIONAL SEARCH REPORT ON INTERNATIONAL PATENT APPLICATION NO.

US 9305705 SA 76331

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report. The members are as contained in the European Patent Office EDP file on

The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

05/1 05/10/9

Patent document cited in search report				Publication date
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WO-A-9115577	17-10-91	AU-A-	7775991	30-10-91
EP-A-0533350	24-03-93	None		

For more details about this annex : see Official Journal of the European Patent Office, No. 12/82